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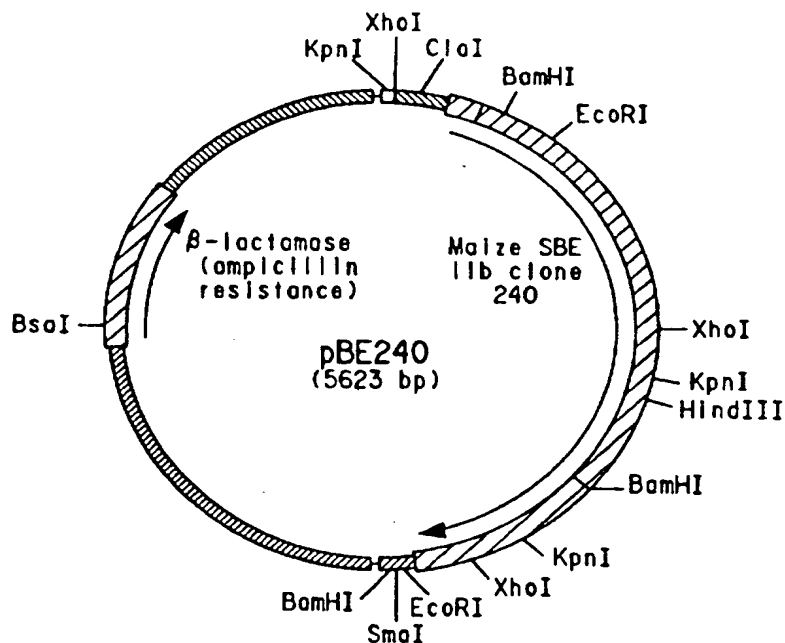
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(57) Abstract

The instant invention discloses utilization of a cDNA clone to construct sense and antisense genes for inhibition of starch branching enzyme enzymatic activity in corn. More specifically, this invention concerns a method of controlling the starch fine structure of starch derived from the grain of corn comprising: (1) preparing a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or a fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination, and (2) transforming corn with said chimeric gene, wherein expression of said chimeric gene results in alteration of the fine structure of starch derived from the grain of said transformed corn compared to the fine structure of starch derived from corn not possessing said chimeric gene.



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TITLENOVEL STARCHES VIA MODIFICATION OF EXPRESSION OF STARCH
BIOSYNTHETIC ENZYME GENESBACKGROUND OF THE INVENTION5 Characteristics and Commercial Utility of Starch

Starch is a mixture of two polysaccharides, amylose and amylopectin. Amylose is an unbranched chain of up to several thousand α -D-glucopyranose units linked by α -1,4 glycosidic bonds. Amylopectin is a highly branched molecule made up of up to 50,000 α -D-glucopyranose residues linked by α -1,4 and α -1,6 glycosidic bonds. Approximately
10 5% of the glycosidic linkages in amylopectin are α -1,6 bonds, which leads to the branched structure of the polymer.

Amylose and amylopectin molecules are organized into granules that are stored in plastids. The starch granules produced by most plants are 15-30% amylose and 70-85% amylopectin. The ratio of amylose to amylopectin and the degree of branching of
15 amylopectin affects the physical and functional properties of the starch. Functional properties, such as viscosity and stability of a gelatinized starch determine the usefulness and hence the value of starches in food and industrial applications. Where a specific functional property is needed, starches obtained from various crops such as corn, rice, or potatoes may meet the functionality requirements. If a starch does not meet a required
20 functional property, if for example it must have stable viscosity under high temperatures and acidic conditions, the functionality can sometimes be achieved by chemically modifying the starch. Various types and degrees of chemical modification are used in the starch industry, and the labeling and use of chemically modified starches must meet government regulations.

25 Within the starch bearing organs of plants, the proportion of amylose to amylopectin and the degree of branching of amylopectin are under genetic control. For example, plants homozygous recessive for the *waxy* (*wx*) gene lack a granule-bound starch synthase enzyme and produce nearly 100% amylopectin. Plants homozygous recessive for the *amylose extender* (*ae*) gene can produce starch granules that are up to
30 90% amylose (see U. S. Pat. No. 5,300,145). The *dull* gene has been shown to influence the levels of activity of a starch synthase and a starch branching enzyme.

Most cereal crops are handled as commodities, and many of the industrial and animal feed requirements for these crops can be met by common varieties which are widely grown and produced in volume. However, there exists at present a growing
35 market for crops with special end-use properties which are not met by grain of standard composition. Most commonly, specialty corn is differentiated from "normal" corn, also known as field corn, by altered endosperm properties, such as an overall change in the ratio of amylose to amylopectin as in waxy or high amylose corn, an increased

accumulation of sugars as in sweet corn, or an alteration in the degree of endosperm hardness as in food grade corn or popcorn; Glover, D. V. and E. T. Mertz, (1987) in Corn: Nutritional Quality of Cereal Grains; Genetic and Agronomic Improvement, R. A. Olson and K. J. Frey, eds. American Society of Agronomy, Madison Wisconsin, pp. 183-336. Rooney, L. W. and S. O. Serna-Saldivar, (1987) Food Uses of Whole Corn and Dry-milled Fractions, in Corn: Chemistry and Technology, S. A. Watson and P. E. Ramstead, eds. American Association of Cereal Chemists, Inc., St. Paul, Minnesota, pp. 399-429. The current invention offers the buyers of specialty corn a source of starch having properties distinct from waxy starch and offers farmers the opportunity to grow a higher value-added crop than normal or waxy corn.

Purified starch is obtained from plants by a milling process. Corn starch is extracted from kernels through the use of a wet milling process. Wet milling is a multi-step process involving steeping and grinding of the kernels and separation of the starch, protein, oil and fiber fractions. A review of the corn wet milling process is given by S. R. Eckhoff in the Proceedings of the Fourth Corn Utilization Conference, June 24-26, 1992, St. Louis, MO., printed by the National Corn Growers Association, CIBA-GEIGY Seed Division and the United States Department of Agriculture. Starch is used in numerous food and industrial applications and is the major source of carbohydrates in the human diet. Typically, starch is mixed with water and cooked to form a thickened gel. Three important properties of a starch are the temperature at which it cooks, the viscosity the gel reaches, and the stability of the gel viscosity over time. The physical properties of unmodified starch during heating and cooling limit its usefulness in many applications. As a result, considerable effort and cost is needed to chemically modify starch in order to overcome these limitations of starch and to expand the usefulness of starch in industrial applications.

Some limitations of unmodified starches and properties of modified starches are given in Modified Starches: Properties and Uses, O. B. Wurzburg, ed., (1986) CRC Press Inc., Boca Raton, FL. Unmodified starches have very limited use in food products because the granules swell and rupture easily, thus forming weak bodied, undesirable gels. Chemical modifications are used to stabilize starch granules thereby making the starch suitable for thousands of food and industrial applications including baby foods, powdered coffee creamer, surgical dusting powders, paper and yarn sizings and adhesives. Common chemical modifications include cross linking in which chemical bonds are introduced to act as stabilizing bridges between starch molecules, and substitution in which substituent groups such as hydroxyethyl, hydroxypropyl or acetyl groups are introduced into starch molecules.

The use of chemically modified starches in the United States is regulated by the Food and Drug Administration (FDA). "Food starch-modified" starches may be used in food but must meet specified treatment limits, and "industrial starch-modified" starches may be used in items such as containers that come in contact with food and must also meet specified treatment requirements; Code of Federal Regulations, Title 21, Chapter 1, Part 172, Food Additives Permitted in Food for Human Consumption, Section 172.892, Food Starch-Modified, U. S. Government Printing Office, Washington, D. C. 1981; (a) Part 178, Indirect Food Additives, Sect. 178.3520, Industrial Starch-Modified. These regulations limit the degree of chemical modification by defining the maximum amount of chemical reagent that can be used in the modification steps. The levels of by-products in starch resulting from the modification process are also regulated. For example, propylene chlorohydrin residues in hydroxypropyl starch are of special concern; Tuschhoff, J. V., (1986) Hydroxypropylated Starches, In Modified Starches: Properties and Uses, O. B. Wurzburg, ed., CRC Press, Boca Raton, FL, pp. 55-57.

15 Alteration of Starch Fine Structure Through Molecular Genetic Manipulation of Starch-Bearing Plants

Differences in the degree of starch branching or polymerization are known to result in a change in the physiochemical properties of starch. It has been suggested that starches, tailor-made for specific applications, may be generated by alteration of the branch chain distribution of the amylopectin molecule, the relative proportion of amylose to amylopectin or the degree of polymerization of amylose. However, achieving phenotypic alteration of starch composition has been problematic; while key enzymes in starch biosynthesis have been identified, their exact roles remain uncertain. Thus, correlation of activities of particular enzymes with particular molecular characteristics of starch structure and, in turn, with starch function in food and industrial products has been difficult. Although desirable functional properties that an ideal starch might need can be envisioned, there is only a vague understanding of what the molecular structure of the starch should be to achieve this and little understanding of how particular starch biosynthetic enzymes specifically affect those parameters. For example, the role of individual enzymes in determining the branching patterns and length of branches is as yet unclear and is compounded by the lack of understanding of how branching enzymes and starch synthases interact.

WO 94/09144 discusses the generation of plants with improved ability to synthesize starch at elevated temperatures. This publication proposes that the limiting factor in grain filling at elevated temperature is the lability of certain starch biosynthetic enzymes, particularly starch synthase (SS) and starch branching enzyme (SBE). The

introduction of genes encoding enzymes that have a higher optimum temperature for activity or that have a higher tolerance to heating into plants may afford an increase in the amount of starch deposited in the corn kernel. Moreover, it is claimed that this strategy may be used to generate starch of altered fine structure as a result of the introduction of donor genes whose expression may alter the balance of the different starch biosynthetic enzymes. Suggested donor genes include those that encode enzymes that display improved kinetic or allosteric properties relative to the endogenous enzyme or an extra copy of the endogenous gene that would compensate for losses in enzyme activity incurred due to heat lability. As a means to alter starch structure, WO 94/09144 also suggests the use of sense and antisense genes to alter the natural ratios of the different starch synthase and branching enzymes in the recipient plant. This publication discloses the effect of temperature on catalytic activity and enzyme stability for certain starch biosynthetic enzymes, however, no data are presented to substantiate the proposed molecular strategies.

The results of attempts to inhibit SBE expression in potato using an antisense approach were recently reported by Virgin et al. at the 4th International Congress of Plant Molecular Biology (June, 1994) and by Christensen et al. and Kossman et al. at the Plant Polysaccharide Symposium (July, 1994). In all cases, although SBE activity was almost completely abolished, the amylose-to-amylopectin ratio remained unaltered. Both Virgin et al. and Kossman et al. reported no change in amylopectin structure. However, Christensen et al. did report a change in the distribution of branch chains on the amylopectin molecule with an increase in the number of long branches.

The results in potato are unexpected, since only a single starch branching enzyme has been purified and only a single gene has been detected on Southern blots of potato genomic DNA, even under conditions of low stringency (Kooshnoodi, J. et al. (1993) *FEBS Letters* 332:132-138; Kossman, J. et al. (1991) *Mol. Gen. Genet.* 230:39-44). Thus, antisense suppression of the single starch branching enzyme gene in potato, resulting in significant reduction of enzyme levels and a concomitant decrease in starch branching enzyme activity, was expected to result in a measurable, reproducible change in starch composition and starch fine structure. The contrary and inconsistent results reported in the literature suggest that other starch branching enzyme genes that share little homology with the identified gene may also play a role in determining amylopectin structure in potato. Alternatively, branching enzyme activity in potato may be encoded by a single gene, but the protein may be present in such large excess that amylopectin quantities or structure are not affected even when greater than 90% of the enzyme activity is inhibited.

Alteration of starch fine structure in corn is complicated by the fact that three SBE isoforms have been identified. In corn endosperm, the three isoforms that demonstrate starch branching enzyme activity are SBEI, SBEIIa and SBEIIb. In the *amylose extender (ae)* mutant, SBEIIb activity has been found to be deficient while in the *dull (du)* mutant, decreased levels of SBEIIa are observed (Boyer, C. D. and Preiss, J. (1981) *Plant Physiol.* 67:1141-1145). Studies of the catalytic properties of the corn starch branching enzymes indicate that the isoforms differ in substrate preference and in the length of glucan chain that is transferred. SBEI activity is higher when amylose serves as the substrate, and longer chains are preferentially transferred. The SBEII isoforms display higher activity with more highly branched substrates such as amylopectin. These enzymes preferentially transfer shorter glucan chains (Guan et al. (1993) *Plant Physiol.* 102:1269-1273; Takeda et al. (1993) *Carbohydrate Res.* 240:253-263).

A corn SBEI cDNA has been cloned and sequenced (Baba et al. (1991) *Biochem. Biophys. Res. Commun.* 181:87-94; Fisher et al. (1995) *Plant Physiol.* 108:1313-1314). In addition, a corn SBEII cDNA clone has been isolated and the nucleotide sequence of the clone has been published (Fisher et al. (1993) *Plant Physiol.* 102:1045-1046). This cDNA clone maps to the *ae* locus, confirming that this locus encodes the structural gene for corn SBEIIb (Stinard et al. (1993) *Plant Cell* 5:1553-1566).

Starch isolated from the *ae* mutant is known to differ in structure from that isolated from dent corn (Baba et al. (1984) *Agric. Biol. Chem.* 48:1763-1775). The effect of the *ae* allele on starch properties has been investigated (Yamada et al. (1978) *Starke* 30:145-148). Increasing doses of *ae* in a *waxy (wx)* background produce an increase in the gelatinization temperature so that for the homozygous mutant, incomplete cooking of the starch is observed, even at 95°C. These authors indicate that the increase in viscosity associated with *ae wx* starch is highly desirable and suggest a "target" starch with properties intermediate between *wx* and *ae wx*. While mutations which influence the levels of corn SBEIIa and SBEIIb are available, mutations in the SBEI structural gene have yet to be identified. The lack of SBEI mutants may indicate that the absence of this branching enzyme isoform is lethal to the plant. Alternatively, a SBEI null mutation may give rise to no observable change in seed phenotype or one that is not readily distinguished from existing starch mutants.

Molecular genetic solutions to the generation of starches from corn with altered fine structures have a decided advantage over more traditional plant breeding approaches. Changes to starch fine structure can be produced by specifically inhibiting expression of one or more of the SBE isoforms by antisense inhibition or cosuppression.

An antisense or cosuppression construct would act as a dominant negative regulator of gene activity. While conventional mutations can yield negative regulation of gene activity this effects are most likely recessive. The dominant negative regulation available with a transgenic approach may be advantageous from a breeding perspective.

- 5 Additionally the ability to restrict the expression of the altered starch phenotype to the reproductive tissues of the plant by the use of specific promoters may confer agronomic advantages relative to conventional mutations which will have an effect in all tissues in which the mutant gene is ordinarily expressed. Finally, the variable levels of antisense inhibition or cosuppression that arise from chromosomal position effects could produce a
10 wider range of starch phenotypes than those that result from dosage effects of a mutant allele in corn endosperm.

The complex organization of starch branching enzymes in corn endosperm and the results reported in potato render attempts to manipulate starch fine structure by inhibition of gene expression of one of the known corn isoforms unpredictable.

- 15 Reported scientific evidence indicates that inhibition of expression of a single starch branching enzyme gene and a measurable reduction of starch branching enzyme activity is not predictive of a corresponding change in starch fine structure. Moreover, antisense technology is inherently uncertain in that it is not obvious or predictable whether the entire gene or whether specific fragments and which fragments of a gene will be most
20 effective in mediating strong antisense inhibition. Some results do indicate that strong expression of the antisense gene is required, however, good expression of the antisense transcript does not necessarily correlate with the observation of and the strength of the antisense phenotype (Bourque, J. (1995) *Plant Sci.* 105:125-149). Although several theories have been advanced to explain the phenomenon of cosuppression (Flavell, R. B.
25 (1994) *Proc. Natl. Acad. Sci. (USA)* 91:3490-3496), it has become apparent that no single mechanism appears sufficient to describe all of the observed results. To date, cosuppression effects have been reported in tobacco, canola, soybean, tomato and Arabidopsis, all of which are dicot plants. No data have been reported that indicates that this phenomenon is operable in monocots.

- 30 Notwithstanding the ability to inhibit the expression of SBE genes in corn, a resulting change in starch phenotype remains unpredictable. Although the enzymatic steps are known, the molecular details of starch biosynthesis are not well understood. It is not clear whether the three SBE isoforms contribute equally throughout starch biosynthesis or whether each isoform plays a distinct role in assembling the amylopectin
35 molecule at discrete steps along an obligatory pathway. In consideration of the possible interplay between the starch branching enzymes and the multiple starch synthases that

function in glucan chain elongation, it is impossible to make predictions concerning starch structure based upon the catalytic properties of each isoform.

SUMMARY OF THE INVENTION

The instant invention discloses utilization of a cDNA clone to construct sense and antisense genes for inhibition of starch branching enzyme enzymatic activity in corn grain or endosperm. More specifically, this invention concerns a method of controlling the branch chain distribution of the amylopectin, the relative proportion of amylose to amylopectin and the degree of polymerization of amylose components of starch in corn comprising: (1) preparing a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or a fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment that encodes a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination, and (2) transforming corn with said chimeric gene, wherein expression of said chimeric gene results in alteration of the branch chain distribution of the amylopectin molecular component of starch derived from the grain of said transformed corn compared to the branch chain distribution of the amylopectin molecular component of starch derived from corn not possessing said chimeric gene. This invention also concerns corn varieties prepared by transformation using said method, starch isolated from the grain of a corn variety prepared using said method, and a method of preparing a thickened foodstuff comprising combining a foodstuff, water, and an effective amount of a starch isolated from the grain of a corn variety prepared using the said method, and cooking the resulting composition as necessary to produce said thickened foodstuff.

BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and the sequence descriptions which form a part of this application.

Figure 1 presents a restriction map of plasmid pBE240 that contains a cDNA insert comprising 78 bp of 5' untranslated DNA, a 2397 bp open reading frame encoding the corn SBEIIb coding region and 190 bp of 3' untranslated DNA.

Figure 2 is a restriction map of plasmid pBE44 comprising a 414 bp 3' fragment of the insert of pBE240 in antisense orientation with respect to the corn 27 kd zein promoter.

Figure 3 is a restriction map of plasmid pML103, used as an intermediate cloning vehicle in construction of chimeric genes of the instant invention.

Figure 4 is a restriction map of plasmid p35/Ac encoding, *inter alia*, phosphinothricin acetyl transferase. Introduction of this plasmid into plant cells and tissues confers resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin on the transformed plant cells and tissues.

5 Figure 5 compares RVA profiles of starch from normal dent corn kernels, kernels homozygous for amylose extender (*ae*) and starch from kernels homozygous for the pBE44 construct. Viscosity, in stirring number units (SNU), and temperature (degrees Celsius) have been measured and plotted as a function of time (in minutes).

10 Figure 6 is a restriction map of plasmid pBE43 comprising a 507 bp 5' fragment of the insert of pBE240 in antisense orientation with respect to the corn 27 kd zein promoter.

Figure 7 is a restriction map of plasmid pBE45 comprising a 2165 bp near full length fragment the insert of pBE240 in antisense orientation with respect to the corn 27 kd zein promoter.

15 Figure 8 is a restriction map of plasmid pBE96 comprising a 2087 bp near full length fragment the insert of pBE240 in sense orientation with respect to the corn 27 kd zein promoter.

20 Figure 9 is a restriction map of plasmid pBE68 comprising a 373 bp fragment representing the 3' end of the corn SBEI cDNA insert in pBE65 (SEQ ID NO:13), joined in antisense orientation to the corn 27 kd zein promoter.

Figure 10 is a restriction map of plasmid pBE69 comprising a 570 bp fragment representing the 5' end of the corn SBEI cDNA insert in pBE65 (SEQ ID NO:16), joined in antisense orientation to the corn 27 kd zein promoter.

25 Figure 11 is a restriction map of plasmid pBE72 comprising a 2487 bp near full length fragment the insert of pBE65 in antisense sense orientation with respect to the corn 27 kd zein promoter.

Figure 12 is a restriction map of plasmid pBE108 comprising a hygromycin resistant variant of pBE72.

30 Figure 13 is a restriction map of plasmid pBE97 comprising a 1865 bp near full length fragment the insert the SBEI cDNA of pBE65 (SEQ ID NO:20) joined in sense orientation to the 27 kD zein promoter.

Figure 14 is a restriction map of plasmid pBE110 comprising a 2565 bp cDNA fragment encoding a full length SBEI joined in sense orientation with respect to the maize 10 kd zein promoter.

Figure 15 is a restriction map of plasmid pBE111 comprising a 1810 bp cDNA fragment encoding a truncated SBEI joined in sense orientation with respect to the maize 27 kd zein promoter.

Figure 16 compares RVA profiles of starch from waxy kernels, kernels
5 homozygous for amylose extender (*ae*) and waxy and from kernels containing the pBE44 construct plus waxy. Viscosity, in stirring number units (SNU), and temperature (degrees Celsius) have been measured and plotted as a function of time (in minutes).

SEQ ID NO:1 depicts the nucleotide sequence of the cDNA insert in plasmid pBE240 and the corresponding amino acid sequence of the entire corn SBEIb enzyme.

10 SEQ ID NO:2 depicts the nucleotide sequence of the the 414 bp insert of pBE44.

SEQ ID NOS:3 and 4 depict the PCR primers BE41 and BE42 used for preparation of the 414 bp insert of pBE44.

SEQ ID NO:5 depicts the nucleotide sequence of the the 507 bp insert of pBE43.

15 SEQ ID NOS:6 and 7 depict the PCR primers BE39 and BE40 used for preparation of the 507 bp insert of pBE43.

SEQ ID NO:8 depicts the nucleotide sequence of the the 2165 bp insert of pBE45.

SEQ ID NO:9 depicts the nucleotide sequence of the the 2087 bp insert of pBE96.

20 SEQ ID NOS:10 and 11 depict the PCR primers BE14 and BE15 used for preparation of the probe used to isolate the 2772 bp insert of pBE65. BE15 (SEQ ID NO:11) was also used for the preparation of the insert in plasmid pBE79.

SEQ ID NO:12 depicts the nucleotide sequence of the the 2772 bp insert of pBE65.

25 SEQ ID NO:13 depicts the nucleotide sequence of the the 373 bp insert of pBE68.

SEQ ID NOS:14 and 15 depict the PCR primers BE43 and BE52 used for preparation of the 373 bp insert of pBE68.

30 SEQ ID NO:16 depicts the nucleotide sequence of the the 571 bp insert of pBE69.

SEQ ID NOS:17 and 18 depict the PCR primers BE46 and BE50 used for preparation of the 571 bp insert of pBE69.

SEQ ID NO:19 depicts the nucleotide sequence of the the 2487 bp insert of pBE72.

35 SEQ ID NO:20 depicts the nucleotide sequence of the the 1865 bp insert of pBE97.

SEQ ID NO:21 depicts the PCR primer BE67 used for preparation of the 805 bp insert of pBE83.

SEQ ID NOS:22 and 23 depict the PCR primers BE101 and BB3 used for preparation of a pBE110.

5 SEQ ID NO:24 depicts the nucleotide sequence of the the 2565 bp insert of pBE110.

SEQ ID NO:25 depicts the nucleotide sequence of the the 1809 bp insert of pBE111.

10 The Sequence Descriptions contain the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IYUB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219(2):345-373 (1984) which are incorporated by reference herein.

DETAILED DESCRIPTION

15 In the context of this disclosure, a number of terms shall be utilized. As used herein, the term "starch" refers to a polysaccharide consisting of α -D-(1,4) glucan that may contain a variable proportion of α -D-(1,6) branches. As used herein, the term "starch fine structure" refers to the molecular structure of a starch polymer, the presence, abundance and distribution of α -D-(1,6) bonds and the presence, abundance and length
20 of both branched and unbranched α -D-(1,4) glucans in the polymer. Starch fine structure is described by amylopectin branch chain distribution, or by the relative proportion of amylose to amylopectin, or by the degree of polymerization of amylose. Alteration of any of these structural molecular components results in an altered starch fine structure. One, two or all three of these parameters may be altered independently of
25 one another. The term "degree of polymerization" refers to the number of α -D-glucopyranose units in a molecule or designated portion of a molecule such as a branch chain of amylopectin.

As used herein, the term "branch chain distribution" refers to the distribution of α -1,4-linked glucan chains which is detected following isoamylase digestion of
30 amylopectin and subsequent fractionation of the liberated branches by size exclusion chromatography. The branch chains may be classified according to their size and the number of crystalline regions (regions where many of the α -1,6-linkages (i.e., branch points) occur) which they span in the intact molecule. A chains are unbranched and span a single crystalline region. B1 chains also span a single crystalline region but are
35 branched. B2, B3 and B4+ chains are branched and span 2, 3 and 4 or more crystalline regions, respectively (Hizukuri (1986) *Carbohydrate Res.* 147:342-347). The length of

the repeating crystalline and amorphous units in the starch granule is quite regular with a repeat distance of 9 nm observed in starch from a wide variety of plant species (Jenkins (1993) Starch/Starke 45:417-420). Thus A and B1 chains are less than 9nm in length B2 and B3 chains are between 18 and 27 nm in length and B4+ chains are greater than
5 36 nm.

As used herein, the term "nucleic acid" refers to a large molecule which can be single-stranded or double-stranded, composed of monomers (nucleotides) containing a sugar, phosphate and either a purine or pyrimidine. A "nucleic acid fragment" is a fraction of a given nucleic acid molecule. In higher plants, deoxyribonucleic acid (DNA)
10 is the genetic material while ribonucleic acid (RNA) is involved in the transfer of the information in DNA into proteins. A "genome" is the entire body of genetic material contained in each cell of an organism. The term "nucleotide sequence" refers to a polymer of DNA or RNA which can be single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases capable of incorporation into DNA or
15 RNA polymers.

As used herein, "essentially similar" refers to DNA sequences that may involve base changes that do not cause a change in the encoded amino acid, or which involve base changes which may alter one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. It is therefore understood that
20 the invention encompasses more than the specific exemplary sequences. Modifications to the sequence, such as deletions, insertions, or substitutions in the sequence which produce silent changes that do not substantially affect the functional properties of the resulting protein molecule are also contemplated. For example, alteration in the gene sequence which reflect the degeneracy of the genetic code, or which results in the
25 production of a chemically equivalent amino acid at a given site, are contemplated; thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another hydrophobic amino acid residue such as glycine, valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged
30 residue for another, such as lysine for arginine, can also be expected to produce a biologically equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. In some cases, it may in fact be desirable to make mutants of the sequence in order to study the effect of alteration on the biological
35 activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products.

Moreover, the skilled artisan recognizes that "essentially similar" sequences encompassed by this invention can also be defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein.

5 "Gene" refers to a nucleic acid fragment that encodes all or a portion of a specific protein, and includes regulatory sequences preceding (5' non-coding) and following (3' non-coding) the coding region. "Native gene" refers to the gene as found in nature with its own regulatory sequences. "Chimeric gene" refers to a gene comprising heterogeneous regulatory and coding sequences. "Endogenous gene" refers
10 to the native gene normally found in its natural location in the genome. A "foreign gene" refers to a gene not normally found in the host organism but that is introduced by gene transfer. "Foreign gene" can also refer to a gene that is normally found in the host organism, but that is reintroduced at a location in the genome where it is not normally found, resulting in one or more additional copies of the coding sequence of an
15 endogenous gene.

"Coding sequence" refers to a DNA sequence that codes for a specific protein and excludes the non-coding sequences.

"Initiation codon" and "termination codon" refer to a unit of three adjacent nucleotides in a coding sequence that specifies initiation and chain termination,
20 respectively, of protein synthesis (mRNA translation). "Open reading frame" refers to the amino acid sequence encoded between translation initiation and termination codons of a coding sequence.

"RNA transcript" refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect
25 complementary copy of the DNA sequence, it is referred to as the "primary transcript" or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript. "Messenger RNA" (mRNA) refers to RNA that can be translated into protein by the cell. "cDNA" refers to a double-stranded DNA, one strand of which is complementary to and derived from mRNA by reverse transcription. "Sense" RNA
30 refers to an RNA transcript that includes all or part of an mRNA. "Antisense RNA" refers to an RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene by interfering with the processing, transport, and/or translation of its primary transcript or mRNA. The complementarity of an antisense RNA may be with any part of the specific gene
35 transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns or the coding sequence. In addition, as used herein, antisense RNA may contain regions of

ribozyme sequences that may increase the efficacy of antisense RNA to block gene expression. "Ribozyme" refers to a catalytic RNA and includes sequence-specific endoribonucleases.

As used herein, suitable "regulatory sequences" refer to nucleotide sequences located upstream (5'), within, and/or downstream (3') to a coding sequence, which control the transcription and/or expression of the coding sequences. These regulatory sequences include promoters, translation leader sequences, transcription termination sequences, and polyadenylation sequences. In artificial DNA constructs, regulatory sequences can also control the transcription and stability of antisense RNA.

"Promoter" refers to a DNA sequence in a gene, usually upstream (5') to its coding sequence, which controls the expression of the coding sequence by providing the recognition for RNA polymerase and other factors required for proper transcription. A promoter may also contain DNA sequences that are involved in the binding of protein factors which control the effectiveness of transcription initiation in response to physiological or developmental conditions. It may also contain enhancer elements.

An "enhancer" is a DNA sequence which can stimulate promoter activity. It may be an innate element of the promoter or a heterologous element inserted to enhance the level and/or tissue-specificity of a promoter. "Constitutive" promoters refer to those that direct gene expression in substantially all tissues and demonstrate little temporal or developmental regulation. "Organ-specific" or "development-specific" promoters as referred to herein are those that direct gene expression almost exclusively in specific organs, such as leaves or seeds, or at specific developmental stages in an organ, such as in early or late embryogenesis, respectively.

The term "operably linked" refers to nucleic acid sequences on a single nucleic acid molecule which are associated so that the function of one is affected by the other. For example, a promoter is operably linked with a structural gene (i.e., a gene encoding a starch branching enzyme) when it is capable of affecting the expression of that structural gene (i.e., that the structural gene is under the transcriptional control of the promoter).

The term "expression", as used herein, is intended to mean the production of a functional end-product encoded by a gene. More particularly, "expression" refers to the transcription of the sense (mRNA) or the antisense RNA derived from the nucleic acid fragment(s) of the invention that, in conjunction with the protein apparatus of the cell, results in altered levels of protein product. "Antisense inhibition" refers to the production of antisense RNA transcripts capable of preventing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed

organisms. "Cosuppression" refers to the expression of a gene which is essentially similar to an endogenous gene and results in the suppression of expression of both the ectopic and the endogenous gene. "Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms. The skilled artisan will recognize that the phenotypic effects contemplated by this invention, namely alteration of branch chain distribution in corn starch, can be achieved by alteration of the level of gene product(s) produced in transgenic organisms relative to normal or non-transformed organisms, including a reduction in gene expression mediated by antisense suppression or cosuppression, and enhancement of gene expression by overexpression.

The "3' non-coding sequences" refers to the DNA sequence portion of a gene that contains a polyadenylation signal and any other regulatory signal capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor.

"Transformation" refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as "transgenic" organisms.

The term "pasting" refers to an irreversible physical change in starch granules or a suspension of starch granules characterized by swelling and hydration of granules, a rapid increase in viscosity of a suspension, and the formation of a sol from the suspension. This change is also known as cooking or gelatinization. The abbreviation "SNU" refers to the stirring number unit, approximately equal to 10 centipoise, which is a measure of viscosity. For conversion to SI units (pascal seconds), multiply centipoise by 1000, i.e., 1 PaSec=1000cp. Hence, 1 SNU=0.01 PaSec. The term "sol" refers to a fluid colloidal system. The term "viscosity" is a measure of the internal friction of a fluid that can be thought of as the consistency or thickness of a fluid.

This invention concerns the construction of transgenic corn plants wherein the expression of genes encoding enzymes involved in starch branching are modulated to effect a change in the branch chain distribution of the amylopectin, the relative proportion of amylose to amylopectin, and the degree of polymerization of amylose component of starch. Such modification of starch fine structure results in alteration of the physical properties of starch isolated from said transgenic corn plants. This alteration in the starch fine structure will lead to generation of novel starches possessing properties that are beneficial in food and industrial applications.

A number of genes encoding carbohydrate branching enzymes have been isolated and sequenced. These include glycogen branching enzymes from *Saccharomyces cerevisiae* (Thon et al. (1992) *J. Biol. Chem.* 267:15224-15228), *E. coli* (Baecker et al. (1986) *J. Biol. Chem.* 261:8738-8743), *Bacillus stearothermophilus* (Kiel et al. (1991) *Mol. Gen. Genet.* 230:136-144), *Bacillus caldolyticus* (Kiel et al. (1992) *DNA Seq.* 3: 221-232), human (Thon et al. (1993) *J. Biol. Chem.* 268:7509-7513), *Aspergillus nidulans* (Kiel et al. (1990) *Gene* 89:77-84), *Streptomyces coelicolor* (EMBL accession number X73903), *Streptomyces aurofaciens* (Homerova, D. and Kormanec, J. (1994) *Biochem. Biophys. Acta* 1200:334-336) and starch branching enzymes from corn (Baba et al., (1991) *Biochem. Biophys. Res. Commun.* 181:87-94; Fisher et al. (1993) *Plant Physiol.* 102:1045-1046; Fisher et al. (1995) *Plant Physiol.* 108:1313-1314), pea (Burton et al. (1995) *Plant J.* 7:3-15), potato (Poulsen, P. and Kreiberg, J. D. (1993) *Plant Physiol.* 102:1053-1054), cassava (Salehuzzaman et al. (1992) *Plant Mol. Biol.* 20:809-819), rice (Kawasaki et al. (1993) *Mol. Gen. Genet.* 237:10-16; Mizuno et al. (1993) *J. Biol. Chem.* 268:19084-19091) and *Arabidopsis thaliana* (EMBL accession numbers U18817 and U22428). Preferred among these are the corn starch branching enzyme genes. These genes can be isolated by techniques routinely employed by the skilled artisan for isolation of genes when the nucleotide sequence of the desired gene is known, or when the sequence of a homologous gene from another organism is known. Sequence information about the desired gene can be used to prepare oligonucleotide probes for identification and isolation of the entire branching enzyme gene from an appropriate genetic library. This library may be a genomic library, wherein the coding region may be contained on a single DNA fragment or may be contained on several distinct DNA fragments. Moreover, two or more exons encoding the branching enzyme may be separated by one or more introns. Alternatively, the library may be a cDNA library wherein the likelihood of isolating a cDNA clone comprising the entire coding region as one contiguous sequence is greater. In either instance, the appropriate clone(s) can be identified by DNA-DNA hybridization with probes corresponding to one or more portions of the desired genes. Alternatively, oligonucleotide primers can be prepared and employed as PCR primers in order to amplify and subsequently isolate all or part of the branching enzyme coding region from genomic DNA, or from the genomic or cDNA libraries described above.

Several different assays can be used to measure branching enzyme activity. In the phosphorylase stimulation assay (Boyer, C. D. and Preiss, J. (1978) *Carbohydr. Res.* 61:321-334), activity is measured indirectly by following the ability of branching enzymes to stimulate formation of α -D-glucan from glucose-1-phosphate by

phosphorylase a. The iodine stain assay is based upon the decrease in the absorbance of a glucan-polyiodide complex which occurs as a result of the branching of amylose or amylopectin (ibid). In the third assay, the branch linkage assay, reduced amylose is utilized as the substrate and enzyme activity is followed by measuring the generation of
5 reducing ends following digestion of the product with isoamylase (Takeda et al. (1993) *Carbohydr. Res.* 240:253-262). Guan and Preiss ((1993) *Plant Physiol.* 102:1269-1273) have used the iodine stain and the branch linkage assay, to differentiate the catalytic properties of the three starch branching enzymes in maize. While SBEI exhibits higher activity in branching amylose, SBEIIa and SBEIIb show higher rates of branching with
10 an amylopectin substrate. The isoforms may be further differentiated on the basis of the length of α -1,4-glucan chain that is transferred: SBEI preferentially transfers longer glucan chains while SBEIIa and SBEIIb show a preference in the transfer of shorter chains. Thus, assays which measure enzyme activity may be used to assign a functional activity to proteins which, on the basis of homology at the amino acid level or
15 hybridization at the DNA level, have been identified as starch or glycogen branching enzymes. They may additionally be used to characterize starch or glycogen branching enzymes which have been subjected to mutagenesis schemes designed to identify or alter amino acid residues which play a role in determining catalytic properties. Furthermore, using the findings of Guan and Preiss (Id.), native or mutagenized enzymes may be
20 classified as SBEI or SBEII-like on the basis of substrate or product preferences.

In order to alter the starch fine structure in corn, a chimeric gene is constructed wherein expression of the gene encoding the starch branching enzyme is under the control of regulatory elements suitable to expression of the gene 1) in desired plant tissues, 2) at stages of development that provide the maximum desired effect, and 3) at
25 levels of gene expression that result in alteration of starch branching enzyme function such that expression affects a measurable and significant change in starch fine structure.

The expression of foreign genes in plants is well-established (De Blaere et al. (1987) *Meth. Enzymol.* 143:277-291). Proper level of expression of sense or antisense branching enzyme genes in corn may require the use of different chimeric genes utilizing
30 different regulatory elements. Moreover, effective modulation of endogenous branching enzyme gene expression by cosuppression or antisense suppression may require construction of chimeric genes comprising different regions of the branching enzyme sense or antisense sequences. The well-known unpredictability of the cosuppression and antisense techniques indicates that even while using different genetic constructs, multiple
35 plants may have to be screened in order to identify those with the desired phenotype.

Promoters utilized to drive gene expression in transgenic plants can be derived from many sources so long as the chosen promoter(s) have sufficient transcriptional activity to accomplish the invention by expressing translatable mRNA or antisense RNA in the desired host tissue. For example, promoters for expression in a wide array of plant
5 organs include those directing the 19S and 35S transcripts in Cauliflower mosaic virus (Odell et al. (1985) *Nature* 313:810-812; Hull et al. (1987) *Virology* 86:482-493), small subunit of ribulose 1,5-bisphosphate carboxylase (Morelli et al. (1985) *Nature* 315:200-204; Broglie et al. (1984) *Science* 224:838-843; Herrera-Estrella et al. (1984) *Nature* 310:115-120; Coruzzi et al. (1984) *EMBO J.* 3:1671-1679; Faciotti et al. (1985)
10 *Bio/Technology* 3:241 and chlorophyll a/b binding protein (Lamppa et al. (1986) *Nature* 316:750-752).

Depending upon the application, it may be desirable to select promoters that are specific for expression in one or more organs of the plant. Examples include the light-inducible promoters of the small subunit of ribulose 1,5-bisphosphate carboxylase, if the
15 expression is desired in photosynthetic organs, or promoters active specifically in seeds.

Preferred promoters are those that allow expression specifically in seeds. This may be especially useful, since seeds are the primary location of long-term starch accumulation. In addition, seed-specific expression may avoid any potential deleterious effects that branching enzyme modulation may have on non-seed organs. Examples of
20 seed-specific promoters include, but are not limited to, the promoters of seed storage proteins. The seed storage proteins are strictly regulated, being expressed almost exclusively in seeds in a highly organ-specific and stage-specific manner (Higgins et al. (1984) *Ann. Rev. Plant Physiol.* 35:191-221; Goldberg et al. (1989) *Cell* 56:149-160; Thompson et al. (1989) *BioEssays* 10:108-113). Moreover, different seed storage
25 proteins may be expressed at different stages of seed development.

There are currently numerous examples for seed-specific expression of seed storage protein genes in transgenic plants. These include genes from monocotyledonous plants such as for barley β -hordein (Marris et al. (1988) *Plant Mol. Biol.* 10:359-366) and wheat glutenin (Colot et al. (1987) *EMBO J.* 6:3559-3564). Moreover, promoters
30 of seed-specific genes, operably linked to heterologous coding sequences in chimeric gene constructs, also maintain their temporal and spatial expression pattern in transgenic plants. Such examples include linking either the Phaseolin or Arabidopsis 2S albumin promoters to the Brazil nut 2S albumin coding sequence and expressing such combinations in tobacco, Arabidopsis, or *Brassica napus* (Altenbach et al. (1989) *Plant*
35 *Mol. Biol.* 13:513-522; Altenbach et al. (1992) *Plant Mol. Biol.* 18:235-245; De Clercq et al. (1990) *Plant Physiol.* 94:970-979), bean lectin and bean β -phaseolin promoters to

express luciferase (Riggs et al. (1989) *Plant Sci.* 63:47-57), and wheat glutenin promoters to express chloramphenicol acetyl transferase (Colot et al. (1987) *EMBO J.* 6:3559-3564).

Of particular use in the expression of the nucleic acid fragment(s) of the invention will be promoters from several extensively characterized corn seed storage protein genes such as endosperm-specific promoters from the 10 kD zein gene (Kiriwara et al. (1988) *Gene* 71:359-370), the 15 kD zein gene (Hoffman et al. (1987) *EMBO J.* 6:3213-3221; Schernthaner et al. (1988) *EMBO J.* 7:1249-1253; Williamson et al. (1988) *Plant Physiol.* 88:1002-1007), the 27 kD zein gene (Prat et al. (1987) *Gene* 52:51-49; Gallardo et al. (1988) *Plant Sci.* 54:211-281), and the 19 kD zein gene (Marks et al. (1985) *J. Biol. Chem.* 260:16451-16459). The relative transcriptional activities of these promoters in corn have been reported (Kodrzyck et al. (1989) *Plant Cell* 1:105-114) providing a basis for choosing a promoter for use in chimeric gene constructs for corn. Moreover, promoters that drive the expression of genes encoding enzymes involved in starch biosynthesis may be used in the practice of this invention. These include the 5' regulatory sequences of the sucrose synthase (Yang, N.-S. and Russell, D. (1990) *Proc. Natl. Acad. Sci.* 87:4144-4148) and the waxy or granule-bound starch synthase I (Unger et al. (1991) *Plant Physiol.* 96:124) genes. Promoter elements may be derived from other starch synthase (granule-bound and soluble isoforms) genes when these become available, and from the sh2 (Bhave et al. (1990) *Plant Cell* 2:581-588) and bt2 (Bae et al. (1990) *Maydica* 35:317-322) genes whose products constitute the enzyme ADP-glucose pyrophosphorylase. The isolation of genomic clones encoding the starch branching enzyme genes may be accomplished using the corresponding cDNA clones (Baba et al. (1991) *Biochem. Biophys. Res. Commun.* 181:87-94; Fisher et al. (1993) *Plant Physiol.* 102:1045-1046) as hybridization probes. These would provide a useful starting point for the isolation of promoter fragments of these genes. For assembly of SBE constructs, the upstream sequences may be donated by the cognate SBEII gene or, alternatively, by the SBEI gene.

It is envisioned that the introduction of enhancers or enhancer-like elements into other promoter constructs will also provide increased levels of primary transcription to accomplish the invention. These would include viral enhancers such as that found in the 35S promoter (Odell et al. (1988) *Plant Mol. Biol.* 10:263-272), enhancers from the opine genes (Fromm et al. (1989) *Plant Cell* 1:977-984), or enhancers from any other source that result in increased transcription when placed into a promoter operably linked to the nucleic acid fragment of the invention.

Introns isolated from the maize Adh-1 and Bz-1 genes (Callis et al. (1987) *Genes Dev.* 1:1183-1200), and intron 1 and exon 1 of the maize Shrunken-1 (sh-1) gene (Maas et al. (1991) *Plant Mol. Biol.* 16:199-207) may also be of use to increase expression of introduced genes. Results with the first intron of the maize alcohol dehydrogenase (Adh-1) gene indicate that when this DNA element is placed within the transcriptional unit of a heterologous gene, mRNA levels can be increased by 6.7-fold over normal levels. Similar levels of intron enhancement have been observed using intron 3 of a maize actin gene (Luehrsen, K. R. and Walbot, V. (1991) *Mol. Gen. Genet.* 225:81-93). Enhancement of gene expression by Adh1 intron 6 (Oard et al. (1989) *Plant Cell Rep* 8:156-160) has also been noted. Exon 1 and intron 1 of the maize sh-1 gene have been shown to individually increase expression of reporter genes in maize suspension cultures by 10 and 100-fold, respectively. When used in combination, these elements have been shown to produce up to 1000-fold stimulation of reporter gene expression (Maas et al. (1991) *Plant Mol. Biol.* 16:199-207).

Any 3' non-coding region capable of providing a polyadenylation signal and other regulatory sequences that may be required for proper expression can be used to accomplish the invention. This would include the 3' end from any storage protein such as the 3' end of the 10kd, 15kd, 27kd and alpha zein genes, the 3' end of the bean phaseolin gene, the 3' end of the soybean b-conglycinin gene, the 3' end from viral genes such as the 3' end of the 35S or the 19S cauliflower mosaic virus transcripts, the 3' end from the opine synthesis genes, the 3' ends of ribulose 1,5-bisphosphate carboxylase or chlorophyll a/b binding protein, or 3' end sequences from any source such that the sequence employed provides the necessary regulatory information within its nucleic acid sequence to result in the proper expression of the promoter/coding region combination to which it is operably linked. There are numerous examples in the art that teach the usefulness of different 3' non-coding regions (for example, see Ingelbrecht et al. (1989) *Plant Cell* 1:671-680).

Various methods of introducing a DNA sequence (i.e., of transforming) into eukaryotic cells of higher plants are available to those skilled in the art (see EPO publications 0 295 959 A2 and 0 138 341 A1). Such methods include high-velocity ballistic bombardment with metal particles coated with the nucleic acid constructs (see Klein et al. (1987) *Nature* (London) 327:70-73, and see U.S. Pat. No. 4,945,050), as well as those based on transformation vectors based on the Ti and Ri plasmids of *Agrobacterium spp.*, particularly the binary type of these vectors. Ti-derived vectors transform a wide variety of higher plants, including monocotyledonous and dicotyledonous plants, such as soybean, cotton and rape (Pacciotti et al. (1985)

Bio/Technology 3:241; Byrne et al. (1987) *Plant Cell, Tissue and Organ Culture* 8:3; Sukhapinda et al. (1987) *Plant Mol. Biol.* 8:209-216; Lorz et al. (1985) *Mol. Gen. Genet.* 199:178-182; Potrykus et al. (1985) *Mol. Gen. Genet.* 199:183-188).

Other transformation methods are available to those skilled in the art, such as
 5 direct uptake of foreign DNA constructs (see EPO publication 0 295 959 A2), and techniques of electroporation (see Fromm et al. (1986) *Nature* (London) 319:791-793). Once transformed, the cells can be regenerated by those skilled in the art. Also relevant are several recently described methods of introducing nucleic acid fragments into commercially important crops, such as rapeseed (see De Block et al. (1989) *Plant*
 10 *Physiol.* 91:694-701), sunflower (Everett et al., (1987) *Bio/Technology* 5:1201-1204), soybean (McCabe et al. (1988) *Bio/Technology* 6:923-926; Hinchey et al. (1988) *Bio/Technology* 6:915-922; Chee et al. (1989) *Plant Physiol.* 91:1212-1218; Christou et al. (1989) *Proc. Natl. Acad. Sci USA* 86:7500-7504; EPO Publication 0 301 749 A2), and corn (Gordon-Kamm et al. (1990) *Plant Cell* 2:603-618; Fromm et al. (1990)
 15 *Bio/Technology* 8:833-839).

One skilled in the art is familiar with still other means for the production of transgenic maize plants including introduction of DNA into protoplasts and regeneration of plants from said protoplasts (Omirulleh et al. (1993) *Plant Mol. Biol.* 21:415-423), electroporation of intact tissues (D'Hulluin et al. (1992) *Plant Cell* 4:1495-1505;
 20 Laursen et al. (1994) *Plant Mol. Biol.* 24:51-61), silica carbide mediated fiber transformation of maize cells (Kaeppler et al. (1992) *Theor. Appl. Genet.* 84:560-566; Frame et al. (1994) *Plant J.* 6:941-948). In addition to the method of particle bombardment of maize callus cells described above, one skilled in the art is familiar with particle bombardment of maize scutellar or suspension cultures to yield fertile transgenic
 25 plants (Koziel et al. (1993) *Bio/Technology* 11:194-200; Walters et al. (1992) *Plant Mol. Biol.* 18:189-200).

Once transgenic plants are obtained by one of the methods described above, it will be necessary to screen individual transgenics for those that most effectively display the desired phenotype. It is well known to those skilled in the art that individual
 30 transgenic plants carrying the same construct may differ in expression levels; this phenomenon is commonly referred to as "position effect". For example, when the construct in question is designed to express higher levels of the gene of interest, individual plants will vary in the amount of the protein produced and thus in enzyme activity; this in turn will effect the phenotype.

35 The person skilled in the art will know that special considerations are associated with the use of antisense or cosuppression technologies in order to reduce expression of

particular genes. U. S. Pat. Nos. 5,190,931, 5,107,065 and 5,283,323 have taught the feasibility of these techniques, but it is well known that their efficiency is unpredictable. In either case, in order to save time, the person skilled in the art will make multiple genetic constructs containing one or more different parts of the gene to be suppressed, 5 since the art does not teach a method to predict which will be most effective for a particular gene. Furthermore, even the most effective constructs will give an effective suppression phenotype only in a fraction of the individual transgenic lines isolated. For example, WO93/11245 and WO94/11516 teach that when attempting to suppress the expression of fatty acid desaturase genes in canola, actual suppression was obtained in 10 less than 1% of the lines tested. In other species the percentage is somewhat higher, but in no case does the percentage reach 100.

This should not be seen as a limitation on the present invention, but instead as practical matter that is appreciated and anticipated by the person skilled in this art. Accordingly, skilled artisan will develop methods for screening large numbers of 15 transformants. The nature of these screens will generally be chosen on practical grounds, and is not an inherent part of the invention. In the instant case, for example, one can screen by looking for changes in starch phenotype using chromatography to determine relative proportions of amylose to amylopectin, amylopectin branch chain distribution, RVA analysis (as is done in the examples), or other means. One could equally use 20 antibodies specific for the protein encoded by the gene being suppressed, or one could establish assays that specifically measure enzyme activity. A preferred method will be one which allows large numbers of samples to be processed rapidly, since it will be expected that the majority of samples will be negative.

Plants that are identified to have the altered starch fine structure in the grain 25 present unique genetic material which provide advantages over traditional corn lines and known starch mutants. Use of lines with inhibited expression of SBE isoforms in corn breeding provide a dominant trait that can simplify and speed the breeding process. Known starch mutants can be used but they are often recessive and present more complications. Further, the use of antisense or cosuppression to inhibit SBE isoforms 30 leads to variable levels of inhibition due to chromosomal position effects. The resulting variable levels of SBE activities would lead to a wide range of phenotypes that is not possible using traditional mutants which can result in a limited dosage series of a mutant allele in corn endosperm. Additional unique and potentially valuable starch fine structures will result from crossing the newly developed corn lines with inhibited SBE 35 with each other and/or known starch mutants such as *wx* or *ae*.

EXAMPLES

The present invention is further defined in the following examples. It will be understood that the examples are given for illustration only and the present invention is not limited to uses described in the examples. The present invention can be used to generate transgenic corn plants whose altered starches may be used for any purpose where its properties are useful such as in, but not limited to, foods, paper, plastics, adhesives, or paint. From the above discussion and the following examples, one skilled in the art can ascertain, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions. All such modifications are intended to fall within the scope of the intended claims.

EXAMPLE 1

Preparation of Transgenic Corn Expressing a 3' Antisense Transcript
of Corn Starch Branching Enzyme I Ib

The cDNA insert of plasmid clone pBE240 was used as the starting point in the assembly of DNA constructs designed to achieve suppression of SBEI Ib expression in transgenic corn plants. The cDNA clone pBE240, encoding corn starch branching enzyme I Ib (hereinafter SBEI Ib), has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852), and bears the following accession number: ATCC 97365. pBE 240 (Figure 1) comprises a 2.7 kbp EcoRI-XhoI fragment isolated from a corn cDNA library, inserted into the plasmid vector pbluescriptTMSK+ (Stratagene). The insert (SEQ ID NO:1) consists of 78 bp of 5' untranslated DNA, a 2397 bp open reading frame encoding the corn SBEI Ib coding region and 190 bp of 3' untranslated DNA.

Preparation of the Expression Vector Encoding the 3' Antisense Construct

The chimeric gene inserted into plasmid construct pBE44 (Figure 2) comprises a 3' fragment of the SBEI Ib cDNA in antisense orientation with respect to the maize 27 kD zein promoter that is located 5' to the SBEI Ib fragment, and the 10 kD zein 3' end that is located 3' to the SBEI Ib fragment. The SBEI Ib fragment of this construct was generated by polymerase chain reaction (PCR) of pBE240 using appropriate oligonucleotide primers. These primers were synthesized on a Beckman Oligo 1000TM DNA Synthesizer. The 414 bp fragment of pBE44 (SEQ ID NO:2) was generated using the oligonucleotide pair BE41 (SEQ ID NO:3) and BE42 (SEQ ID NO:4):

BE41 5' - GAATTC CGGGGTGTTCAACTTCCACTGC - 3' (SEQ ID NO:3)
BE42 5' - GAATTC CATGGGACACCTTGAAGGTCTT - 3' (SEQ ID NO:4)

Cloning sites (NcoI or SmaI) were incorporated into the oligonucleotides to provide antisense orientation of the DNA fragments when inserted into the digested vector pML103 as described below. Amplification was performed in a 100 µl volume in a standard PCR mix consisting of 0.4 mM of each oligonucleotide and 0.3 µM of pBE240 in 10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% w/v gelatin, 200 mM dGTP, 200 mM dATP, 200 mM dTTP, 200 mM dCTP and 0.025 unit Amplitaq™ DNA polymerase. Reactions were carried out in a Perkin-Elmer Cetus Thermocycler™ for 30 cycles comprising 1 minute at 95°C, 2 minutes at 55°C and 3 minutes at 72°C, with a final 7 minute extension at 72°C after the last cycle. The amplified DNA was digested with restriction enzymes NcoI and SmaI and fractionated on a 0.7% low melting point agarose gel in 40 mM Tris-acetate, pH 8.5, 1 mM EDTA. The appropriate band was excised from the gel, melted at 68°C and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103 (Figure 3). Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852), and bears the following accession number: ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA were ligated at 15°C overnight, essentially as described (Sambrook, J., Fritsch, E. F. and Maniatis, T. (1989) Molecular Cloning, Cold Spring Harbor Laboratory Press, New York; hereinafter "Maniatis"). The ligated DNA was used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants were screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U. S. Biochemical). The resulting plasmid construct, pBE44, comprises a chimeric gene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a 3' fragment of the corn SBEIIb cDNA, and the 10 kD zein 3' region.

Larger quantities of pBE44 plasmid DNA were prepared by the alkaline lysis method, followed by purification by CsCl density gradient centrifugation.

Transformation of Corn with the 3' Antisense Construct

Immature corn embryos were dissected from developing caryopses derived from crosses of the inbred corn lines H99 and LH132. The embryos were isolated 10 to 11 days after pollination when they were 1.0 to 1.5 mm long. The embryos were placed with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al. (1975), *Sci. Sin. Peking* 18:659-668). The embryos were kept in the dark at 27°C.

Friable embryogenic callus consisting of undifferentiated masses of cells with somatic proembryoids and embryoids borne on suspensor structures proliferates from the scutellum of these immature embryos. The embryogenic callus isolated from the primary explant was cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (Figure 4; obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, Germany) was used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al. (1987), *Nature* 327:70-73) was used to transfer genes to the callus culture cells. Gold particles (1 μm in diameter) were coated with DNA using the following technique. Plasmid DNAs (10 μg of p35S/Ac and 10 μg of pBE44) were added to 50 μl of a suspension of gold particles (60 mg per ml). Calcium chloride (50 μl of a 2.5 M solution) and spermidine free base (20 μl of a 1.0 M solution) were added to the particles. The suspension was vortexed during the addition of these solutions. After 10 minutes, the tubes were briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles were resuspended in 200 μl of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse was performed again and the particles resuspended in a final volume of 30 μl of ethanol. An aliquot (5 μl) of the DNA-coated gold particles was placed in the center of a KaptonTM flying disc (Bio-Rad Labs). The particles were accelerated into the corn tissue with a BiolisticTM PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue was placed on filter paper over agarose-solidified N6 medium. The tissue was arranged as a thin lawn and covered a circular area of about 5 cm in diameter. The petri dish containing the tissue was placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber was then evacuated to a vacuum of 28 inches of Hg. The macrocarrier was accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock tube reaches 1000 psi.

Seven days after bombardment the tissue was transferred to N6 medium that contained gluphosinate (2 mg per liter) and lacked casein or proline. The tissue

continued to grow slowly on this medium. After an additional 2 weeks the tissue was transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus were identified on some of the plates containing the glufosinate-supplemented medium. These calli continued to grow when sub-cultured on the selective medium.

Plants were regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue was transferred to regeneration medium (Fromm et al. (1990) *BioTechnology* 8:833-839). A total of 9 corn plants were regenerated from a single transformation experiment using the pBE44 construct.

Molecular Analysis of Transgenic Corn Plants Containing the 3' Antisense Construct

Total DNA was isolated from leaf tissue of plants regenerated from the transformation experiment using pBE44 essentially as described by Dellaporta et al. (Dellaporta et al. (1983) *Plant Mol. Biol. Rep.* 1 (4):9). Lyophilized tissue was frozen in liquid nitrogen, ground to a fine powder and suspended in a buffer consisting of 100 mM Tris-HCl, pH 8.0, 50 mM EDTA, 10 mM b-mercaptoethanol and 0.5 M NaCl. Cells were lysed by the addition of SDS to 1% and the DNA precipitated with isopropanol. The dissolved DNA was treated with DNase-free RNase and then re-precipitated with iso-propanol. The isolated DNAs were dissolved in 10 mM Tris-HCl, pH 8.0, 1 mM EDTA and stored at -20°C until use.

For Southern blot analysis, 5 mg of isolated DNA was digested with restriction enzyme (10 units/mg DNA) in the appropriate buffer for approximately 6 hrs at 37°C. The restricted DNA was loaded onto a 0.8% agarose gel in Tris-borate-EDTA buffer (Maniatis) and electrophoresed at 40 V overnight. Following denaturation and neutralization, the DNA was transferred to an Immobilon™ membrane (Millipore Corporation) using 10X SSC. The Immobilon™ membrane was pre-hybridized at 65°C in an aqueous buffer system consisting of 6X SSPE, 5X Denhardt's reagent, 0.5% SDS and 100 mg/mL denatured salmon sperm DNA as described (Maniatis). The SBE fragment of pBE44 was labeled by nick translation (BRL Nick Translation Kit) and added to the above buffer supplemented with 5% dextran sulfate at a level of 1-2 x 10⁶ cpm/ml. Hybridization was allowed to proceed at 65°C for 18 h. The membrane was sequentially washed with 2X SSC, 0.1% SDS for 15 minutes at room temperature, 1X SSC, 0.1% SDS for 15 minutes at room temperature and 0.1X SSC, 0.5% SDS for 15 minutes at 50°C. Washed membranes were exposed to Dupont Reflection™ film with an intensifying screen at -80°C.

For Northern blot analysis, total RNA was isolated from kernels harvested 20-22 days after pollination (DAP). Approximately 10 kernels per plant were pooled and frozen in liquid nitrogen. The frozen tissue was ground to a fine powder. A mixture of phenol-chloroform-isoamyl alcohol (24:24:1; 3 ml) was added and the tissue slurry
5 briefly homogenized by hand. 4.5 mL extraction buffer (1 M Tris-HCl, pH 9.0, 1% SDS, 5% β -mercaptoethanol) was mixed in and the suspension was centrifuged (4°C, 7500 rpm, SS-34) to remove debris. The supernatant was extracted with phenol-chloroform-isoamyl alcohol and the nucleic acids collected by ethanol precipitation. RNA was isolated from the dissolved pellet by selective precipitation with 0.2 M LiCl
10 followed by a second precipitation with ethanol. RNA was dissolved in sterile water and stored at -80°C prior to use. RNA concentration was calculated by measuring the absorption of solutions at 260 nm (assuming that $A_{260} = 1$ corresponds to 40 mg/mL).

Total RNA was denatured by reaction with glyoxal and fractionated on a 1% agarose gel in 10 mM sodium phosphate buffer, pH 7.0 (Maniatis). RNA was
15 transferred to a Hybond™ nylon membrane using 20X SSC as the transfer medium and then fixed to the solid support by irradiation in a UV Stratalinker™ (Stratagene). Blots were pre-hybridized at 42°C for 18 h. in a buffer consisting of 50% formamide, 6X SSPE, 5X Denhardt's, 0.5% SDS, 100 mg/mL denatured salmon sperm DNA. Hybridization was carried out at 42°C for 18-24 h in the same buffer supplemented with
20 5% dextran sulfate and containing $1-2 \times 10^6$ cpm/mL denatured, nick translated probe. Blots were washed at room temperature for 15 minutes in 2X SSC, 0.1% SDS, followed by 15 minutes in 1X SSC, 0.1% SDS. This was followed by a third wash for 15 minutes at 50°C in 0.1X SSC, 0.5% SDS. Washed blots were exposed at -80°C while still damp to Dupont Reflection™ film with an intensifying screen.

25 Of the 9 transgenic plant lines that were regenerated from particle bombardments performed with the pBE44 construct, seven of these were identified by Southern blot analysis to contain the trait gene. Northern blots of total RNA isolated from these lines showed variable levels of SBEIIb RNA; in 6 of the analyzed lines, a 500 base transcript was also observed. The size of this hybridizing RNA is consistent with that predicted for
30 the antisense transcript from the chimeric gene of pBE44.

Analysis of Starch from Transformed Corn Plants Containing the 3' Antisense Construct

Starch was extracted from single seeds obtained from corn plants transformed with the 3' antisense construct. Seeds were steeped in a solution containing 1.0% lactic acid and 0.3% sodium metabisulfite, pH 3.82, held at 52°C for 22-24 h. Seeds were
35 drained, rinsed and homogenized individually in 8-9 mL of a solution of 100 mM NaCl. Five mL of toluene were added to each tube and vigorously shaken twice for 6 minutes

using a paint mixer, and allowed to settle for 30 minutes. Two mL of 100 mM NaCl was sprayed onto the solution, allowed to settle for 30 minutes, and the protein-toluene layer was aspirated off. The toluene wash step was repeated. Twelve mL water was added and shaken in a paint shaker for 45 seconds. This solution was centrifuged for
5 10 minutes and the water was removed. The water wash was repeated, followed by a final wash with 12 mL of acetone. After shaking and centrifugation steps, the acetone was drained and allowed to evaporate for 1 h. Starch extracts were incubated in a 40°C oven overnight.

Extracted starches were enzymatically debranched as follows. Extracted starches
10 (10 mg) from individual seeds were gelatinized in 2 mL water by heating to 115°C for 0.5 h. Four units of isoamylase (Sigma) in 50 mM NaOAc buffer, pH 4.5, were added to each of the gelatinized starches and placed in a water bath at 45°C for 2.5 h. Enzyme was inactivated by heating samples to 115°C for 5 minutes. Each sample was filtered through a 0.45 micron filter, and placed into individual autosampler vials. Samples were
15 held at 45°C until injection.

Fifty mL of debranched starch sample were injected and run through four columns (3 x 250 Å and 1 x 500 Å ultrahydrogel™; Waters) arranged in series at 45°C and eluted with 50 mM NaOAc at a flow rate of 0.7 mL/min. Sampling interval was 65 minutes. A refractive index detector (Waters), integrator/plotter (Spectra-Physics)
20 and computer were used for sample detection, recording of retention times and chromatogram storage, respectively. Retention times of collected samples were compared to retention times of pullulan standards (380K, 100K, 23.7K, 5.8K, 728 and 180 mw).

Spectra-Physics software was used to make any baseline corrections to the
25 chromatogram including subtraction of a blank chromatogram. Spectra-Physics GPC-PC software was used to enter molecular weights and retention times of pullulan standards. The data were imported to Microsoft Excel for parsing and stripping of all data except molecular weight and area percent of the chromatogram. The remaining data were used to determine branch chain distribution of the amylopectin using Jandel Scientific Peakfit
30 software. A series of six Gaussian curves were fit to the amylopectin portion of the chromatograms as described by Ong et al. ((1994) *Carbohydrate Res.* 260:99-117).

Amylopectin is typically described by its distribution of branch chains in the molecule. The amylopectin molecule is comprised of alternating crystalline and amorphous regions. The crystalline region is where many of the branch points (α -1,6
35 linkages) occur, while the amorphous region is an area of little to no branching and few branch chains. The type of chain is designated A or B. A chains are unbranched and

span a single crystalline region. B1 chains also span a single crystalline region but are branched. B2, B3 and B4+ chains are branched and span 2, 3 and 4 or more crystalline regions, respectively (Hizukuri (1986) *Carbohydrate Res.* 147:342-347). The relative area under the six Gaussian curves fit to the amylopectin portion of the chromatograms using Peakfit software was used to determine the area percentage of the A, B1, B2, B3 and B4+ chains. The areas of the first and second peaks were summed to give the relative amount of A and B1 chains, the third and fourth peaks represent the B2 and B3 chains, respectively, and the sum of the fifth and sixth peaks represent the relative area of the B4+ chains. The mass average DP of the A and B1, B2, B3, and B4 chains were 14, 22, 43 and 69 respectively.

Starches from individual R1 kernels of plants transformed with pBE44 (the 3' antisense construct for corn SBEIIb) were analyzed using the procedure described above. As known to those skilled in the art, the antisense phenomenon is generally not observed in every individual transgenic line. Therefore, individual kernels from multiple lines were examined and as expected, some, but not all lines possessed kernels demonstrating an altered starch phenotype. Individual kernels from a negative control plant (Transformation Negative Control Line 03376; this line has been through the transformation process but does not carry the antisense gene) were included in each set of assays, and duplicate assays were performed on starches from individual kernels. Table 1 presents the results for individual kernels (kernel Nos. 1 and 7) from a transformed corn line (0693) which did show a phenotype. The data represent the percentage difference of the various branches between kernels of the transformed line and kernels from a negative control (line 03376, which has been through the transformation process but does not contain the antisense gene).

Table 1. Percentage Difference of Branch Chain Distribution of Amylopectin from Starch Isolated from Individual Seed from 3' Antisense SBEIIb Transgenic Corn Line (0693) Compared to Starch Isolated from Negative Control Line (03376).

<u>Starch Source</u>	<u>A + B1</u>	<u>B2</u>	<u>B3</u>	<u>B4+</u>
06931	80	95	104	226
06937	91	90	100	194

Both the experimental (06931 and 06937) and control (03376) data are the average of duplicate assays of starches isolated from individual kernels. As can be seen, there is an approximately 2-fold increase (226% of control and 194% of control for 06931 and 06937, respectively) in long (B4+) chains, indicating that long chains (B4+) are favored at the expense of shorter chains (A's, B1's and B2's) in starches possessing

the antisense gene relative to control starch. The instant transgenic plants thus demonstrate a unique starch branching phenotype compared to non-transgenic control plants. This data indicates that alteration of corn starch branching enzyme activity by suppressing expression of the corresponding genes encoding starch branching enzymes results in an altered starch phenotype.

R1 kernels from the pBE44 line, 0693, were planted and R2 grain was produced. Individual R2 kernels were analyzed using the same procedure as described above for analysis of R1 kernels. Individual kernels from a negative control line (04659, which has been through the transformation process but does not carry the antisense gene) were included in this set of assays. Table 2 presents the results for R2 kernels. The data represent the percentage difference of the various branches between R2 kernels and kernels from the negative control.

Table 2. Percentage Difference of Branch Chain Distribution of Amylopectin From Starch Isolated From Individual R2 Seed From 3' Antisense SBEIIb Transgenic Corn Line (05985) Compared to Starch Isolated From Negative Control Line (04659).

<u>Starch Source</u>	<u>A + B1</u>	<u>B2</u>	<u>B3</u>	<u>B4+</u>
059852	69	91	132	476
0598510	71	92	129	455

As can be seen, long chains (B3 and B4+) are favored at the expense of shorter chains (A's, B1's and B2's) in the amylopectin derived from R2 kernels possessing the antisense gene relative to control starch (04659). The instant transgenic plant thus demonstrates a unique starch branching phenotype compared to non-transgenic control plants. This data also indicates that the phenotype observed in the R2 seed is stronger than that of the R1 seed (Table 1) which may be due to segregation.

R4 grain (line XAY00681) was produced, harvested and starch was extracted. For starch branch chain distribution and determination of amylose content, starch digestion was modified from that in previous examples slightly as follows. Seven mg of each starch sample was added to a screw cap test tube with 1.1 mL of water. The tubes were heated to 120°C for 30 minutes and then placed in a water bath at 45°C. Debranching solution was made by diluting 50 μ L of isoamylase (5×10^6 units/mL, Sigma) per mL of sodium acetate buffer (50 mM, pH 4.5). 40 μ L of debranching solution was added to each starch sample and incubated for 3 h at 45°C. Reactions were stopped by heating to 110°C for 5 minutes. Debranched starch samples were lyophilized and redissolved in DMSO for analysis by gel permeation chromatography (GPC). One hundred μ L of debranched starch was injected and run through 2 columns (Polymer

Labs, Mixed Bed-C)) in series at 100°C and eluted with DMSO at a flow rate of 1.0 mL/min. Sampling interval was 25 minutes. A refractive index detector (Waters) was used with a computer running Chemstation Software (version A.02.05, Hewlett Packard) for detection and data collection and storage, respectively. Retention times of pullulan standards (380K, 100K, 23.7K, 5.8K, 728 and 180 mw) were used to define molecular weight ranges for the debranched starch samples. The proportion of the total starch was determined for 24 ranges of degree of polymerization (DP) spanning both the amylose and amylopectin portions of the chromatogram. For purposes of comparison to data reported above, the percentage area in appropriate DP ranges was summed to give values for A and B1 chains, B2, B3 and B4+ chains of the amylopectin portion of the chromatogram. The portion of the total area above DP 150 was used to determined amylose content.

Starch from line XAY00681 (R4) and dent starch (control) were debranched and analyzed. The results are shown in Tables 3 and 4 below:

Table 3. The percentage of total chromatographic area within given degree of polymerization (DP) ranges for starch derived from R4 grain containing the 3' antisense transcript of corn SBE IIb and normal dent starch (control). Averages (n=12) and standard errors of the mean (SE) are reported.

DP range	Dent Starch		XAY00681	
	Average	SE	Average	SE
>5k	5.45	0.14	5.59	0.63
3-5k	2.62	0.05	3.15	0.06
1.8-3k	3.03	0.04	3.89	0.09
1.2-1.8k	2.49	0.05	3.54	0.10
0.9-1.2k	1.92	0.04	2.67	0.06
600-900	2.86	0.03	3.91	0.09
400-600	2.78	0.05	3.83	0.08
250-400	2.83	0.05	3.83	SE
150-250	2.43	0.04	3.50	0.09
90-150	2.38	0.04	3.50	0.09
60-90	4.04	0.08	6.10	0.07
48-60	4.08	0.07	4.81	0.04
40-48	3.95	0.09	3.96	0.05
32-40	4.52	0.13	4.45	0.05
28-32	3.45	0.12	2.89	0.04
24-28	3.69	0.17	3.37	0.06

21-24	4.72	0.18	3.74	0.05
18-21	6.01	0.03	4.83	0.10
15-18	8.42	0.05	6.18	0.12
13-15	7.24	0.21	5.34	0.11
11-15	6.64	0.17	4.49	0.10
9-11	6.20	0.08	4.54	0.11
7-9	4.48	0.06	3.40	0.07
5-7	3.67	0.07	2.91	0.05

Table 4. Percentage Difference of Branch Chain Distribution of Amylopectin (expressed as A+B1, B2, B3 and B4+) and Amylose Content (% of Total Starch) from Starch Isolated from R4 Grain containing the 3' Antisense Transcript of Corn SBEIIb (XAY00681) as Compared to Control (Dent). DP range is indicated.

<u>A+B1 (5-15)</u>	<u>B2 (15-32)</u>	<u>B3 (32-60)</u>	<u>B4+ (60-150)</u>	<u>Amylose (>150)</u>
83.3	89.0	117.4	184.5	128.4

As can be seen in Tables 3 and 4, the relative amount of amylose increased as did the proportion of longer branches on amylopectin in starch which contained the 3' antisense transcript of corn SBE IIb compared to a dent control.

Functional Analysis of Starch from Lines Homozygous for the 3' Antisense Construct

Kernels of plants of a line (XAT00025), homozygous for the pBE44 construct, were isolated from the progeny of line 05985 in order to obtain sufficient quantities of starch for functionality testing. Starch was extracted from dry mature kernels from line XAT00025, dent, and *ae* corn. For each line 15 g of kernels were weighed into a 50 mL Erlenmeyer flask and steeped in 50 mL of steep solution (same as above) for 18 h at 52°C. The kernels were drained and rinsed with water. The kernels were then homogenized using a 20 mm Polytron probe (Kinematica GmbH, Kriens-Luzern, Switzerland) in 50 mL of cold 50 mM NaCl. The homogenate was filtered through a 72 micron mesh screen. The filtrate was brought up to a total volume of 400 mL with 50 mM NaCl and an equal volume of toluene was added. The mixture was stirred with a magnetic stir bar for 1 h at sufficient speed to completely emulsify the two phases. The emulsion was allowed to separate overnight in a covered beaker. The upper toluene layer was aspirated from the beaker and discarded. The starch slurry remaining in the bottom of the beaker was resuspended, poured into a 250 mL centrifuge bottle and centrifuged 15 minutes at 25,000 RCF. The supernatant was discarded and the starch was washed sequentially with water and acetone by shaking and centrifuging as above.

After the acetone wash and centrifugation the acetone was decanted and the starch allowed to dry overnight in a fume hood at room temperature.

A Rapid Visco Analyzer (Newport Scientific; Sydney, Australia) with high sensitivity option and Thermocline software was used for pasting curve analysis. For each line, 1.50 g of starch was weighed into the sample cup and 25 mL of phosphate/citrate buffer (pH 6.50) containing 1% NaCl was added. Pasting curve analysis was performed using the following temperature profile: Idle temperature 50°C, hold at 50°C for 0.5 minutes, linear heating to 95°C for 2.5 minutes, linear cooling to 50°C over 4 minutes, hold at 50°C for four minutes.

Results of the Rapid Visco Analyzer pasting analysis are shown in Figure 5. It can be seen that the starch produced by line XAT00025 differs in its pasting properties both from normal dent starch and from a line homozygous for the *ae* mutation. This result demonstrates that the alteration of starch fine structure produced by suppressing expression of a starch branching enzyme can create a starch of novel functionality.

EXAMPLE 2

Preparation of Transgenic Corn Expressing a 5' Antisense Transcript of Corn Starch Branching Enzyme IIb

Preparation of the Expression Vector Encoding the 5' Antisense Construct

The chimeric gene inserted into plasmid construct pBE43 (Figure 6) comprises a 5' fragment of the SBEIIb cDNA in antisense orientation with respect to the maize 27 kD zein promoter, located 5' to the SBEIIb fragment, and the 10 kD zein 3' end, located 3' to the SBEIIb fragment. The SBEIIb fragment of this construct was generated by polymerase chain reaction (PCR) of pBE240 using appropriate oligonucleotide primers. These primers were synthesized on a Beckman Oligo 1000™ DNA Synthesizer. The 507 bp fragment of pBE43 (SEQ ID NO:5) was synthesized using the oligonucleotide pair BE39 (SEQ ID NO:6) and BE40 (SEQ ID NO:7):

BE39 5' -GAATTCCCGGGACCCGGATTTCGCTCTT-3' (SEQ ID NO:6)

BE40 5' -GAATTCATGGTCTATAGAGGCTGTACCG-3' (SEQ ID NO:7)

Cloning sites (NcoI or SmaI) were incorporated into the oligonucleotides to provide antisense orientation of the DNA fragments when inserted into the digested vector pML103 as described below. Amplification was performed in a 100 µl volume in a standard PCR mix consisting of 0.4 mM of each oligonucleotide and 0.3 pM of pBE240 in 10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% w/v gelatin, 200 mM dGTP, 200 mM dATP, 200 mM dTTP, 200 mM dCTP and 0.025 unit Amplitaq™ DNA polymerase. Reactions were carried out in a Perkin-Elmer Cetus

Thermocycler™ for 30 cycles comprising 1 minute at 95°C, 2 minutes at 55°C and 3 minutes at 72°C, with a final 7 minute extension at 72°C after the last cycle. The amplified DNA was digested with restriction enzymes NcoI and SmaI and fractionated on a 0.7% low melting point agarose gel in 40 mM Tris-acetate, pH 8.5, 1 mM EDTA.

- 5 The appropriate band was excised from the gel, melted at 68°C and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103 (Figure 3). The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA was ligated at 15°C overnight,
- 10 essentially as described (Maniatis). The ligated DNA was used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue'; Stratagene™). Bacterial transformants were screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U. S. Biochemical). The resulting plasmid construct, pBE43, comprises
- 15 a chimeric gene encoding in the 5' to 3' direction, the maize 27 kD zein promoter, a 5' fragment of the corn SBEIIb gene in antisense orientation, and the 10 kD zein 3' region.

Larger quantities of pBE43 plasmid DNA were prepared by the alkaline lysis method, followed by purification by CsCl density gradient centrifugation.

Transformation of Corn with the 5' Antisense Construct

- 20 The 5' antisense construct (pBE43) was introduced into embryogenic corn tissue by the particle bombardment method essentially as described in Example 1. Seven days after bombardment the tissue was transferred to N6 medium that contained glufosinate (2 mg per liter) and lacked casein or proline. The tissue continued to grow slowly on this medium. After an additional 2 weeks the tissue was transferred to fresh N6 medium
- 25 containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus were identified on some of the plates containing the glufosinate-supplemented medium. These calli continued to grow when sub-cultured on the selective medium.

- Plants were regenerated from the transgenic callus by first transferring clusters of
- 30 tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue was transferred to regeneration medium (Fromm et al. (1990) *Bio/Technology* 8:833-839). Ninety-nine transgenic plant lines were generated from 2 separate particle bombardment experiments performed with the DNA construct pBE43.

Molecular Analysis of Transformed Corn Plants Containing the 5' Antisense Construct

- 35 Southern blot and Northern blot analyses of DNA and RNA from corn plants transformed with the 5' antisense construct (pBE43) were performed as described in

Example 1. For Southern, the DNA probe was prepared as described in Example 1. Of the ninety-nine transgenic plant lines that were generated from particle bombardment experiments, twenty-eight were subjected to Southern blot analysis using a 666 bp EcoRI-BamHI fragment of the SBEIIb cDNA as a hybridization probe. Twenty lines carrying the trait gene were identified. The pattern of hybridizing bands ranged from fairly simple to rather complex, consistent with duplication and rearrangement of the construct DNA upon to integration into the corn genome.

Total RNA was isolated from 35 pBE43-transformed plant lines. The RNA was denatured, fractionated by gel electrophoresis, blotted onto nylon membranes and hybridized to a probe encompassing the complete SBEIIb cDNA or a 5' portion of it. The level of SBEIIb RNA was found to vary considerably from line to line but in no case was a complete absence of RNA found. This result is not unexpected given that the RNA was prepared from a segregating population of seed. In addition to the 2.7 kb SBEIIb RNA, a smaller RNA species was observed in some of the analyzed plant lines. The intensity of this band was found to vary with 8 lines showing moderate to weak signals and 4 lines showing strong signals. The size of this RNA band, approximately 600 bases, matches that expected from the antisense transcript derived from the chimeric gene.

This identity was confirmed by hybridizing Northern blots to strand specific riboprobes. For generation of single stranded RNA probes, the SBEIIb DNA fragment of construct pBE43 was subcloned into a modified pBLUESCRIPT SK+ vector which contains an NcoI site in place of the XbaI site in the polylinker. For synthesis of the sense (RNA identical) strand, the plasmid was first linearized by digestion with NcoI and transcription carried out by T7 RNA polymerase in the presence of (α - 32 P)rUTP using an RNA Transcription Kit (Stratagene). For synthesis of the antisense RNA probe, the plasmid was linearized by digestion with EcoRI, followed by transcription catalyzed by T3 RNA polymerase. Pre-hybridization of Northern blots was accomplished at 60°C in 50% formamide, 6X SSPE, 1 x Denhardt's solution and 100 mg/ml yeast t-RNA. Hybridization was carried out in the same buffer supplemented with 5% dextran sulfate and containing 1×10^6 cpm/ml of RNA probe for approximately 18 hrs at 60°C. Blots were washed for 15 minutes at room temperature in 2X SSPE, 30 minutes at 70°C in 1X SSPE, 0.1% SDS followed by 30 minutes at 70°C in 0.1X SSPE, 0.5% SDS. Washed blots were exposed at -80°C while still damp to Dupont Reflection film with an intensifying screen. The probe corresponding to the antisense RNA strand detected only the endogenous SBEIIb RNA while the sense probe detected only the 600 base RNA

species. This result is consistent with the identity of the 600 base RNA of the antisense transcript of pBE43.

Analysis of Starch from Transformed Corn Plants Containing the 5' Antisense Construct

Starches from individual R1 kernels of plants transformed with pBE43 (the 5' antisense construct for corn SBEIIb) were extracted and analyzed using the procedure described in Example 1. As known to those skilled in the art, the antisense or cosuppression phenomenon is generally not observed in every individual transgenic line. Therefore, individual kernels from multiple lines were examined. No alterations in starch branch chain distribution were observed for the transgenic lines that were screened. It is believed that the number of lines tested was too small to insure finding a plant where an effective antisense event occurred. As described above, the number of plants that must be screened can be unpredictable and large. It is assumed that if a sufficiently large number of individuals were examined such an event would be detected. It may be that this particular configuration is less efficient for suppressing expression of this gene; it is for this reason that multiple constructs were prepared and tested.

EXAMPLE 3

Preparation of Transgenic Corn Expressing a Near Full Length Antisense Transcript of Corn Starch Branching Enzyme IIb

Preparation of the Expression Vector Encoding the Near Full Length Antisense Construct

The construct pBE45 is similar to pBE43 and pBE44 except that the SBEIIb fragment is 2.16 kb and contains the entire 5' untranslated region as well as 2.08 kb of the coding region (SEQ ID NO:8). pBE240 was first digested with EcoRI and then subjected to an end filling reaction with the Klenow fragment of DNA polymerase I (Maniatis). The blunt-ended DNA was fractionated on a low melting point agarose gel and the excised band combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103 (Figure 3). The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA were ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA was used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants were screened for the presence of and the orientation of the added DNA by restriction enzyme digestion with KpnI and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U. S. Biochemical). According to this analysis, in pBE45, the SBEIIb fragment is present in inverse orientation relative to the 27 kD zein promoter.

The resulting plasmid construct, pBE45, comprises a chimeric gene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, the near full length fragment of corn SBEIIb in antisense orientation, and the 10 kD zein 3' region (Fig. 7).

Larger quantities of pBE45 plasmid DNA were prepared by the alkaline lysis method, followed by purification by CsCl density gradient centrifugation.

Transformation of Corn with the Near Full Length Antisense Construct

The near full length antisense construct (pBE45) was introduced into embryogenic corn tissue by the particle bombardment method essentially as described in Example 1. Seven days after bombardment, the tissue was transferred to N6 medium that contained glufosinate (2 mg per liter) and lacked casein or proline. The tissue continued to grow slowly on this medium. After an additional 2 weeks the tissue was transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus were identified on some of the plates containing the glufosinate-supplemented medium. These calli continued to grow when sub-cultured on the selective medium.

Plants were regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue was transferred to regeneration medium (Fromm et al. (1990) *Bio/Technology* 8:833-839). Ten transgenic plant lines were generated from a single particle bombardment experiment performed with the DNA construct pBE45.

Molecular Analysis of Transformed Corn Plants Containing the Near Full Length Antisense Construct

Southern blot and Northern blot analyses of DNA and RNA from corn plants transformed with the near full length antisense construct (pBE45) were performed essentially as described in Example 1. For Southern blots, the DNA probe, an EcoRI-BamHI 5' fragment of pBE240, was prepared essentially as described in Example 1. Of the 10 transgenic plant lines that were generated, 5 tested positive for the presence of the introduced trait gene.

Northern blots of total RNA revealed only a single band when probed with the EcoRI-BamHI 5' fragment of the SBEIIb cDNA. Since the SBEIIb RNA and the pBE45 antisense transcript are similar in size, 2.7 and 2.4 kb respectively, it seemed possible that the two species might not be adequately resolved during agarose gel electrophoresis. For this reason, Northern blots were also hybridized to strand specific RNA probes, essentially as described in Example 1. However, while the antisense strand detected the endogenous SBEIIb mRNA, no signal was evident when the sense strand probe was employed.

amplify this region of the gene by PCR using corn total DNA as the template. A 571 bp 5' fragment was isolated, sequenced and found to be identical to the cDNA over nucleotides 49 to 188. pBE65 was then used as a starting point in the generation of sense and antisense SBEI constructs including pBE68 and pBE97 described below. In the time since these constructs were made and introduced into corn, a second SBEI sequence became available (Fisher et al. (1995) *Plant Physiol.* 108:1313-1314). The 5' terminal 165 bp of pBE65 showed poor agreement with this sequence as it did with the previous SBEI sequence. As a result of subsequent experiments, it is now concluded that pBE65 contains a 165 bp 5' terminal segment that is not related to SBEI but which presumably arose as an artifact during the cloning of corn cDNA. This region is followed by 2607 bp of SBEI cDNA which encodes 42 amino acids of the SBEI transit peptide, the 760 amino acids of the mature SBEI protein and contains 194 bp of 3' untranslated DNA. The plasmid pBE65 has been deposited under the terms of the Budapest Treaty at the ATCC (American Type Tissue Culture Collection, 12301 Parklawn Drive, Rockville, MD 20852) and bears the following accession number: _____.

Preparation of Expression Vectors Encoding SBEI Antisense Constructs

Since it was not known which portions of the cDNA sequence would be most effective in mediating suppression of SBEI expression, three constructs bearing different SBEI fragments in antisense orientation were made. The chimeric gene of plasmid pBE68 (Figure 9) comprises a 3' fragment of the SBEI cDNA in antisense orientation with respect to the maize 27 kD zein promoter that is located 5' to the SBEI fragment, and the 10 kD zein 3' end that is located 3' to the SBEI fragment. The 373 bp SBEI fragment of this construct (SEQ ID NO:13) was obtained by PCR of pBE65 using the oligonucleotide primer pair BE43 (SEQ ID NO:14) and BE52 (SEQ ID NO:15):

BE43 5' - GAATTCCCGGGCCGAAC TCGTTCAAAG - 3' (SEQ ID NO:14)

BE52 5' - GAATTC CATGGCGGTGATGAGACAC CAGTC - 3' (SEQ ID NO:15)

The chimeric gene of pBE69 (Figure 10) is analogous to that of pBE68 except that the SBEI fragment consists of a 5' portion of the SBEI cDNA. The 571 bp fragment of this construct (SEQ ID NO:16) was obtained by amplification of pBE65 using the primer pair BE46 (SEQ ID NO:17) and BE50 (SEQ ID NO:18):

BE46 5' - GAATTC CATGGCCATCTTATGGTTTGCACC - 3' (SEQ ID NO:17)

BE50 5' - GAATTC CCGGCATAGCATAGATATGACGGC - 3' (SEQ ID NO:18)

Cloning sites (NcoI and SmaI) were incorporated into the above oligonucleotides to provide antisense orientation of the DNA fragments when inserted into the vector pML103 described in Example 1. Amplification was performed in a 100 ml volume in a standard PCR reaction mix as defined in Example 1. Reactions were carried out in a
5 Perkin-Elmer Cetus ThermocyclerTM for 30 cycles comprising 1 minute at 95°C, 2 minutes at 55°C and 3 minutes at 72°C, with a final 7 minute extension at 72°C after the last cycle. Amplified DNAs were digested with the restriction enzymes NcoI and SmaI and fractionated on a 0.7% low melting point agarose gel in 40 mM Tris-acetate, pH8.5, 1 mM EDTA. The bands corresponding to the 3' and 5' fragments of the SBEI
10 cDNA were excised from the gel, melted at 68°C and each was combined with the 4.9 kb NcoI-SmaI fragment of plasmid pML103 (Figure 3) described in Example 1. Vector and insert DNAs were ligated at 15°C overnight, essentially as described in Maniatis. The chimeric gene of construct pBE72 (Figure 11) consists of a 2.49 kb SBEI fragment in antisense orientation with respect to the maize 27 kD zein promoter that is located 5'
15 to the SBEI fragment and the 10 kD zein 3' end that is located 3' to the SBEI fragment. The SBEI fragment of pBE72 (SEQ ID NO:19) was obtained by restriction enzyme digestion of pBE65 with EcoRI and HindIII followed by reaction with the Klenow fragment of *E. coli* DNA polymerase. The blunt-ended fragment was ligated to the Klenow-treated 4.9 kb NcoI-SmaI fragment of pML103 essentially as described in
20 Maniatis.

The ligated DNAs were used to transform *E. coli* XL1-Blue (Epicurean Coli XL-1 BlueTM; Stratagene). Bacterial transformants were initially screened by restriction enzyme digestion of plasmid DNA. For pBE68 and pBE69 transformants, the presence of the insert was detected by combined digestion with NcoI and SmaI. For pBE72
25 transformants, digestion of the DNA with SalI was used to confirm the presence of insert DNA and to determine the orientation of the SBEI fragment relative to the 27 kD zein promoter. Identified transformants were further characterized by limited nucleotide sequence analysis using the dideoxy chain termination method (SequenaseTM DNA Sequencing Kit; U. S. Biochemical).

30 The chimeric gene of pBE72 was subsequently introduced into the vector pKS17, described in Example 4. The 27 kD zein-SBEI-10 kD zein DNA fragment of pBE72 was liberated by partial digestion with BamHI and cloned into the BamHI site of pKS17 to give a hygromycin resistant equivalent of pBE72 termed pBE108 (Figure 12).
Transformation of Corn with the SBEI Antisense Constructs

35 In separate experiments, each SBEI antisense construct was introduced into embryogenic corn tissue by the particle bombardment method essentially as described in

Analysis of Starch from Transformed Corn Plants Containing the Near Full Length Antisense Construct

Starches from individual R1 kernels of plants transformed with pBE45 (the near full length antisense construct for corn SBEIIb) were analyzed using the procedure described in Example 1. As known to those skilled in the art, the antisense phenomenon is generally not observed in every individual transgenic line. Therefore, individual kernels from multiple lines were examined and as expected, some, but not all lines possessed kernels demonstrating an altered starch phenotype. Table 5 presents the results for kernels from a transformed corn line which did show a phenotype. The data represent the percentage difference of the various branches between kernels of the transformed line and kernels from a negative control (line 03376, which has been through the transformation process but does not contain the antisense gene).

Table 5. Percentage Difference of Branch Chain Distribution of Amylopectin from Starch Isolated from Individual Seed from Near Full Length Antisense SBEIIb Transgenic Corn Line (9228) Compared to Starch Isolated from Negative Control Line (03376).

<u>Starch Source</u>	<u>A + B1</u>	<u>B2</u>	<u>B3</u>	<u>B4+</u>
92283	92	97	81	192

As can be seen, long chains (B4+) are favored at the expense of shorter chains (A's and B1's, B2's and B3's) in the starch derived from the corn plant possessing the antisense gene relative to control starch (03376). The instant transgenic plant thus demonstrates a unique starch branching phenotype compared to non-transgenic control plants. This data indicates that alteration of corn starch branching enzyme activity by suppressing expression of the corresponding genes encoding starch branching enzymes results in an altered starch phenotype.

EXAMPLE 4

Preparation of Transgenic Corn Expressing a Near Full Length Sense Transcript of Corn Starch Branching Enzyme IIb

Preparation of the Expression Vector Encoding the Near Full Length Sense Construct

Plasmid pBE96 comprises a 2.09 kb fragment of the SBEIIb cDNA (SEQ ID NO:9) joined in the sense orientation to the 27 kD zein promoter and the 10 kD zein 3' end (Figure 8). The SBEIIb fragment commences at the initiating ATG codon of the coding region and terminates 312 bp 5' of the translation termination codon. pBE240 was subjected to site specific mutagenesis (Sculptor™ Mutagenesis Kit, Amersham) to generate an NcoI site at the ATG start site. The mutagenized plasmid was first digested with EcoRI and then rendered blunt-ended by reaction with Klenow. The DNA

fragment was liberated by digestion with NcoI, fractionated by electrophoresis on a low melting point agarose gel, and ligated to the NcoI-SmaI fragment of pML103 as described above. Transformants in *E. coli* XL1-Blue were tested for the presence of the SBEIIb fragment by restriction enzyme digestion with NcoI and HindIII followed by nucleotide sequence determination. From this analysis, pBE71 was identified. pBE71 was digested with PvuII to release the full chimeric gene (27 kD zein promoter-truncated SBEIIb-10 kD zein 3' end) and this fragment was cloned into the vector pKS17. pKS17 contains the hygromycin B phosphotransferase gene which confers resistance to the antibiotic hygromycin. pKS17 was assembled by the addition of a T7promoter -HPT-T7 terminator chimeric gene to a multicopy vector from which the b-lactamase gene had been deleted. The resultant plasmid containing the 27 kD zein-truncated SBEIIb-10 kD zein insert in pKS17 is termed pBE96.

EXAMPLE 5

Preparation of Transgenic Corn Expressing Antisense Transcripts of Corn Starch Branching Enzyme I

A corn SBEI DNA fragment was generated from the published sequence of the SBEI cDNA (Baba et al. (1991) *Biochem. Biophys. Res. Commun.* 181:87-94) by the polymerase chain reaction (PCR) using primers BE14 (SEQ ID NO:10) and BE15 (SEQ ID NO:11):

BE14 5' -AAGCTTGAATTCTGCTCGGTGATGAGACAC-3' (SEQ ID NO:10)

BE15 5' -AAGCTTGAATTCCTTGGAGGTGATGGCTAC-3' (SEQ ID NO:11)

BE14 and BE15 were combined with lambda DNA prepared from plate lysates of a 12 DAP corn cDNA library in lambda ZapII (Stratagene) in a standard PCR reaction mix consisting of 0.4 mM of each oligonucleotide and 0.8 mg of template DNA in 10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% w/v gelatin, 200 mM dGTP, 200 mM dATP, 200 mM dTTP, 200 mM dCTP and 0.025 unit Amplitaq™ DNA polymerase in a 100 ml volume. The 875 bp PCR fragment was digested with the restriction enzyme AccI to release a 325 bp fragment (encompassing nucleotides 2290-2610 of the published sequence) that was then used as a hybridization probe to screen the 12 DAP corn cDNA library for full length SBEI clones. One of the isolated clones, designated pBE65, contained a 2772 bp EcoRI insert (SEQ ID NO:12). Nucleotides 165 to 2772 of this clone were found to be more than 99 % identical to the sequence of the maize SBEI cDNA clone published by Baba et al. ((1991) *Biochem. Biophys. Res. Commun.* 181: 87-94). However, the 5' terminal 164 bp of the insert did not agree with the published sequence. To resolve this discrepancy, we attempted to

Example 1. Seven days after bombardment, the tissue was transferred to N6 medium that contained gluphosinate (2 mg per liter) and lacked casein or proline. The tissue continued to grow slowly on this medium. After an additional 2 weeks, the tissue was transferred to fresh N6 medium containing gluphosinate. After 6 weeks, areas of about 1
5 cm in diameter of actively growing callus were identified on some of the plates containing the gluphosinate supplemented medium. These calli continued to grow when sub-cultured on the selective medium.

Plants were regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks, the
10 tissue was transferred to regeneration medium (Fromm et al. (1990) *Bio/Technology* 8:833-839). Nine transgenic plant lines were regenerated from particle bombardment experiments performed with the DNA construct pBE68. 20 transgenic lines were regenerated from particle bombardments performed with the DNA construct pBE69 and 9 transgenic lines were regenerated from particle bombardment experiments performed
15 with the DNA construct pBE72.

Molecular Analysis of Transgenic Corn Plants Containing the SBEI Antisense Constructs

Total DNA was isolated from leaf tissue of transgenic plants essentially as described in Example 1. For Southern blot analysis of pBE68, pBE69 and pBE72 transformants, 10 mg of isolated DNA was digested with the restriction enzyme XbaI at
20 37°C for 6 hrs in the buffer supplied by the manufacturer. The restricted DNAs were electrophoresed at 40 volts overnight on a 0.8 % agarose gel in Tris-phosphate-EDTA buffer (Maniatis) and transferred to ImmobilonTM membranes. The blots were pre-hybridized, hybridized with nick translated pBE65 insert, and washed as described in Example 1.

25 Total RNA was isolated from developing (20-22 DAP) kernels of transgenic plants and Northern blots were prepared as described in Example 1. Blots were probed with nick translated pBE65 insert DNA and subsequently washed according to the regimen outlined in Example 1.

Of the 9 transgenic plant lines that were regenerated from particle bombardments with the pBE68 construct, 5 were identified by Southern blot analysis to contain the trait
30 gene. Northern blot analysis showed variable levels of the 2.7 kb SBEI mRNA in 4 of the Southern positive lines. In addition, 2 of these lines contained a 400 base transcript that presumably corresponds to the antisense RNA specified by the chimeric gene of pBE68. Of the 20 transgenic plant lines that were generated from bombardments with pBE69, 8 were found to contain pBE69 DNA. RNA isolated from two of the pBE69
35 transgenic plant lines showed the presence of the 600 base antisense transcript. Of the 9

available pBE72 transgenic plant lines, 6 were found by Southern blot analysis to be positive for the presence of the trait gene.

Analysis of Starch from Transformed Corn Plants Containing the 3' and 5' SBE1 Antisense Constructs

5 Starches from individual kernels of plants transformed with pBE68 (the 3' antisense construct for corn SBE1) and pBE69 (the 5' antisense construct for corn SBE1) were extracted using the procedure described in Example 1. As known to those skilled in the art, the antisense phenomenon is generally not observed in every individual transgenic line. Therefore, individual kernels from multiple lines were examined and as
10 expected, some, but not all lines possessed kernels demonstrating an altered phenotype. Starch digestion was modified from that in previous examples slightly as follows. 7.0 mg of each starch sample was added to a screw cap test tube with 1.1 mL of water. The tubes were heated to 120°C for 30 minutes and then placed in a water bath at 45°C. Debranching solution was made by diluting 50 µL of isoamylase (5×10^6 units/mL, Sigma) per mL of sodium acetate buffer (50 mM, pH 4.5). Forty µL of debranching
15 solution was added to each starch sample and incubated for 3 hours at 45°C. Reactions were stopped by heating to 110°C for 5 minutes. Debranched starch samples were lyophilized and redissolved in DMSO for analysis by gel permeation chromatography (GPC). One hundred µL of debranched starch was injected and run through 2 columns (Polymer Labs, Mixed Bed-C)) in series at 100°C and eluted with DMSO at a flow rate
20 of 1.0mL/min. Sampling interval was 25 minutes. A refractive index detector (Waters) was used with a computer running Chemstation Software (version A.02.05, Hewlett Packard) for detection and data collection and storage, respectively. Retention times of pullulan standards (380K, 100K, 23.7K, 5.8K, 728 and 180 mw) were used to define
25 molecular weight ranges for the debranched starch samples. The proportion of the total starch was determined for 24 ranges of degree of polymerization (DP) spanning both the amylose and amylopectin portions of the chromatogram. For purposes of comparison to data reported above the percentage area in appropriate DP ranges was summed to give values for A and B1 chains, B2, B3 and B4+ chains of the amylopectin portion of the
30 chromatogram. The proportion of the total area above DP 150 was used to determine amylose content.

 Starch was prepared from twelve individual R4 kernels from a line (XAY01414) positive for the pBE69 construct, debranched and analyzed as described above and compared to twelve individual kernels from untransformed corn. Tables 6 and 7 show
35 the average and standard error for line XAY01414 and the untransformed control.

Table 6. The Percentage of Total Chromatographic Area within Given Degree of Polymerization (DP) Ranges for Starch Derived from R4 Kernels Containing the 5' Antisense Transcript of Corn SBE I (XAY01414) and Dent Starch (control). Average of 12 individual seed and standard errors of the mean (SE) are provided.

5

<u>DP range</u>	<u>Dent Starch</u>		<u>XAY01414</u>	
	<u>Average</u>	<u>SE</u>	<u>Average</u>	<u>SE</u>
>5k	5.45	0.14	5.92	0.14
3-5k	2.62	0.05	2.58	0.04
1.8-3k	3.03	0.04	2.95	0.08
1.2-1.8k	2.49	0.05	2.66	0.03
0.9-1.2k	1.92	0.04	2.01	0.04
600-900	2.86	0.03	2.94	0.06
400-600	2.78	0.05	3.07	0.04
250-400	2.83	0.05	3.23	0.04
150-250	2.43	0.04	2.97	0.05
90-150	2.38	0.04	3.61	0.06
60-90	4.04	0.08	5.72	0.15
48-60	4.08	0.07	4.94	0.10
40-48	3.95	0.09	4.86	0.04
32-40	4.52	0.13	5.59	0.14
28-32	3.45	0.12	3.58	0.17
24-28	3.69	0.17	4.40	0.08
21-24	4.72	0.18	4.06	0.18
18-21	6.01	0.03	5.64	0.23
15-18	8.42	0.05	6.17	0.16
13-15	7.24	0.21	5.92	0.28
11-15	6.64	0.17	5.33	0.15
9-11	6.20	0.08	4.71	0.13
7-9	4.48	0.06	3.58	0.09
5-7	3.67	0.07	3.44	0.06

Table 7. Percentage Difference of Branch Chain Distribution of Amylopectin (expressed as A+B1, B2, B3 and B4+) and Amylose Content (% of Total Starch) from Starch Isolated from R4 Kernels Containing the 5' Antisense Transcript of Corn SBE I (XAY01414) as Compared to Control (Dent). DP range is indicated.

<u>A+B1 (5-15)</u>	<u>B2 (15-32)</u>	<u>B3 (32-60)</u>	<u>B4+ (60-150)</u>	<u>Amylose (>150)</u>
83.5	93.1	126.0	149.4	107.3

The transformant has alterations in both the amylose and amylopectin fractions of the starch. The overall amylose content is increased somewhat in the XAY01414 line. The amylopectin structure is also altered in that the longer chains (B3 and B4+) are increased relative to the dent control and the shorter chains are less abundant than in the dent starch.

Starch was prepared from twelve individual R4 kernels from a line (XAY00013) positive for the pBE68 construct and analyzed as described above. Tables 8 and 9 show the results of this analysis.

Table 8. The Percentage of Total Chromatographic Area within Given Degree of Polymerization (DP) Ranges for Starch Derived from R4 Kernels Containing the 3' Antisense Transcript of Corn SBE I (XAY00013) and Dent Starch (control). Average of 12 individual seed and standard errors of the mean (SE) are provided.

<u>DP range</u>	<u>Dent Starch</u>		<u>XAY00013</u>	
	<u>Average</u>	<u>SE</u>	<u>Average</u>	<u>SE</u>
>5k	5.45	0.14	6.13	0.39
3-5k	2.62	0.05	2.46	0.06
1.8-3k	3.03	0.04	2.92	0.05
1.2-1.8k	2.49	0.05	2.51	0.06
0.9-1.2k	1.92	0.04	2.02	0.04
600-900	2.86	0.03	2.93	0.05
400-600	2.78	0.05	3.02	0.06
250-400	2.83	0.05	3.19	0.05
150-250	2.43	0.04	2.83	0.06
90-150	2.38	0.04	3.15	0.07
60-90	4.04	0.08	5.33	0.10
48-60	4.08	0.07	4.77	0.13
40-48	3.95	0.09	4.73	0.16
32-40	4.52	0.13	5.62	0.18
28-32	3.45	0.12	3.99	0.16
24-28	3.69	0.17	3.97	0.19

45

21-24	4.72	0.18	4.67	0.18
18-21	6.01	0.03	5.40	0.12
15-18	8.42	0.05	6.64	0.16
13-15	7.24	0.21	5.73	0.22
11-15	6.64	0.17	5.23	0.11
9-11	6.20	0.08	5.27	0.10
7-9	4.48	0.06	4.08	0.09
5-7	3.67	0.07	3.31	0.10

Table 9. Percentage Difference of Branch Chain Distribution of Amylopectin (expressed as A+B1, B2, B3 and B4+) and Amylose Content (% of Total Starch) from Starch Isolated from R4 Kernels containing the 3' Antisense Transcript of Corn SBE I (XAY00013) as Compared to Control (Dent). DP range is indicated.

<u>A+B1 (5-15)</u>	<u>B2 (15-32)</u>	<u>B3 (32-60)</u>	<u>B4+ (60-150)</u>	<u>Amylose (>150)</u>
85.6	95.9	123.1	135.1	106.0

Like the XAY01414 line, the line transformed with the pBE68 construct has alterations in both the amylose and amylopectin fractions of the starch. Amylose content is increased relative to the control and longer chains (B4+ and B3) are increased in the amylopectin. The majority of the increase in amylose content is due to an increase in the Amylose of DP greater than 5000.

The instant transgenic plants thus demonstrate a unique starch branching pattern compared to the control plants. This data indicates that alteration of corn starch branching enzyme activity by suppressing expression of the corresponding genes encoding starch branching enzymes results in an altered starch phenotype.

EXAMPLE 6

Preparation of Transgenic Corn Expressing Sense Transcripts of Corn Starch Branching

Enzyme I

Preparation of the Expression Vector Encoding the Near Full Length Sense Construct

Plasmid pBE97 comprises a 1.87 kb fragment of the SBEI cDNA of pBE65 (SEQ ID NO:20) joined in the sense orientation to the 27 kD zein promoter and the 10 kD zein 3' end (Figure 13). The SBEI fragment encompasses nucleotides 55 through 1919 of the cDNA clone pBE65 and thus contains 117 bp of unknown sequence preceding the remaining 1748 bp of SBEI coding region DNA. This DNA fragment was generated by PCR-mediated site specific mutagenesis to introduce an NcoI site at nucleotide position 53 of the pBE65 sequence. The appropriate nucleotide primers were combined with pBE65 template DNA in a standard PCR reaction defined in Example 1.

The PCR fragment that was generated contains a ClaI site followed by an NcoI site and terminates at nucleotide 612 of the pBE65 sequence. This DNA fragment was digested with ClaI and PstI and exchanged with the corresponding region in pBE65 to give pBE79. pBE79 was digested with BstEII and rendered blunt-ended by reaction with the Klenow fragment of DNA polymerase (Maniatis). The DNA fragment was liberated by partial digestion with NcoI, fractionated by electrophoresis on a low melting point agarose gel, and ligated to the NcoI-SmaI fragment of pML103 described in Example 1. Transformants in *E. coli* XL-Blue were screened for the presence of the SBEI fragment by restriction enzyme digestion with NcoI and BamHI. From this analysis, pBE88 was identified. pBE88 was subjected to partial digestion with BamHI and the 3.87 kb fragment containing the 27 kD zein-truncated SBEI-10 kD zein chimeric gene was isolated by electrophoresis on a 0.7 % low melting point agarose gel (Maniatis). The DNA fragment was cloned into BamHI digested vector pKS17 described in Example 4. The resultant plasmid containing the 27 kD zein-truncated SBEI-10 kD zein insert in pKS17 is termed pBE97.

Two additional sense constructs of maize SBEI were made: pBE110 and pBE111. The full length and truncated sense fragments of these constructs were generated by removal of the artifactual 5' sequences of pBE65 and replacement with the correct 5' terminal sequences of the SBEI coding region. In order to generate a full length sense construct, the plasmid pBE79 described above was modified to incorporate a SmaI restriction site following nt 2674 of the insert sequence of pBE65. To accomplish this, a 805 bp 3' fragment of SBEI cDNA was obtained by PCR using the oligonucleotide pair BE15 (SEQ ID NO:11) and BE67 (SEQ ID NO:21):

BE15 5'-AAGCTTGAATTCCTTGGAGGTGATGGCTAC-3' (SEQ ID NO: 11)
 BE67 5'-CGCGGATCCCGGGTTCCAAGGGCGCCAGCGG-3' (SEQ ID NO: 21)

and pBE65 as the template DNA in a standard PCR reaction mixture as defined in Example 1. The PCR product was digested with the restriction enzymes BstEII and SmaI and the digestion product cloned into BstEII and SmaI digested pBE79 to give pBE83. The SBEI coding region fragment of pBE83 was subcloned into the vector pCC6 in two steps: first as an NcoI-SmaI fragment representing the 3' end and then as an NcoI fragment representing the 5' end of the coding region fragment. The vector pCC6 contains a 924 bp EcoRI-NcoI promoter fragment of the maize 10 kD zein gene followed by a 453 bp NcoI-SmaI fragment bearing the 10 kD zein coding region and a 944 bp 3' segment of the 10 kD zein gene in the cloning vector pTZ18R (Pharmacia). The pCC6 derivative which contains the NcoI-SmaI SBEI fragment is designated

pBE85. pBE85 was subjected to partial digestion with PvuII and the 4.7 kb 10 kD zein-SBEI-10 kD zein fragment was inserted into PvuII digested pKS17 (Example 4). The resultant construct designated pBE98, contains 110 bp of unidentified sequence at the 5' end of SBEI cDNA segment. The correct 5' sequence of the SBEI cDNA was obtained by PCR using oligonucleotides BE101 (SEQ ID NO:22) and BB3 (SEQ ID NO:23):

BE101 5' -AACTGCAGAAGGATCCCATGGTGTGCCTCGTGTGCCCC-3' (SEQ ID NO:22)

BB3 5' -GGATGCTTAAATGTGTACC-3' (SEQ ID NO:23)

10

and lambda DNA prepared from plate lysates of a 19 DAP corn endosperm cDNA library (Stratagene) as the template. The 748 bp PCR product was digested with NcoI and SstI to yield a 673 bp fragment. This DNA segment was exchanged with the corresponding region in pBE98 to give pBE110. The construct pBE110 is 7203 bp in length and consists of a 2565 bp segment of SBEI cDNA (SEQ ID NO:24) that includes the entire 823 amino acids of the SBEI coding region and 96 bp of 3' untranslated DNA (Figure 14). The SBEI DNA fragment is preceded by the promoter region of the maize 10 kD zein gene and is followed by the 3' end of the maize 10 kD zein gene.

The truncated sense SBEI construct pBE111 was generated by assembling a shortened SBEI coding region fragment in the vector pBC24. pBC24 is a pSK+ derivative in which the XbaI site has been blunted by reaction with the Klenow fragment of DNA polymerase and ligated to NcoI linkers. pBC24 thus lacks the XbaI site and contains a unique NcoI site in the polylinker region. The 5' SBEI fragment described above was digested with the restriction enzymes NcoI and BamHI and the 694 bp fragment was cloned into NcoI-BamHI digested pBC24. This intermediate was then digested with BamHI and SmaI and ligated to the 1874 bp BamHI-SmaI fragment of pBE83 to yield pBE112. pBE112 was digested with BstEII, reacted with Klenow and then subjected to partial digestion with NcoI. The liberated 1809 bp fragment was cloned into NcoI-partial SmaI digested pBT752. The vector, pBT752 is a derivative of pKS17 described in Example 4 which contains a 27 kD zein-maize high sulfur zein-10 kD zein chimeric gene and lacks the NcoI site at the translational start site of the hygromycin phosphotransferase gene. Analytical digests of the resultant transformants in NovaBlue (Novagen) cells revealed that the 10 kD zein 3' end was removed as a SmaI fragment during the cloning procedure. This 963 bp SmaI segment was thus isolated from pBT752 and inserted into a blunted HindIII site that is located just downstream from BstEII/SmaI junction in the intermediate plasmid, pBE110.5. Transformants were screened by digestion with DraI in order to determine the orientation of the 3' end fragment relative to the chimeric SBEI gene. From this analysis, pBE111 was identified.

pBE111 contains an 1809 bp fragment of the SBEI cDNA (SEQ ID NO:25) which is preceded by the 27 kD zein promoter and is followed by the 10 kD zein 3' end (Figure 15).

EXAMPLE 7

5 Use of Transgenic Corn Expressing Antisense Transcripts of Corn Starch Branching Enzyme IIb in Combination with the Waxy Mutant

A corn line carrying the 3' antisense transcript of corn starch branching enzyme IIb (pBE 44) was crossed with the well characterized corn starch mutant, waxy (wx). Individual segregants homozygous for the waxy mutation were identified in the
10 progeny of this cross. Kernels from line XAY00096 (homozygous wx) carrying the 3' antisense construct were selected. Starch was extracted from these kernels and subjected to Rapid Visco Analyzer pasting analysis as described in Example 1. Waxy (wx) and the homozygous double mutant, amylose extender waxy (ae wx), are shown for comparative purposes. A unique functionality was observed for line XAY00096 in
15 Figure 16. As can be seen from Figure 16, the pasting properties of the XAY00096 starch increased the pasting temperature as compared to waxy, but was lower than that of the homozygous ae wx. Viscosity was much higher than that of ae wx and was retained even after cooling, unlike wx which loses viscosity during pasting. This novel starch thus leads to unique pasting properties that are distinct than those observed in
20 waxy alone, in the SBEIIb null mutation (ae) in the combination of these two mutants (ae wx), or in transgenic line alone. The instant invention thus demonstrates the ability to produce starch with unique functionality by combining transgenic lines with known starch mutants.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: E. I. DU PONT DE NEMOURS AND COMPANY
 - (B) STREET: 1007 MARKET STREET
 - (C) CITY: WILMINGTON
 - (D) STATE: DELAWARE
 - (E) COUNTRY: UNITED STATES OF AMERICA
 - (F) POSTAL CODE (ZIP): 19898
 - (G) TELEPHONE: 302-992-4927
 - (H) TELEFAX: 302-773-0164
 - (I) TELEX: 6717325
- (ii) TITLE OF INVENTION: NOVEL STARCHES VIA MODIFICATION OF
EXPRESSION OF STARCH BIOSYNTHESIS
ENZYME GENES
- (iii) NUMBER OF SEQUENCES: 25
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: DISKETTE, 3.50 INCH
 - (B) COMPUTER: IBM PC COMPATIBLE
 - (C) OPERATING SYSTEM: WINDOWS 3.1
 - (D) SOFTWARE: MICROSOFT WORD 6.0A
- (v) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vi) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 06/009,113
 - (B) FILING DATE: DECEMBER 20, 1995
- (vii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: BRUCE W. MORRISSEY
 - (B) REGISTRATION NUMBER: 30,663
 - (C) REFERENCE/DOCKET NUMBER: BB-1066

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH 2665 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 79..2476

(xi) SEQUENCE DESCRIPTION SEQ ID NO:1:

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ACCCGGATTT CGCTCTTGCG GTCGCTGGGG TTTAGCATT GGCTGATCAG TTCGATCCGA      60
TCCGGCTGCG AAGGCGAG ATG GCG TTC CGG GTT TCT GGG GCG GTG CTC GGT      111
                Met Ala Phe Arg Val Ser Gly Ala Val Leu Gly
                1                               5                               10

GGG GCC GTA AGG GCT CCC CGA CTC ACC GGC GGC GGG GAG GGT AGT CTA      159
Gly Ala Val Arg Ala Pro Arg Leu Thr Gly Gly Gly Glu Gly Ser Leu
                15                               20                               25

GTC TTC CGG CAC ACC GGC CTC TTC TTA ACT CGG GGT GCT CGA GTT GGA      207
Val Phe Arg His Thr Gly Leu Phe Leu Thr Arg Gly Ala Arg Val Gly
                30                               35                               40

TGT TCG GGG ACG CAC GGG GCC ATG CGC GCG GCG GCC GCG GCC AGG AAG      255
Cys Ser Gly Thr His Gly Ala Met Arg Ala Ala Ala Ala Arg Lys
                45                               50                               55

GCG GTC ATG GTT CCT GAG GGC GAG AAT GAT GGC CTC GCA TCA AGG GCT      303
Ala Val Met Val Pro Glu Gly Glu Asn Asp Gly Leu Ala Ser Arg Ala
                60                               65                               70                               75

GAC TCG GCT CAA TTC CAG TCG GAT GAA CTG GAG GTA CCA GAC ATT TCT      351
Asp Ser Ala Gln Phe Gln Ser Asp Glu Leu Glu Val Pro Asp Ile Ser
                80                               85                               90

GAA GAG ACA ACG TGC GGT GCT GGT GTG GCT GAT GCT CAA GCC TTG AAC      399
Glu Glu Thr Thr Cys Gly Ala Gly Val Ala Asp Ala Gln Ala Leu Asn
                95                               100                               105

AGA GTT CGA GTG GTC CCC CCA CCA AGC GAT GGA CAA AAA ATA TTC CAG      447
Arg Val Arg Val Val Pro Pro Pro Ser Asp Gly Gln Lys Ile Phe Gln
                110                               115                               120

ATT GAC CCC ATG TTG CAA GGC TAT AAG TAC CAT CTT GAG TAT CGG TAC      495
Ile Asp Pro Met Leu Gln Gly Tyr Lys Tyr His Leu Glu Tyr Arg Tyr
                125                               130                               135

AGC CTC TAT AGA AGA ATC CGT TCA GAC ATT GAT GAA CAT GAA GGA GGC      543
Ser Leu Tyr Arg Arg Ile Arg Ser Asp Ile Asp Glu His Glu Gly Gly
                140                               145                               150                               155

TTG GAA GCC TTC TCC CGT AGT TAT GAG AAG TTT GGA TTT AAT GCC AGC      591
Leu Glu Ala Phe Ser Arg Ser Tyr Glu Lys Phe Gly Phe Asn Ala Ser
                160                               165                               170

GCG GAA GGT ATC ACA TAT CGA GAA TGG GCT CCT GGA GCA TTT TCT GCA      639
Ala Glu Gly Ile Thr Tyr Arg Glu Trp Ala Pro Gly Ala Phe Ser Ala
                175                               180                               185

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GCA	TTG	GTG	GGT	GAC	TTC	AAC	AAC	TGG	GAT	CCA	AAT	GCA	GAT	CGT	ATG	687
Ala	Leu	Val	Gly	Asp	Phe	Asn	Asn	Trp	Asp	Pro	Asn	Ala	Asp	Arg	Met	
		190					195					200				
AGC	AAA	AAT	GAG	TTT	GGT	GTT	TGG	GAA	ATT	TTT	CTG	CCT	AAC	AAT	GCA	735
Ser	Lys	Asn	Glu	Phe	Gly	Val	Trp	Glu	Ile	Phe	Leu	Pro	Asn	Asn	Ala	
	205					210					215					
GAT	GGT	ACA	TCA	CCT	ATT	CCT	CAT	GGA	TCT	CGT	GTA	AAG	GTG	AGA	ATG	783
Asp	Gly	Thr	Ser	Pro	Ile	Pro	His	Gly	Ser	Arg	Val	Lys	Val	Arg	Met	
220					225					230					235	
GAT	ACT	CCA	TCA	GGG	ATA	AAG	GAT	TCA	ATT	CCA	GCC	TGG	ATC	AAG	TAC	831
Asp	Thr	Pro	Ser	Gly	Ile	Lys	Asp	Ser	Ile	Pro	Ala	Trp	Ile	Lys	Tyr	
				240					245					250		
TCA	GTG	CAG	GCC	CCA	GGA	GAA	ATA	CCA	TAT	GAT	GGG	ATT	TAT	TAT	GAT	879
Ser	Val	Gln	Ala	Pro	Gly	Glu	Ile	Pro	Tyr	Asp	Gly	Ile	Tyr	Tyr	Asp	
			255					260					265			
CCT	CCT	GAA	GAG	GTA	AAG	TAT	GTG	TTC	AGG	CAT	GCG	CAA	CCT	AAA	CGA	927
Pro	Pro	Glu	Glu	Val	Lys	Tyr	Val	Phe	Arg	His	Ala	Gln	Pro	Lys	Arg	
		270					275					280				
CCA	AAA	TCA	TTG	CGG	ATA	TAT	GAA	ACA	CAT	GTC	GGA	ATG	AGT	AGC	CCG	975
Pro	Lys	Ser	Leu	Arg	Ile	Tyr	Glu	Thr	His	Val	Gly	Met	Ser	Ser	Pro	
	285					290					295					
GAA	CCG	AAG	ATA	AAC	ACA	TAT	GTA	AAC	TTT	AGG	GAT	GAA	GTC	CTC	CCA	1023
Glu	Pro	Lys	Ile	Asn	Thr	Tyr	Val	Asn	Phe	Arg	Asp	Glu	Val	Leu	Pro	
300					305					310					315	
AGA	ATA	AAA	AAA	CTT	GGA	TAC	AAT	GCA	GTG	CAA	ATA	ATG	GCA	ATC	CAA	1071
Arg	Ile	Lys	Lys	Leu	Gly	Tyr	Asn	Ala	Val	Gln	Ile	Met	Ala	Ile	Gln	
				320					325					330		
GAG	CAC	TCA	TAT	TAT	GGA	AGC	TTT	GGA	TAC	CAT	GTA	ACT	AAT	TTT	TTT	1119
Glu	His	Ser	Tyr	Tyr	Gly	Ser	Phe	Gly	Tyr	His	Val	Thr	Asn	Phe	Phe	
			335					340					345			
GCG	CCA	AGT	AGT	CGT	TTT	GGT	ACC	CCA	GAA	GAT	TTG	AAG	TCT	TTG	ATT	1167
Ala	Pro	Ser	Ser	Arg	Phe	Gly	Thr	Pro	Glu	Asp	Leu	Lys	Ser	Leu	Ile	
		350					355					360				
GAT	AGA	GCA	CAT	GAG	CTT	GGT	TTG	CTA	GTT	CTC	ATG	GAT	GTG	GTT	CAT	1215
Asp	Arg	Ala	His	Glu	Leu	Gly	Leu	Leu	Val	Leu	Met	Asp	Val	Val	His	
	365					370					375					
AGT	CAT	GCG	TCA	AGT	AAT	ACT	CTG	GAT	GGG	TTG	AAT	GGT	TTT	GAT	GGT	1263
Ser	His	Ala	Ser	Ser	Asn	Thr	Leu	Asp	Gly	Leu	Asn	Gly	Phe	Asp	Gly	
380					385					390					395	
ACA	GAT	ACA	CAT	TAC	TTT	CAC	AGT	GGT	CCA	CGT	GGC	CAT	CAC	TGG	ATG	1311
Thr	Asp	Thr	His	Tyr	Phe	His	Ser	Gly	Pro	Arg	Gly	His	His	Trp	Met	
				400					405					410		
TGG	GAT	TCT	CGC	CTA	TTT	AAC	TAT	GGG	AAC	TGG	GAA	GTT	TTA	AGA	TTT	1359
Trp	Asp	Ser	Arg	Leu	Phe	Asn	Tyr	Gly	Asn	Trp	Glu	Val	Leu	Arg	Phe	
			415					420					425			
CTT	CTC	TCC	AAT	GCT	AGA	TGG	TGG	CTC	GAG	GAA	TAT	AAG	TTT	GAT	GGT	1407
Leu	Leu	Ser	Asn	Ala	Arg	Trp	Trp	Leu	Glu	Glu	Tyr	Lys	Phe	Asp	Gly	
		430					435					440				

TTC	CGT	TTT	GAT	GGT	GTG	ACC	TCC	ATG	ATG	TAC	ACT	CAC	CAC	GGA	TTA	1455
Phe	Arg	Phe	Asp	Gly	Val	Thr	Ser	Met	Met	Tyr	Thr	His	His	Gly	Leu	
	445					450					455					
CAA	GTA	ACA	TTT	ACG	GGG	AAC	TTC	AAT	GAG	TAT	TTT	GGC	TTT	GCC	ACC	1503
Gln	Val	Thr	Phe	Thr	Gly	Asn	Phe	Asn	Glu	Tyr	Phe	Gly	Phe	Ala	Thr	
	460				465					470					475	
GAT	GTA	GAT	GCA	GTG	GTT	TAC	TTG	ATG	CTG	GTA	AAT	GAT	CTA	ATT	CAT	1551
Asp	Val	Asp	Ala	Val	Val	Tyr	Leu	Met	Leu	Val	Asn	Asp	Leu	Ile	His	
				480					485					490		
GGA	CTT	TAT	CCT	GAG	GCT	GTA	ACC	ATT	GGT	GAA	GAT	GTT	AGT	GGA	ATG	1599
Gly	Leu	Tyr	Pro	Glu	Ala	Val	Thr	Ile	Gly	Glu	Asp	Val	Ser	Gly	Met	
			495					500					505			
CCT	ACA	TTT	GCC	CTT	CCT	GTT	CAC	GAT	GGT	GGG	GTA	GGT	TTT	GAC	TAT	1647
Pro	Thr	Phe	Ala	Leu	Pro	Val	His	Asp	Gly	Gly	Val	Gly	Phe	Asp	Tyr	
		510					515					520				
CGG	ATG	CAT	ATG	GCT	GTG	GCT	GAC	AAA	TGG	ATT	GAC	CTT	CTC	AAG	CAA	1695
Arg	Met	His	Met	Ala	Val	Ala	Asp	Lys	Trp	Ile	Asp	Leu	Leu	Lys	Gln	
	525					530					535					
AGT	GAT	GAA	ACT	TGG	AAG	ATG	GGT	GAT	ATT	GTG	CAC	ACA	CTG	ACA	AAT	1743
Ser	Asp	Glu	Thr	Trp	Lys	Met	Gly	Asp	Ile	Val	His	Thr	Leu	Thr	Asn	
	540				545					550					555	
AGG	AGG	TGG	TTA	GAG	AAG	TGT	GTA	ACT	TAT	GCT	GAA	AGT	CAT	GAT	CAA	1791
Arg	Arg	Trp	Leu	Glu	Lys	Cys	Val	Thr	Tyr	Ala	Glu	Ser	His	Asp	Gln	
				560					565					570		
GCA	TTA	GTC	GGC	GAC	AAG	ACT	ATT	GCG	TTT	TGG	TTG	ATG	GAC	AAG	GAT	1839
Ala	Leu	Val	Gly	Asp	Lys	Thr	Ile	Ala	Phe	Trp	Leu	Met	Asp	Lys	Asp	
			575					580					585			
ATG	TAT	GAT	TTC	ATG	GCC	CTC	GAT	AGA	CCT	TCA	ACT	CCT	ACC	ATT	GAT	1887
Met	Tyr	Asp	Phe	Met	Ala	Leu	Asp	Arg	Pro	Ser	Thr	Pro	Thr	Ile	Asp	
		590					595					600				
CGT	GGG	ATA	GCA	TTA	CAT	AAG	ATG	ATT	AGA	CTT	ATC	ACA	ATG	GGT	TTA	1935
Arg	Gly	Ile	Ala	Leu	His	Lys	Met	Ile	Arg	Leu	Ile	Thr	Met	Gly	Leu	
	605					610					615					
GGA	GGA	GAG	GGC	TAT	CTT	AAT	TTC	ATG	GGA	AAT	GAG	TTT	GGA	CAT	CCT	1983
Gly	Gly	Glu	Gly	Tyr	Leu	Asn	Phe	Met	Gly	Asn	Glu	Phe	Gly	His	Pro	
	620				625					630					635	
GAA	TGG	ATA	GAT	TTT	CCA	AGA	GGT	CCG	CAA	AGA	CTT	CCA	AGT	GGT	AAG	2031
Glu	Trp	Ile	Asp	Phe	Pro	Arg	Gly	Pro	Gln	Arg	Leu	Pro	Ser	Gly	Lys	
				640					645					650		
TTT	ATT	CCA	GGG	AAT	AAC	AAC	AGT	TAT	GAC	AAA	TGT	CGT	CGA	AGA	TTT	2079
Phe	Ile	Pro	Gly	Asn	Asn	Asn	Ser	Tyr	Asp	Lys	Cys	Arg	Arg	Arg	Phe	
			655					660					665			
GAC	CTG	GGT	GAT	GCA	GAC	TAT	CTT	AGG	TAT	CAT	GGT	ATG	CAA	GAG	TTT	2127
Asp	Leu	Gly	Asp	Ala	Asp	Tyr	Leu	Arg	Tyr	His	Gly	Met	Gln	Glu	Phe	
		670					675					680				
GAT	CAG	GCA	ATG	CAA	CAT	CTT	GAG	CAA	AAA	TAT	GAA	TTC	ATG	ACA	TCT	2175
Asp	Gln	Ala	Met	Gln	His	Leu	Glu	Gln	Lys	Tyr	Glu	Phe	Met	Thr	Ser	
	685					690					695					

53

GAT CAC CAG TAT ATT TCC CGG AAA CAT GAG GAG GAT AAG GTG ATT GTG	2223
Asp His Gln Tyr Ile Ser Arg Lys His Glu Glu Asp Lys Val Ile Val	
700 705 710 715	
TTC GAA AAG GGA GAT TTG GTA TTT GTG TTC AAC TTC CAC TGC AAC AAC	2271
Phe Glu Lys Gly Asp Leu Val Phe Val Phe Asn Phe His Cys Asn Asn	
720 725 730	
AGC TAT TTT GAC TAC CGT ATT GGT TGT CGA AAG CCT GGG GTG TAT AAG	2319
Ser Tyr Phe Asp Tyr Arg Ile Gly Cys Arg Lys Pro Gly Val Tyr Lys	
735 740 745	
GTG GTC TTG GAC TCC GAC GCT GGA CTA TTT GGT GGA TTT AGC AGG ATC	2367
Val Val Leu Asp Ser Asp Ala Gly Leu Phe Gly Gly Phe Ser Arg Ile	
750 755 760	
CAT CAC GCA GCC GAG CAC TTC ACC GCC GAC TGT TCG CAT GAT AAT AGG	2415
His His Ala Ala Glu His Phe Thr Ala Asp Cys Ser His Asp Asn Arg	
765 770 775	
CCA TAT TCA TCC TCG GTT TAT ACA CCA AGC AGA ACA TGT GTC GTC TAT	2463
Pro Tyr Ser Ser Ser Val Tyr Thr Pro Ser Arg Thr Cys Val Val Tyr	
780 785 790 795	
GCT CCA GTG GAG T GATAGCGGGG TACTCGTTGC TCGCGGGCAT GTGTGGGGCT	2516
Ala Pro Val Glu	
GTGCATGTGA GGAAAAACCT TCTTCCAAAA CCGGCAGATG CATGCATGCA TGCTACAATA	2576
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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 414 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GACACCTTGA AGGTCTTATA TATATAGAGA GGACAACGCA GCGAGATGCA TGGGCTTTCC	60
AGCATCGATT AAAGTATCAG AACCTTATTG TAGCATGCAT GCATGCATCT GCCGGTTTTG	120
GAAGAAGGTT TTTCTCACA TCGACAGCCC CACACATGCC GCGCAGCAAC GAGTACCCCCG	180
CTATCACTCC ACTGGAGCAT AGACGACACA TGTTCTGCTT GGTGTATAAA CCGAGGATGA	240
ATATGGCCTA TTATCATGCG AACAGTCGGC GETGAAGTGC TCGGCTGCGT GATGGATCCT	300
GCTAAATCCA CCAAATAGTC CAGCGTCGGA GTCCAAGACC ACCTTATACA CCCCAGGCTT	360
TCGACAACCA ATACGGTAGT CAAAATAGCT GTTGTTGCAG TGGAAGTTGA ACAC	414

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid

54

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GAATTCCCGG GGTGTTCAAC TTCCACTGC

29

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GAATTCATG GGACACCTTG AAGGTCTT

28

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 507 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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CTGGAATATT TTTTGTCCAT CGCTTGGTGG GGGGACCACT CGAACTCTGT TCAAGGCTTG 120
AGCATCAGCC ACACCAGCAC CGCACGTTGT CTCTTCAGAA ATGTCTGGTA CCTCCAGTTC 180
ATCCGACTGG AATTGAGCCG AGTCAGCCCT TGATGCGAGG CCATCATTCT CGCCCTCAGG 240
AACCATGACC GCCTTCCTGG CCGCGGCCGC CGCGCGCATG GCCCCGTGCG TCCCCGAACA 300
TCCAACTCGA GCACCCCGAG TTAAGAAGAG GCCGGTGTGC CGGAAGACTA GACTACCCTC 360
CCCGCCGCCG GTGAGTCGGG GAGCCCTTAC GGCCCCACCG AGCACCGCCC CAGAAACCCG 420
GAACGCCATC TCGCCTTCGC AGCCGGATCG GATCGAACTG ATCAGCCAAT GCTAAAACCC 480
CAGCGACCGC AAGAGCGAAA TCCGGGT 507

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GAATTCCCGG GACCCGGATT TCGCTCTT

28

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GAATTCCATG GTCTATAGAG GCTGTACCG

29

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2165 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

AATTCATATT TTTGCTCAAG ATGTTGCATT GCCTGATCAA ACTCTTGCAT ACCATGATAC 60
 CTAAGATAGT CTGCATCACC CAGSTCAAAT CTTGACGAC ATTTGTCATA ACTGTTGTTA 120
 TTCCCTGGAA TAAACTTACC ACTTGGAAGT CTTTGGCGAC CTCTTGAAA ATCTATCCAT 180
 TCAGGATGTC CAAACTCATT TCCCATGAAA TTAAGATAGC CCTCTCCTCC TAAACCCATT 240
 GTGATAAGTC TAATCATCTT ATGTAATGCT ATCCCACGAT CAATGGTAGG AGTTGAAGGT 300
 CTATCGAGGG CCATGAAATC ATACATATCC TTGTCCATCA ACCAAAACGC AATAGTCTTG 360
 TCGCCGACTA ATGCTTGATC ATGACTTTCA GCATAAGTTA CACACTTCTC TAACCACCTC 420
 CTATTTGTCA GTGTGTGCAC AATATCACCC ATCTTCCAAG TTTCATCACT TTGCTTGAGA 480
 AGGTCAATCC ATTTGTCAGC CACAGCCATA TGCATCCGAT AGTCAAAACC TACCCACCA 540
 TCGTGAACAG GAAGGGCAAA TGTAGGCATT CCACTAACAT CTTACCAAT GGTTACAGCC 600
 TCAGGATAAA GTCCATGAAT TAGATCATTT ACCAGCATCA AGTAAACCAC TGCATCTACA 660
 TCGGTGGCAA AGCCAAAATA CTCATTGAAG TTCCCCGTAA ATGTTACTTG TAATCCGTGG 720
 TGAGTGTA CAATGGAGGT CACACCATCA AAACGGAAAC CATCAAACCTT ATATTCCTCG 780
 AGCCACCATC TAGCATTGGA GAGAAGAAAT CTTAAACCTT CCCAGTCCC ATAGTTAAAT 840
 AGGCGAGAAT CCCACATCCA GTGATGGCCA CGTGGACCAC TGTGAAAGTA ATGTGTATCT 900
 GTACCATCAA AACCATTCAA CCCATCCAGA GTATTACTTG ACGCATGACT ATGAACCACA 960
 TCCATGAGAA CTAGCAAACC AAGCTCATGT GCTCTATCAA TCAAAGACTT CAAATCTTCT 1020
 GGGGTACCAA AACGACTACT TGGCGCAAAA AAATTAGTTA CATGGTATCC AAAGCTTCCA 1080

TAATATGAGT GCTCTTGGAT TGCCATTATT TGCACATGCAT TGTATCCAAG TTTTTTTTATT 1140
 CTTGGGAGGA CTTTCATCCCT AAAGTTTACA TATGTGTTTA TCTTCGGTTC CGGGCTACTC 1200
 ATTCCGACAT GTGTTTCATA TATCCGCAAT GATTTTGGTC GTTTAGGTTG CGCATGCCTG 1260
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 TCCATTCTCA CCTTTACACG AGATCCATGA GGAATAGGTG ATGTACCATC TGCATTGTTA 1440
 GGCAGAAAAA TTTCCCAAAC ACCAACTCA TTTTGTCTCA TACGATCTGC ATTTGGATCC 1500
 CAGTTGTTGA CGTCACCCAC CAATGCTGCA GAAAATGCTC CAGGAGCCCA TTCTCGATAT 1560
 GTGATACCTT CCGCGCTGGC ATTAAATCCA AACTTCTCAT AACTACGGGA GAAGGCTTCC 1620
 AAGCCTCCTT CATGTTTCATC AATGTCTGAA CGGATTCTTC TATAGAGGCT GTACCGATAC 1680
 TCAAGATGGT ACTTATAGCC TTGCAACATG GGGTCAATCT GGAATATTTT TTGTCCATCG 1740
 CTTGGTGGGG GGACCACTCG AACTCTGTTC AAGGCTTGAG CATCAGCCAC ACCAGCACCG 1800
 CACGTTGTCT CTTTCAGAAAT GTCTGGTACC TCCAGTTCAT CCGACTGGAA TTGAGCCGAG 1860
 TCAGCCCTTG ATGCGAGGCC ATCATTCTCG CCCTCAGGAA CCATGACCGC CTTCTGGGCC 1920
 GCGGCCGCCG CGCGCATGGC CCCGTGCGTC CCCGAACATC CAACTCGAGC ACCCCGAGTT 1980
 AAGAAGAGGC CGGTGTGCCG GAAGACTAGA CTACCCTCCC CGCCGCCGGT GAGTCGGGGA 2040
 GCCCTTACGG CCCACCGAG CACCGCCCCA GAAACCCGGA ACGCCATCTC GCCTTCGCAG 2100
 CCGGATCGGA TCGAACTGAT CAGCCAATGC TAAACCCCA GCGACCGCAA GAGCGAAATC 2160
 CGGGT 2165

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 2087 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ATGGCGTTCC GGGTTTCTGG GGCGGTGCTC GGTGGGGCCG TAAGGGCTCC CCGACTCACC 60
 GGCGGCGGGG AGGGTAGTCT ACTCTTCCGG CACACCGGCC TCTTCTTAAC TCGGGGTGCT 120
 CGAGTTGGAT GTTCGGGGAC GCACGGGGCC ATGCGCGCGG CGGCCGCGGC CAGGAAGGCG 180
 GTCATGGTTC CTGAGGGCGA GAATGATGGC CTCGCATCAA GGGCTGACTC GGCTCAATTC 240
 CAGTCGGATG AACTGGAGGT ACCAGACATT TCTGAAGAGA CAACGTGCGG TGCTGGTGTG 300
 GCTGATGCTC AAGCCTTGAA CAGAGTTCGA GTGGTCCCCC CACCAAGCGA TGGACAAAAA 360
 ATATTCCAGA TTGACCCCAT GTTGCAAGGC TATAAGTACC ATCTTGAGTA TCGGTACAGC 420

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CTCTATAGAA GAATCCGTTT AGACATTGAT GAACATGAAG GAGGCTTGGG AGCCTTCTCC 480
CGTAGTTATG AGAAGTTTGG ATTTAATGCC AGCGCGGAAG GTATCACATA TCGAGAATGG 540
GCTCCTGGAG CATTTTCTGC AGCATTGGTG GGTGACTTCA ACAACTGGGA TCCAAATGCA 600
GATCGTATGA GCAAAAATGA GTTTGGTGTT TGGGAAATTT TTCTGCCTAA CAATGCAGAT 660
GGTACATCAC CTATTCCTCA TGGATCTCGT GTAAAGGTGA GAATGGATAC TCCATCAGGG 720
ATAAAGGATT CAATTCCAGC CTGGATCAAG TACTCAGTGC AGGCCCCAGG AGAAATACCA 780
TATGATGGGA TTTATTATGA TCCTCCTGAA GAGGTAAAGT ATGTGTTTCA GCATGCGCAA 840
CCTAAACGAC CAAAATCATT GCGGATATAT GAAACACATG TCGGAATGAG TAGCCCGGAA 900
CCGAAGATAA ACACATATGT AAACCTTAGG GATGAAGTCC TCCCAAGAAT AAAAAAATT 960
GGATACAATG CAGTGCAAAT AATGGCAATC CAAGAGCACT CATATTATGG AAGCTTTGGA 1020
TACCATGTAA CTAATTTTTT TGCGCCAAGT AGTCGTTTTG GTACCCCAAG AGATTTGAAG 1080
TCTTTGATTG ATAGAGCACA TGAGCTTGGT TTGCTAGTTC TCATGGATGT GGTTCATAGT 1140
CATGCGTCAA GTAATACTCT GGATGGGTTG AATGGTTTTG ATGGTACAGA TACACATTAC 1200
TTTCACAGTG GTCCACGTGG CCATCACTGG ATGTGGGATT CTCGCCTATT TAACTATGGG 1260
AACTGGGAAG TTTTAAGATT TCTTCTCTCC AATGCTAGAT GGTGGCTCGA GGAATATAAG 1320
TTTGATGGTT TCCGTTTTGA TGGTGTGACC TCCATGATGT AACTCACCA CGGATTACAA 1380
GTAACATTTA CGGGGAATT CAATGAGTAT TTTGGCTTTG CCACCGATGT AGATGCAGTG 1440
GTTTACTTGA TGCTGGTAAA TGATCTAATT CATGGACTTT ATCCTGAGGC TGTAAACATT 1500
GGTGAAGATG TTAGTGGAAAT GCCTACATTT GCCCTTCCTG TTCACGATGG TGGGGTAGGT 1560
TTTGAATATC GGATGCATAT GGCTGTGGCT GACAAATGGA TTGACCTTCT CAAGCAAAGT 1620
GATGAAACTT GGAAGATGGG TGATATTGTG CACACACTGA CAAATAGGAG GTGGTTAGAG 1680
AAGTGTGTAA CTTATGCTGA AAGTCATGAT CAAGCATTAG TCGGCGACAA GACTATTGCG 1740
TTTTGGTTGA TGGACAAGGA TATGTATGAT TTCATGGCCC TCGATAGACC TTCAACTCCT 1800
ACCATTGATC GTGGGATAGC ATTACATAAG ATGATTAGAC TTATCACAAT GGTTTATAGG 1860
GGAGAGGGCT ATCTTAATTT CATGGGAAAT GAGTTTGGAC ATCCTGAATG GATAGATTTT 1920
CCAAGAGGTC CGCAAAGACT TCCAAGTGGT AAGTTTATTC CAGGGAATAA CAACAGTTAT 1980
GACAAATGTC GTCGAAGATT TGACCTGGGT GATGCAGACT ATCTTAGGTA TCATGGTATG 2040
CAAGAGTTTG ATCAGGCAAT GCAACATCTT GAGCAAAAAT ATGAATT 2087

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(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

AAGCTTGAAT TCTGCTCGGT GATGAGACAC

30

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 30 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAGCTTGAAT TCCTTGGAGG TGATGGCTAC

30

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2772 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 49..2580

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

TGCTGATCGA GTGAGGGAAT TCAGCAGCAG CAGCAGCAGG TAGCATAG CAT AGA TAT 57
 His Arg Tyr
 1

GAC GGC GGC GGA GGT GGA GGC CGC CAA GGA CAT CGC CGA GGA GAA GGC 105
 Asp Gly Gly Gly Gly Gly Gly Gly Arg Gln Gly His Arg Arg Gly Glu Gly
 5 10 15

CGT CGT GCC GTT GCC ACC GTC GCC CGC CAA GCC GGC CGA CGA CGA CTC 153
 Arg Arg Ala Val Ala Thr Val Ala Arg Gln Ala Gly Arg Arg Arg Leu
 20 25 30 35

CAA GGC CAT CGT CGC TCT TGC TCG CAT GCT GAT CGG GCG GCA CCG CCG 201
 Gln Gly His Arg Arg Ser Cys Ser His Ala Asp Arg Ala Ala Pro Pro
 40 45 50

GGG ATC GCG GGT GGC GGC AAT GTG CGC CTG AGT GTG TTG TCT GTC CAG 249
 Gly Ile Ala Gly Gly Gly Asn Val Arg Leu Ser Val Leu Ser Val Gln
 55 60 65

TGC AAG GCT CGC CGG TCA GGG GTG CGG AAG GTC AAG AGC AAA TTC GCC 297
 Cys Lys Ala Arg Arg Ser Gly Val Arg Lys Val Lys Ser Lys Phe Ala
 70 75 80

ACT GCA GCT ACT GTG CAA GAA GAT AAA ACT ATG GCA ACT GCC AAA GGC 345
 Thr Ala Ala Thr Val Gln Glu Asp Lys Thr Met Ala Thr Ala Lys Gly
 85 90 95

GAT Asp 100	GTC Val	GAC Asp	CAT His	CTC Leu	CCC Pro 105	ATA Ile	TAC Tyr	GAC Asp	CTG Leu	GAC Asp 110	CCC Pro	AAG Lys	CTG Leu	GAG Glu	ATA Ile 115	393
TTC Phe	AAG Lys	GAC Asp	CAT His	TTC Phe 120	AGG Arg	TAC Tyr	CGG Arg	ATG Met	AAA Lys 125	AGA Arg	TTC Phe	CTA Leu	GAG Glu	CAG Gln 130	AAA Lys	441
GGA Gly	TCA Ser	ATT Ile	GAA Glu 135	GAA Glu	AAT Asn	GAG Glu	GGA Gly	AGT Ser 140	CTT Leu	GAA Glu	TCT Ser	TTT Phe	TCT Ser	AAA Lys	GGC Gly	489
TAT Tyr	TTG Leu	AAA Lys 150	TTT Phe	GGG Gly	ATT Ile	AAT Asn	ACA Thr 155	AAT Asn	GAG Glu	GAT Asp	GGA Gly	ACT Thr 160	GTA Val	TAT Tyr	CGT Arg	537
GAA Glu 165	TGG Trp	GCA Ala	CCT Pro	GCT Ala	GCG Ala	CAG Gln 170	GAG Glu	GCA Ala	GAG Glu	CTT Leu	ATT Ile 175	GGT Gly	GAC Asp	TTC Phe	AAT Asn	585
GAC Asp 180	TGG Trp	AAT Asn	GGT Gly	GCA Ala	AAC Asn	CAT His 185	AAG Lys	ATG Met	GAG Glu	AAG Lys 190	GAT Asp	AAA Lys	TTT Phe	GGT Gly	GTT Val 195	633
TGG Trp	TCG Ser	ATC Ile	AAA Lys	ATT Ile 200	GAC Asp	CAT His	GTC Val	AAA Lys	GGG Gly 205	AAA Lys	CCT Pro	GCC Ala	ATC Ile	CCT Pro	CAC His 210	681
AAT Asn	TCC Ser	AAG Lys	GTT Val 215	AAA Lys	TTT Phe	CGC Arg	TTT Phe	CTA Leu 220	CAT His	GGT Gly	GGA Gly	GTA Val	TGG Trp 225	GTT Val	GAT Asp	729
CGT Arg	ATT Ile	CCA Pro 230	GCA Ala	TTG Leu	ATT Ile	CGT Arg 235	TAT Tyr	GCG Ala	ACT Thr	GTT Val	GAT Asp	GCC Ala 240	TCT Ser	AAA Lys	TTT Phe	777
GGA Gly 245	GCT Ala	CCC Pro	TAT Tyr	GAT Asp	GGT Gly 250	GTT Val	CAT His	TGG Trp	GAT Asp	CCT Pro	CCT Pro	GCT Ala	TCT Ser	GAA Glu	AGG Arg	825
TAC Tyr 260	ACA Thr	TTT Phe	AAG Lys	CAT His	CCT Pro 265	CGG Arg	CCT Pro	TCA Ser	AAG Lys	CCT Pro 270	GCT Ala	GCT Ala	CCA Pro	CGT Arg	ATC Ile 275	873
TAT Tyr	GAA Glu	GCC Ala	CAT His	GTA Val 280	GGT Gly	ATG Met	AGT Ser	GGT Gly	GAA Glu	AAG Lys 285	CCA Pro	GCA Ala	GTA Val	AGC Ser	ACA Thr	921
TAT Tyr	AGG Arg	GAA Glu	TTT Phe 295	GCA Ala	GAC Asp	AAT Asn	GTG Val	TTG Leu 300	CCA Pro	CGC Arg	ATA Ile	CGA Arg	GCA Ala 305	AAT Asn	AAC Asn	969
TAC Tyr	AAC Asn	ACA Thr	GTT Val 310	CAG Gln	TTG Leu	ATG Met	GCA Ala 315	GTT Val	ATG Met	GAG Glu	CAT His	TCG Ser	TAC Tyr	TAT Tyr	GCT Ala	1017
TCT Ser	TTC Phe 325	GGG Gly	TAC Tyr	CAT His	GTG Val	ACA Thr 330	AAT Asn	TTC Phe	TTT Phe	GCG Ala	GTT Val 335	AGC Ser	AGC Ser	AGA Arg	TCA Ser	1065

60

GGC	ACA	CCA	GAG	GAC	CTC	AAA	TAT	CTT	GTT	GAT	AAG	GCA	CAC	AGT	TTG	1113
Gly	Thr	Pro	Glu	Asp	Leu	Lys	Tyr	Leu	Val	Asp	Lys	Ala	His	Ser	Leu	
340					345					350					355	
GGT	TTG	CGA	GTT	CTG	ATG	GAT	GTT	GTC	CAT	AGC	CAT	GCA	AGT	AAT	AAT	1161
Gly	Leu	Arg	Val	Leu	Met	Asp	Val	Val	His	Ser	His	Ala	Ser	Asn	Asn	
				360					365					370		
GTC	ACA	GAT	GGT	TTA	AAT	GGC	TAT	GAT	GTT	GGA	CAA	AGC	ACC	CAA	GAG	1209
Val	Thr	Asp	Gly	Leu	Asn	Gly	Tyr	Asp	Val	Gly	Gln	Ser	Thr	Gln	Glu	
			375					380					385			
TCC	TAT	TTT	CAT	GCG	GGA	GAT	AGA	GGT	TAT	CAT	AAA	CTT	TGG	GAT	AGT	1257
Ser	Tyr	Phe	His	Ala	Gly	Asp	Arg	Gly	Tyr	His	Lys	Leu	Trp	Asp	Ser	
		390					395					400				
CGG	CTG	TTC	AAC	TAT	GCT	AAC	TGG	GAG	GTA	TTA	AGG	TTT	CTT	CTT	TCT	1305
Arg	Leu	Phe	Asn	Tyr	Ala	Asn	Trp	Glu	Val	Leu	Arg	Phe	Leu	Leu	Ser	
	405					410					415					
AAC	CTG	AGA	TAT	TGG	TTG	GAT	GAA	TTC	ATG	TTT	GAT	GGC	TTC	CGA	TTT	1353
Asn	Leu	Arg	Tyr	Trp	Leu	Asp	Glu	Phe	Met	Phe	Asp	Gly	Phe	Arg	Phe	
420					425				430						435	
GAT	GGA	GTT	ACA	TCA	ATG	CTG	TAT	CAT	CAC	CAT	GGT	ATC	AAT	GTG	GGG	1401
Asp	Gly	Val	Thr	Ser	Met	Leu	Tyr	His	His	His	Gly	Ile	Asn	Val	Gly	
				440					445					450		
TTT	ACT	GGA	AAC	TAC	CAG	GAA	TAT	TTC	AGT	TTG	GAC	ACA	GCT	GTG	GAT	1449
Phe	Thr	Gly	Asn	Tyr	Gln	Glu	Tyr	Phe	Ser	Leu	Asp	Thr	Ala	Val	Asp	
			455					460					465			
GCA	GTT	GTT	TAC	ATG	ATG	CTT	GCA	AAC	CAT	TTA	ATG	CAC	AAA	CTC	TTG	1497
Ala	Val	Val	Tyr	Met	Met	Leu	Ala	Asn	His	Leu	Met	His	Lys	Leu	Leu	
		470					475					480				
CCA	GAA	GCA	ACT	GTT	GTT	GCT	GAA	GAT	GTT	TCA	GGC	ATG	CCG	GTC	CTT	1545
Pro	Glu	Ala	Thr	Val	Val	Ala	Glu	Asp	Val	Ser	Gly	Met	Pro	Val	Leu	
	485					490					495					
TGC	CGG	CCA	GTT	GAT	GAA	GGT	GGG	GTT	GGG	TTT	GAC	TAT	CGC	CTG	GCA	1593
Cys	Arg	Pro	Val	Asp	Glu	Gly	Gly	Val	Gly	Phe	Asp	Tyr	Arg	Leu	Ala	
500					505					510					515	
ATG	GCT	ATC	CCT	GAT	AGA	TGG	ATT	GAC	TAC	CTG	AAG	AAT	AAA	GAT	GAC	1641
Met	Ala	Ile	Pro	Asp	Arg	Trp	Ile	Asp	Tyr	Leu	Lys	Asn	Lys	Asp	Asp	
				520					525					530		
TCT	GAG	TGG	TCG	ATG	GGT	GAA	ATA	GCG	CAT	ACT	TTG	ACT	AAC	AGG	AGA	1689
Ser	Glu	Trp	Ser	Met	Gly	Glu	Ile	Ala	His	Thr	Leu	Thr	Asn	Arg	Arg	
			535					540					545			
TAT	ACT	GAA	AAA	TGC	ATC	GCA	TAT	GCT	GAG	AGC	CAT	GAT	CAG	TCT	ATT	1737
Tyr	Thr	Glu	Lys	Cys	Ile	Ala	Tyr	Ala	Glu	Ser	His	Asp	Gln	Ser	Ile	
		550					555					560				
GTT	GGC	GAC	AAA	ACT	ATT	GCA	TTT	CTC	CTG	ATG	GAC	AAG	GAA	ATG	TAC	1785
Val	Gly	Asp	Lys	Thr	Ile	Ala	Phe	Leu	Leu	Met	Asp	Lys	Glu	Met	Tyr	
	565					570					575					

ACT	GGC	ATG	TCA	GAC	TTG	CAG	CCT	GCT	TCA	CCT	ACA	ATT	GAT	CGA	GGG	1833
Thr	Gly	Met	Ser	Asp	Leu	Gln	Pro	Ala	Ser	Pro	Thr	Ile	Asp	Arg	Gly	
580					585					590					595	
ATT	GCA	CTC	CAA	AAG	ATG	ATT	CAC	TTC	ATC	ACA	ATG	GCC	CTT	GGA	GGT	1881
Ile	Ala	Leu	Gln	Lys	Met	Ile	His	Phe	Ile	Thr	Met	Ala	Leu	Gly	Gly	
				600					605					610		
GAT	GGC	TAC	TTG	AAT	TTT	ATG	GGA	AAT	GAG	TTT	GGT	CAC	CCA	GAA	TGG	1929
Asp	Gly	Tyr	Leu	Asn	Phe	Met	Gly	Asn	Glu	Phe	Gly	His	Pro	Glu	Trp	
			615					620					625			
ATT	GAC	TTT	CCA	AGA	GAA	GGG	AAC	AAC	TGG	AGC	TAT	GAT	AAA	TGC	AGA	1977
Ile	Asp	Phe	Pro	Arg	Glu	Gly	Asn	Asn	Trp	Ser	Tyr	Asp	Lys	Cys	Arg	
		630					635					640				
CGA	CAG	TGG	AGC	CTT	GTG	GAC	ACT	GAT	CAC	TTG	CGG	TAC	AAG	TAC	ATG	2025
Arg	Gln	Trp	Ser	Leu	Val	Asp	Thr	Asp	His	Leu	Arg	Tyr	Lys	Tyr	Met	
		645				650					655					
AAT	GCG	TTT	GAC	CAA	GCG	ATG	AAT	GCG	CTC	GAT	GAG	AGA	TTT	TCC	TTC	2073
Asn	Ala	Phe	Asp	Gln	Ala	Met	Asn	Ala	Leu	Asp	Glu	Arg	Phe	Ser	Phe	
660					665					670					675	
CTT	TCG	TCG	TCA	AAG	CAG	ATC	GTC	AGC	GAC	ATG	AAC	GAT	GAG	GAA	AAG	2121
Leu	Ser	Ser	Ser	Lys	Gln	Ile	Val	Ser	Asp	Met	Asn	Asp	Glu	Glu	Lys	
				680					685					690		
GTT	ATT	GTC	TTT	GAA	CGT	GGA	GAT	TTA	GTT	TTT	GTT	TTC	AAT	TTC	CAT	2169
Val	Ile	Val	Phe	Glu	Arg	Gly	Asp	Leu	Val	Phe	Val	Phe	Asn	Phe	His	
			695					700					705			
CCC	AAG	AAA	ACT	TAC	GAG	GGC	TAC	AAA	GTG	GGA	TGC	GAT	TTG	CCT	GGG	2217
Pro	Lys	Lys	Thr	Tyr	Glu	Gly	Tyr	Lys	Val	Gly	Cys	Asp	Leu	Pro	Gly	
		710					715					720				
AAA	TAC	AGA	GTA	GCC	CTG	GAC	TCT	GAT	GCT	CTG	GTC	TTC	GGT	GGA	CAT	2265
Lys	Tyr	Arg	Val	Ala	Leu	Asp	Ser	Asp	Ala	Leu	Val	Phe	Gly	Gly	His	
		725				730					735					
GGA	AGA	GTT	GGC	CAC	GAC	GTG	GAT	CAC	TTC	ACG	TCG	CCT	GAA	GGG	GTG	2313
Gly	Arg	Val	Gly	His	Asp	Val	Asp	His	Phe	Thr	Ser	Pro	Glu	Gly	Val	
740					745					750					755	
CCA	GGG	GTG	CCC	GAA	ACG	AAC	TTC	AAC	AAC	CGG	CCG	AAC	TCG	TTC	AAA	2361
Pro	Gly	Val	Pro	Glu	Thr	Asn	Phe	Asn	Asn	Arg	Pro	Asn	Ser	Phe	Lys	
				760					765					770		
GTC	CTT	TCT	CCG	CCC	CGC	ACC	TGT	GTG	GCT	TAT	TAC	CGT	GTA	GAC	GAA	2409
Val	Leu	Ser	Pro	Pro	Arg	Thr	Cys	Val	Ala	Tyr	Tyr	Arg	Val	Asp	Glu	
			775					780					785			
GCA	GGG	GCT	GGA	CGA	CGT	CTT	CAC	GCG	AAA	CGA	GAG	ACA	GGA	AAG	ACG	2457
Ala	Gly	Ala	Gly	Arg	Arg	Leu	His	Ala	Lys	Arg	Glu	Thr	Gly	Lys	Thr	
		790					795					800				
TCT	CCA	GCA	GAG	AGC	ATC	GAC	GTC	AAA	GCT	TCC	AGA	GCT	AGT	AGC	AAA	2505
Ser	Pro	Ala	Glu	Ser	Ile	Asp	Val	Lys	Ala	Ser	Arg	Ala	Ser	Ser	Lys	
		805				810					815					

GAA GAC AAG GAG GCA ACG GCT GGT GGC AAG AAG GGA TGG AAG TTT GCG 2553
 Glu Asp Lys Glu Ala Thr Ala Gly Gly Lys Lys Gly Trp Lys Phe Ala
 820 825 830 835

CGG CAG CCA TCC GAT CAA GAT ACC AAA TGAAGCCAGG AGTCCTTGGT 2600
 Arg Gln Pro Ser Asp Gln Asp Thr Lys
 840

GAGGACTGGA CTGGCTGCCG GCGCCCTGTT AGTAGTCCTG CTCTACTGGA CTAGCCGCCG 2660
 CTGGCGCCCT TGGAACGGTC CTTTCCTGTA GCTTGCAGGC GACTGGTGTC TCATCACC GA 2720
 GCAGGCAGGC ACTGCTTGTA TAGCTTTTCT AGAATAATAA TCAGGGATGG AT 2772

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 373 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CGGTGATGAG ACACCAGTCG CCTGCAAGCT ACAGGAAAGG ACCGTTCCAA GGGCGCCAGC 60
 GGC GGCTAGT CCAGTAGAGC AGGACTACTA ACAGGGCGCC GGCAGCCAGT CCAGTCCTCA 120
 CCAAGGACTC CTGGCTTCAT TTGGTATCTT GATCGGATGG CTGCCGCGCA AACTTCCATC 180
 CCTTCTTGCC ACCAGCCGTT GCCTCCTTGT CTTCTTTGCT ACTAGCTCTG GAAGCTTTGA 240
 CGTCGATGCT CTCTGCTGGA GACGTCTTTC CTGTCTCTCG TTTCGCGTGA AGACGTCGTC 300
 CAGCCCCCTGC TTCGTCTACA CGGTAATAAG CCACACAGGT GCGGGGCGGA GAAAGGACTT 360
 TGAACGAGTT CGG 373

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GAATTCCCGG GCCGAAGTCG TTCAAAG

27

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GAATTCCATG GCGGTGATGA GACACCAGTC

30

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 571 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CCATCTTATG GTTTCACCA TTCCAGTCAT TGAAGTCACC AATAAGCTCT GCCTCCTGCG 60
 CAGCAGGTGC CCATTCACGA TATACAGTTC CATCCTCATT TGTATTAATC CCAAATTTCA 120
 AATAGCCTTT AGAAAAAGAT TCAAGACCTT CCCTCATTTT CTTCAATTGA TCCTTTCTGC 180
 TCTAGGAATC TTTTCATCCG GTACCTGAAA TGGTCCTGA ATATCTCCAG CTGGGGGTCC 240
 AGGTCGTATA TGGGGAGATG GTCGACATCG CCTTTGGCAG TTGCCATAGT TTTATCTTCT 300
 TGCACAGTAG CTGCAGTGGC GAATTTGCTC TTGACCTTCC GCACCCCTGA CCGGCGAGCC 360
 TTGCACTGGA CAGACAACAC ACTCAGGCGC ACATTGCCGC CACCCGCGAT CCGGCGGGT 420
 GCCGCCCGAT CAGCATGCCA GCAAGAGCGA CGATGGCCTT GGAGTCGTCG TCGGCCGGCT 480
 TGGCGGGCGA CGGTGGCAAC GGCACGACGG CCTTCTCCTC GGCGATGTCC TTGGCGGCCT 540
 CCACCTCCGC CGCCGTCATA TCTATGCTAT G 571

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "PCR primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GAATTCCATG GCCATCTTAT GGTTTGCACC

30

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 31 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "PCR primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GAATTCCCGG GCATAGCATA GATATGACGG C

31

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2487. base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

AGCTTTGACG TCGATGCTCT CTGCTGGAGA CGTCTTTCCT GTCTCTCGTT TCGCGTGAAG 60
ACGTCGTCCA GCCCCTGCTT CGTCTACACG GTAATAAGCC ACACAGGTGC GGGGCGGAGA 120
AAGGACTTTG AACGAGTTCG GCCGGTTGTT GAAGTTCGTT TCGGGCACCC CTGGCACCCC 180
TTCAGGCGAC GTGAAGTGAT CCACGTCGTG GCCAACTCTT CCATGTCCAC CGAAGACCAG 240
AGCATCAGAG TCCAGGGCTA CTCTGTATTT CCCAGGCAAA TCGCATCCCA CTTTGTAGCC 300
CTCGTAAGTT TTCTTGGGAT GGAAATTGAA AACAAAACT AAATCTCCAC GTTCAAAGAC 360
AATAACCTTT TCCTCATCGT TCATGTCGCT GACGATCTGC TTTGACGACG AAAGGAAGGA 420
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TTCTCTTGGA AAGTCAATCC ATTCTGGGTG ACCAACTCA TTTCCCATAA AATTCAAGTA 600
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CAGGAGAAAT GCAATAGTTT TGTCGCCAAC AATAGACTGA TCATGGCTCT CAGCATATGC 780
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GCGATAGTCA AACCCAACCC CACCTTCATC AACTGGCCGG CAAAGGACCG GCATGCCTGA 960
AACATCTTCA GCAACAACAG TTGCTTCTGG CAAGAGTTTG TGCATTAAAT GGTGTTGCAAG 1020
CATCATGTAA ACAACTGCAT CCACAGCTGT GTCCAAACTG AAATATTCCT GGTAAGTTTCC 1080
AGTAAACCCC ACATTGATAC CATGGTGATG ATACAGCATT GATGTAATC CATCAAATCG 1140
GAAGCCATCA AACATGAATT CATCCAACCA ATATCTCAGG TTAGAAAGAA GAAACCTTAA 1200

TACCTCCCAG TTAGCATAGT TGAACAGCCG ACTATCCCAA AGTTTATGAT AACCTCTATC 1260
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 TGTGACATTA TTA CTGTCAT GGCTATGGAC AACATCCATC AGAACTCGCA AACCCAAACT 1380
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 CAACTGAACT GTGTTGTAGT TATTTGCTCG TATGCGTGGC AACACATTGT CTGCAAATTC 1560
 CCTATAITGT CTTACTGCTG GCTTTTCACC ACTCATACCT ACATGGGCTT CATAGATACG 1620
 TGGAGCAGCA GGCTTTGAAG GCCGAGGATG CTTAAATGTG TACCTTTCAG AAGCAGGAGG 1680
 ATCCCAATGA ACACCATCAT AGGGAGCTCC AAATTTAGAG GCATCAACAG TCGCATAACG 1740
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 GGAATTGTGA GGGATGGCAG GTTTCCTTT GACATGGTCA ATTTTGATCG ACCAAACACC 1860
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(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1865 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

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66

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TGTGTCTGT CCAGTGCAAG GCTCGCCGGT CAGGGGTGCG GAAGGTCAAG AGCAAATTCG 240
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```

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
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 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

67

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "PCR primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

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31

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
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 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "PCR primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

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38

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
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 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "PCR primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

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19

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2565 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

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AGTGTGTTGT CTGTCCAGTG CAAGGCTCGC CGGTCAGGGG TCGGGAAGGT CAAGAGCAAA 180

TTCGCCACTG CAGCTACTGT GCAAGAAGAT AAAACTATGG CAACTGCCAA AGGCGATGTC 240

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CCTGGGAAAT ACAGAGTAGC CCTGGACTCT GATGCTCTGG TCTTCGGTGG ACATGGAAGA 2160
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(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1809 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

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 AGTGTGTTGT CTGTCCAGTG CAAGGCTCGC CGGTCAGGGG TGCGGAAGGT CAAGAGCAAA 180
 TTCGCCACTG CAGCTACTGT GCAAGAAGAT AAAACTATGG CAACTGCCAA AGGCGATGTC 240
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CLAIMS

What is claimed is:

1. A method of controlling the starch fine structure of starch derived from the grain of corn comprising:

- 5 (a) preparing a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or a fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination,
- 10 (b) transforming corn with the chimeric gene of step (a),

wherein expression of said chimeric gene results in alteration of the fine structure of starch derived from the grain of said transformed corn compared to the fine structure of starch derived from corn not possessing said chimeric gene.

15

2. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the branch chain distribution of the amylopectin molecular component of said starch.

3. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the ratio of the amylose molecular component to the amylopectin molecular component of said starch.

20

4. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the degree of polymerization of the amylose molecular component of said starch.

5. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the branch chain distribution of the amylopectin molecular component and alteration of the ratio of the amylose molecular component to the amylopectin molecular component of said starch.

25

6. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the branch chain distribution of the amylopectin molecular component and alteration of the degree of polymerization of the amylose molecular component of said starch.

30

7. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the ratio of the amylose molecular component to the amylopectin molecular component and alteration of the degree of polymerization of the amylose molecular component of said starch.

35

8. The method of Claim 1 wherein said alteration of starch fine structure comprises alteration of the branch chain distribution of the amylopectin molecular

component, alteration of the ratio of the amylose molecular component to the amylopectin molecular component, and alteration of the degree of polymerization of the amylose molecular component of said starch.

5 9. The method of Claim 1 wherein the nucleic acid fragment encoding the starch branching enzyme structural gene or a fragment thereof is derived from corn.

10 10. The method of Claim 1 wherein the nucleic acid fragment encoding the starch branching enzyme structural gene or a fragment thereof encodes all or a portion of the corn SBEIIb enzyme.

10 11. The method of Claim 1 wherein wherein the nucleic acid fragment encoding the starch branching enzyme structural gene or a fragment thereof encodes all or a portion of the corn SBEI enzyme.

15 12. The method of Claim 1 wherein the nucleic acid fragment encoding the starch branching enzyme structural gene or a fragment thereof is operably linked in the antisense orientation relative to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue on the upstream side, and to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination on the downstream side.

20 13. The method of Claim 1 wherein the nucleic acid fragment encoding the starch branching enzyme structural gene or a fragment thereof is operably linked in the sense orientation relative to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue on the upstream side, and to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination on the downstream side.

25 14. A corn variety prepared by the method of Claim 1, or any progeny thereof

15. The corn variety of Claim 14 wherein the ratio of the amylose molecular component to the amylopectin molecular component of the starch isolated from the grain of said corn variety is increased compared to the ratio of the amylose molecular component to the amylopectin molecular component of starch isolated from the grain of untransformed corn.

30 16. The corn variety of Claim 14 wherein the amylopectin molecular component of the starch isolated from the grain of said corn variety comprises a greater proportion of longer α -1,4-linked glucan chains and a lesser proportion of shorter α -1,4-linked glucan chains compared to the amylopectin molecular component of starch isolated from the grain untransformed corn.

35 17. The corn variety of Claim 16 wherein the amylopectin component of the starch isolated from the grain of said corn variety has a greater proportion of B3 and

B4+ chains compared to the branch chain distribution of the amylopectin molecular component of starch isolated from the grain of untransformed corn.

18. Starch isolated from the grain of a corn variety prepared by the method of Claim 1 or any progeny thereof.

5 19. A method of preparing a thickened foodstuff comprising combining a foodstuff, water, and an effective amount of a starch of Claim 18 and cooking the resulting composition as necessary to produce said thickened foodstuff.

10 20. A corn variety transformed with a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination, or any progeny thereof.

15 21. A method of controlling the branch chain distribution of the amylopectin molecular component of starch in corn comprising:

(a) preparing a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or a fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination,

20 (b) transforming corn with the chimeric gene of step (a), wherein expression of said chimeric gene results in alteration of the branch chain distribution of the amylopectin molecular component of starch derived from the grain of said transformed corn compared to the branch chain distribution of the amylopectin molecular component of starch derived from corn not possessing said chimeric gene.

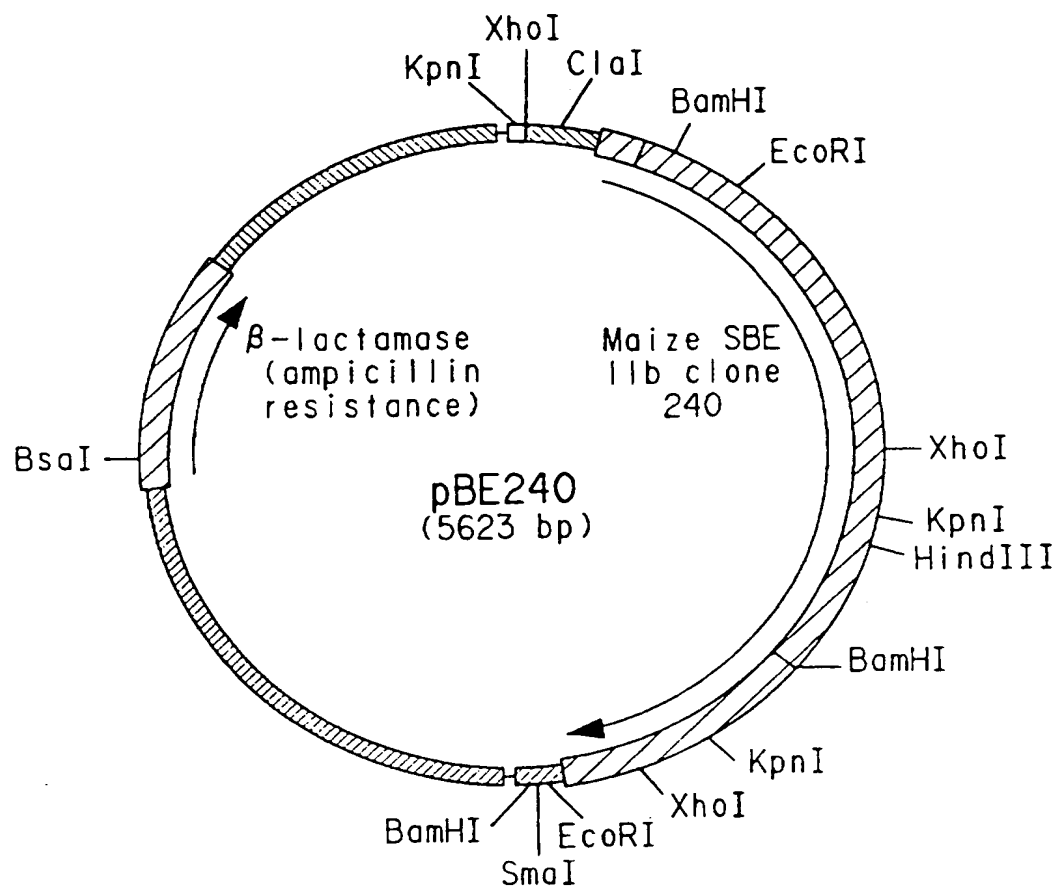


FIG. 1

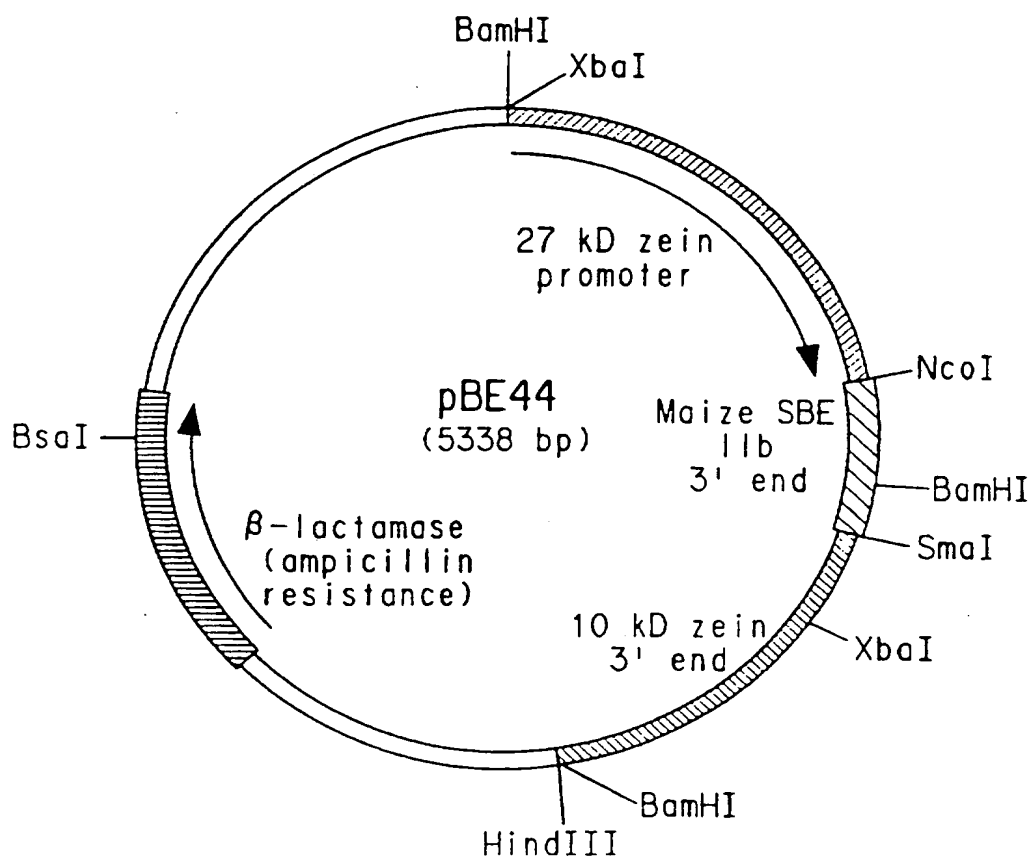


FIG.2

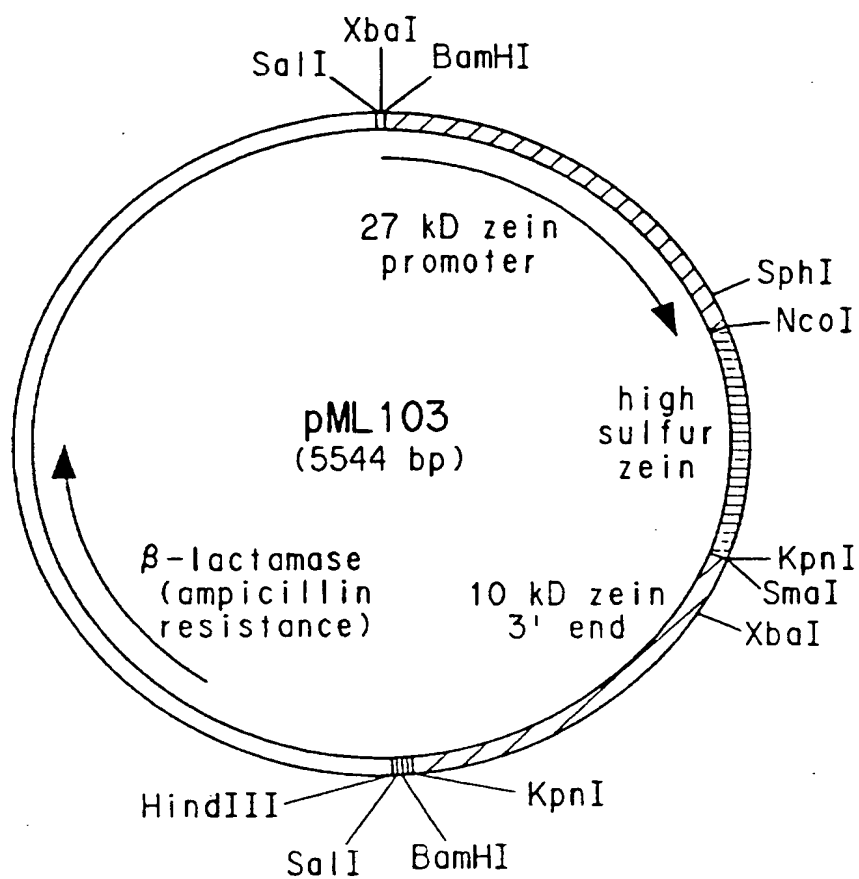


FIG. 3

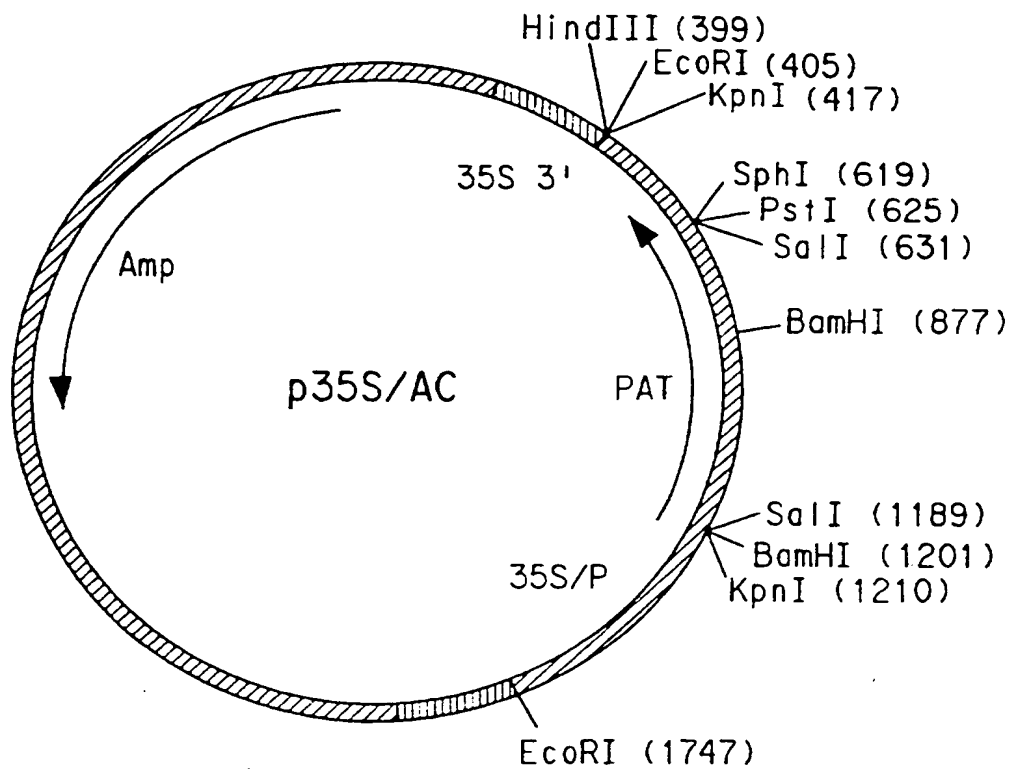
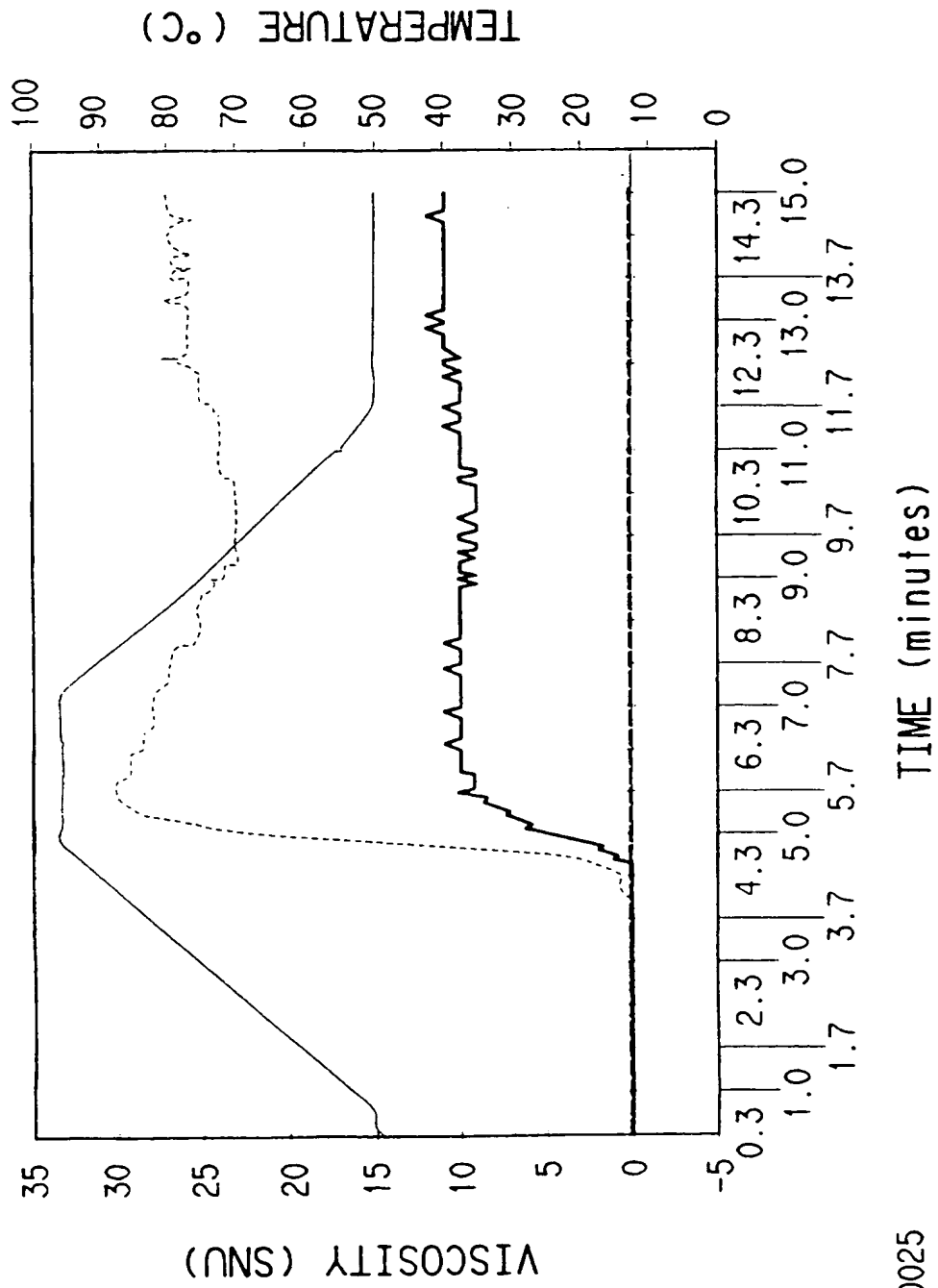


FIG.4

FIG. 5



----- dent
—— XAT00025
----- Ae
—— Temperature

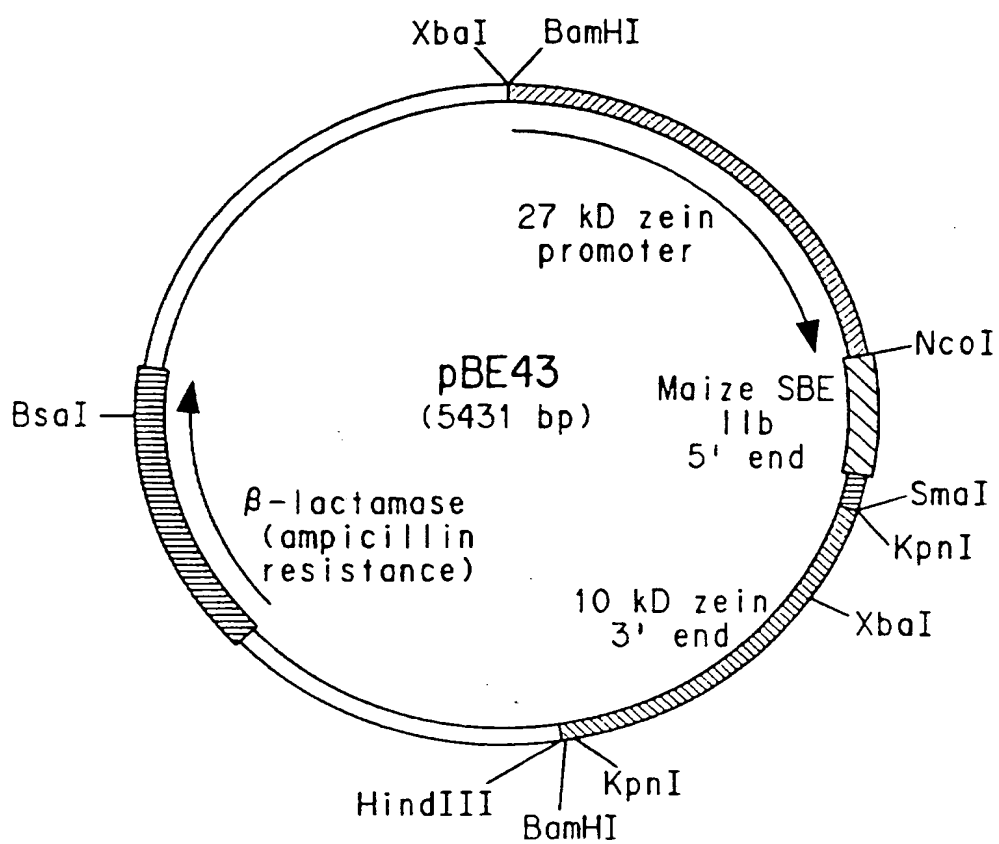


FIG. 6

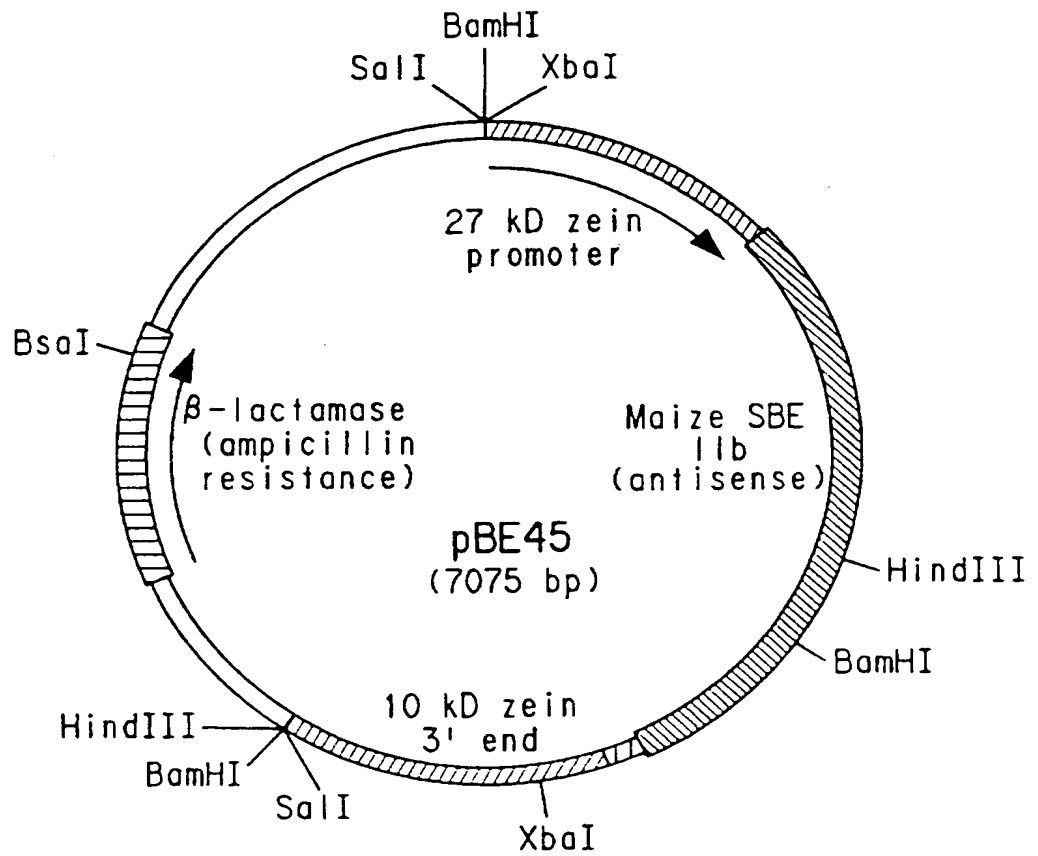


FIG. 7

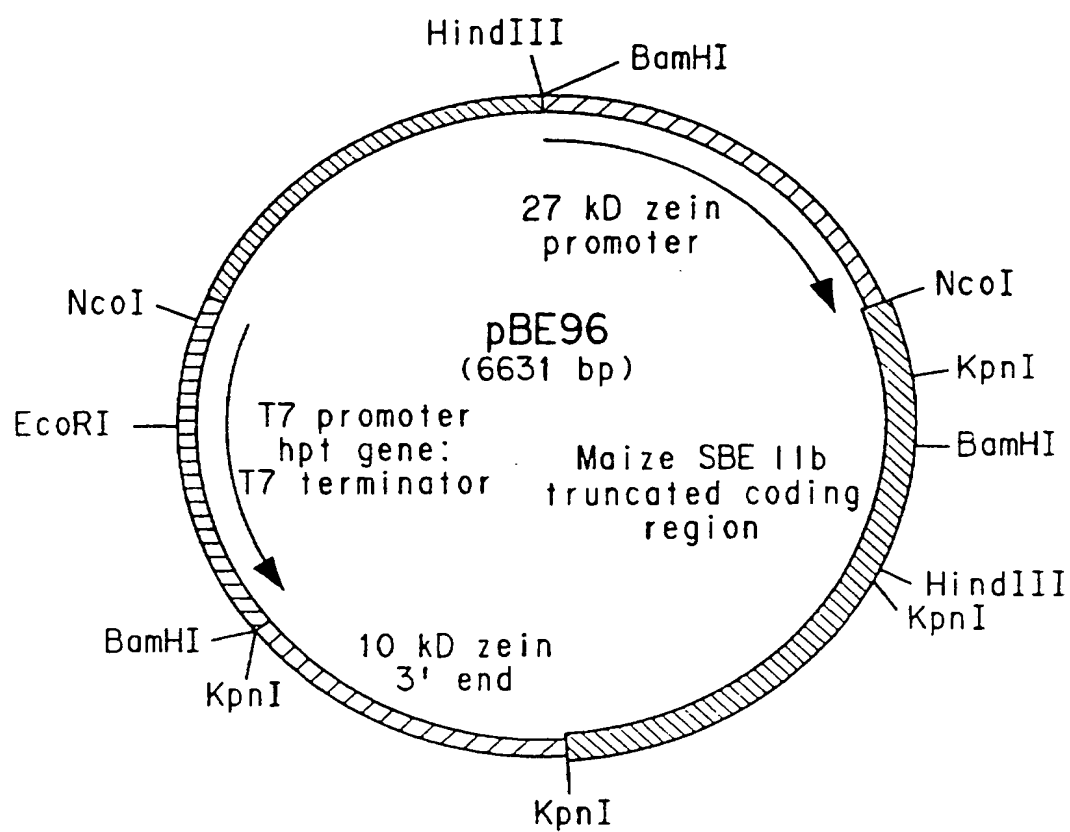


FIG. 8

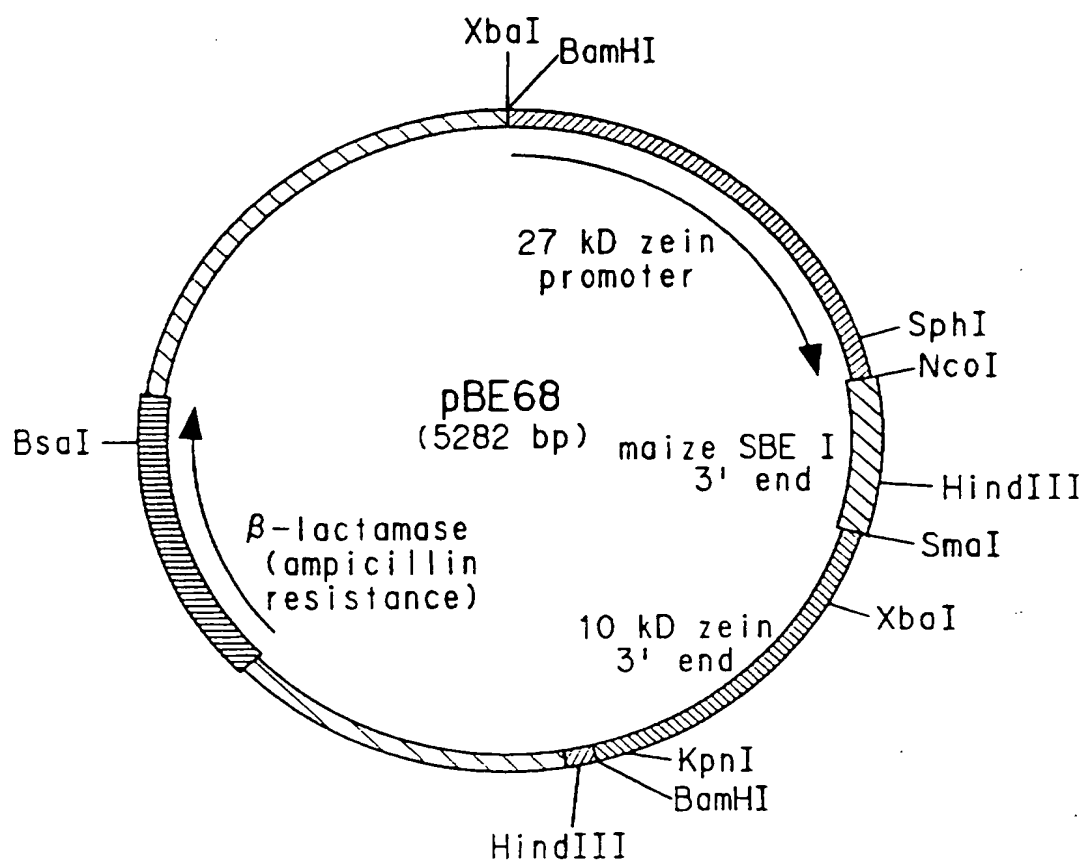


FIG. 9

10/16

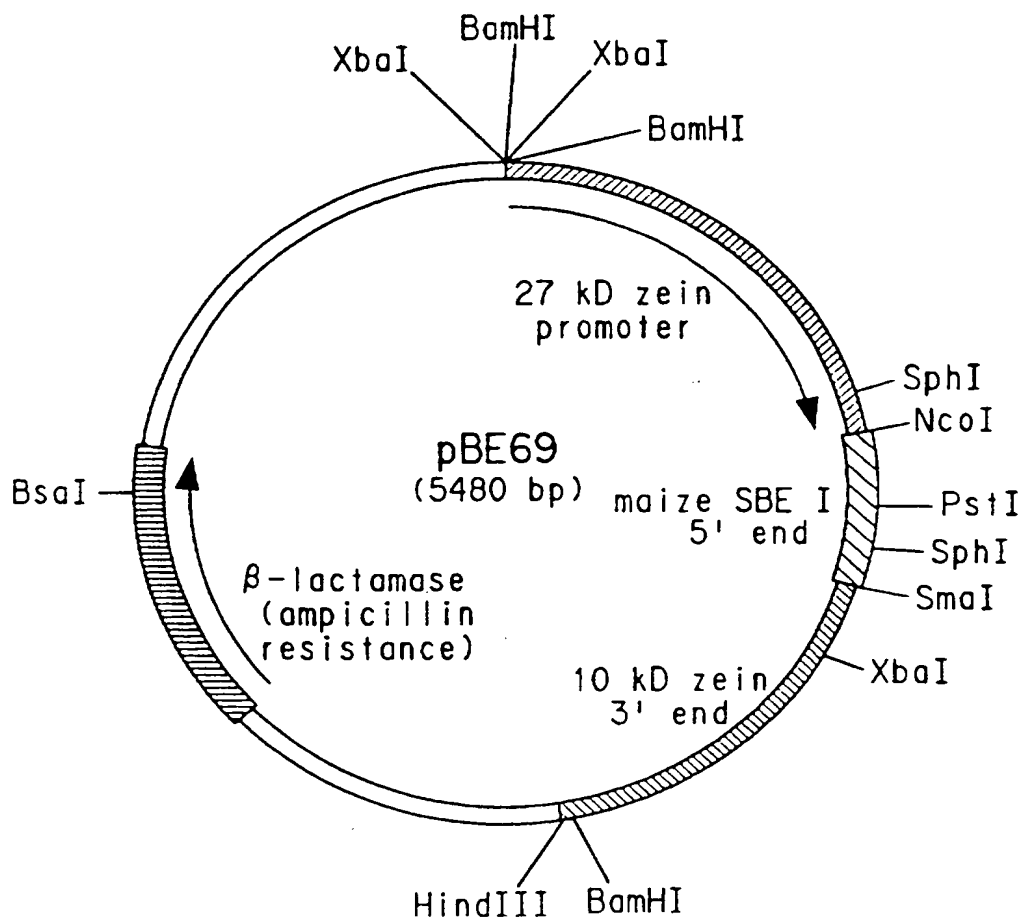


FIG. 10

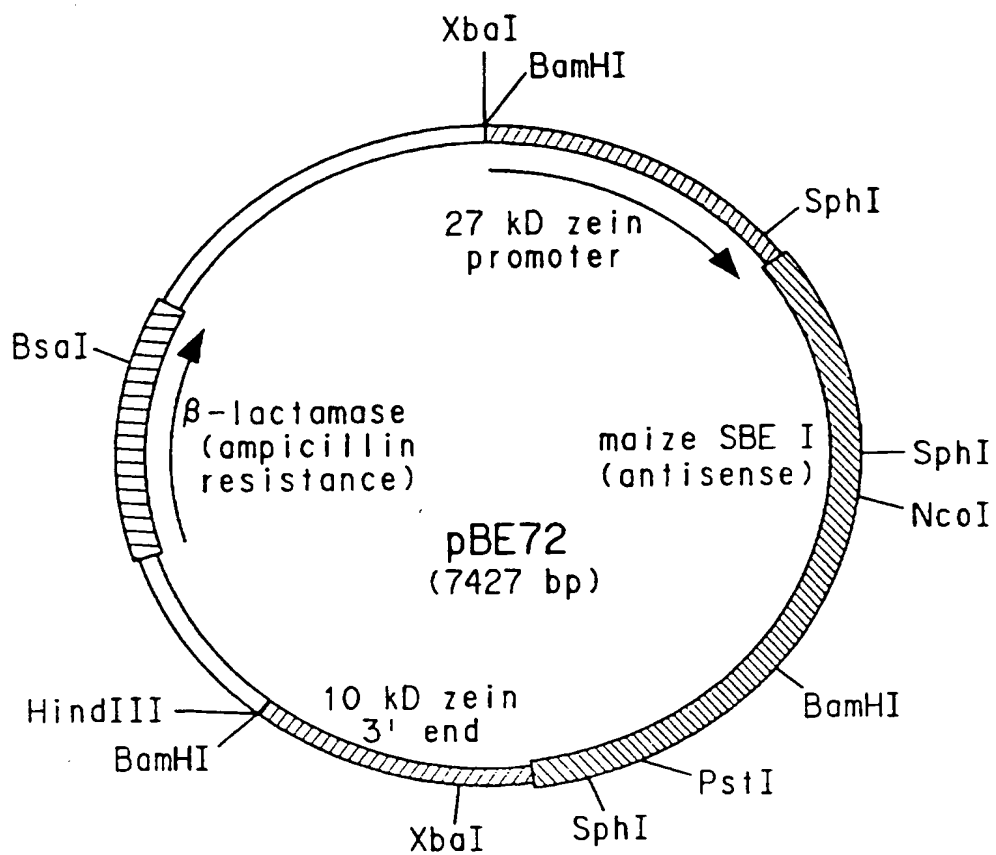


FIG. 11

12/16

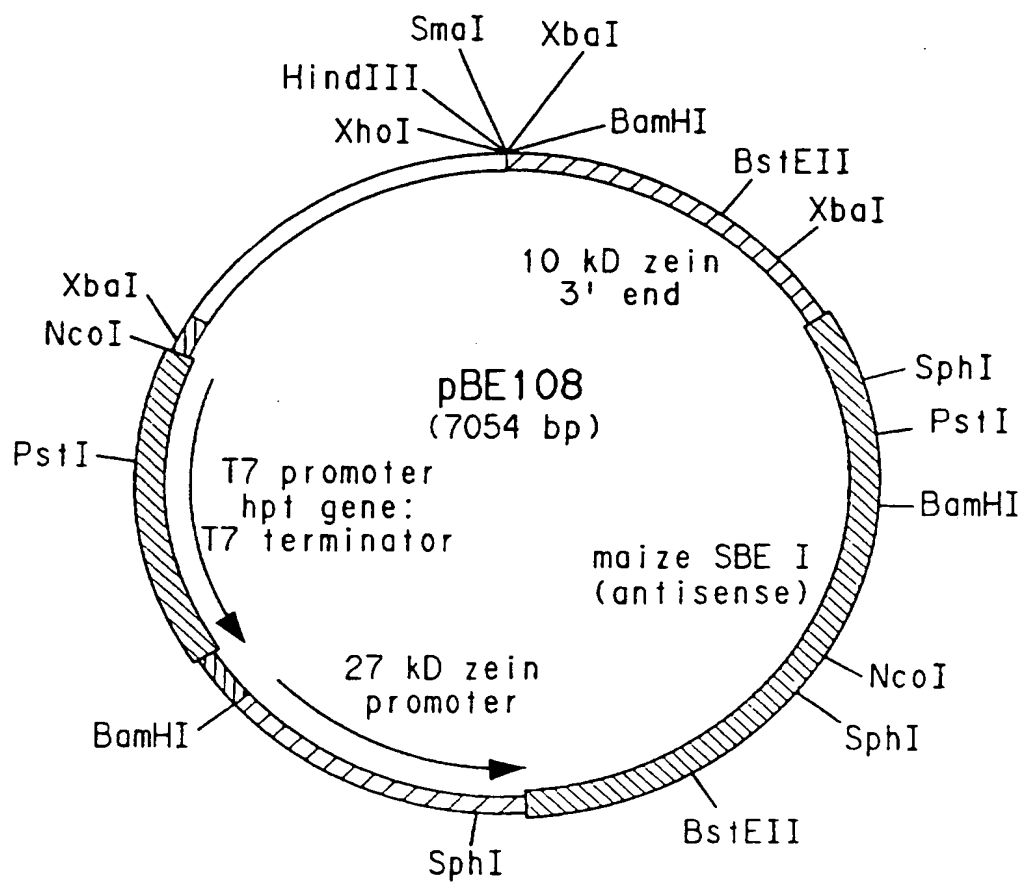


FIG. 12

13/16

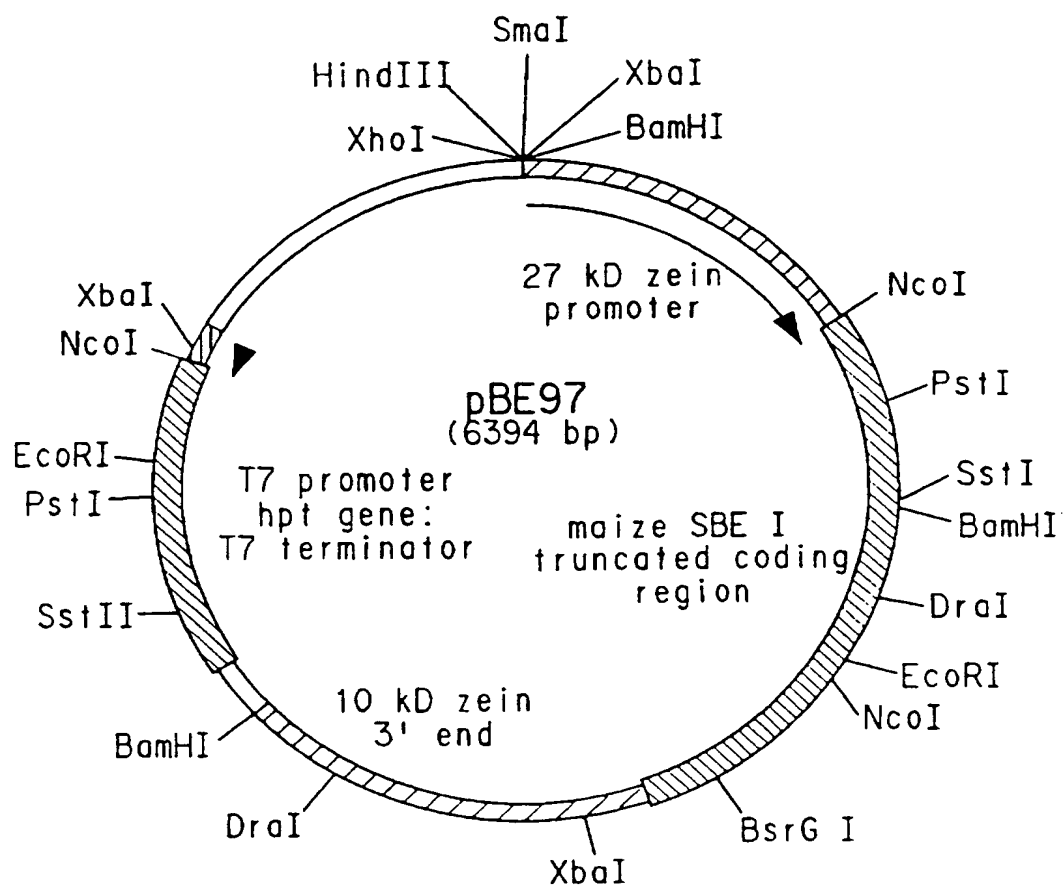


FIG. 13

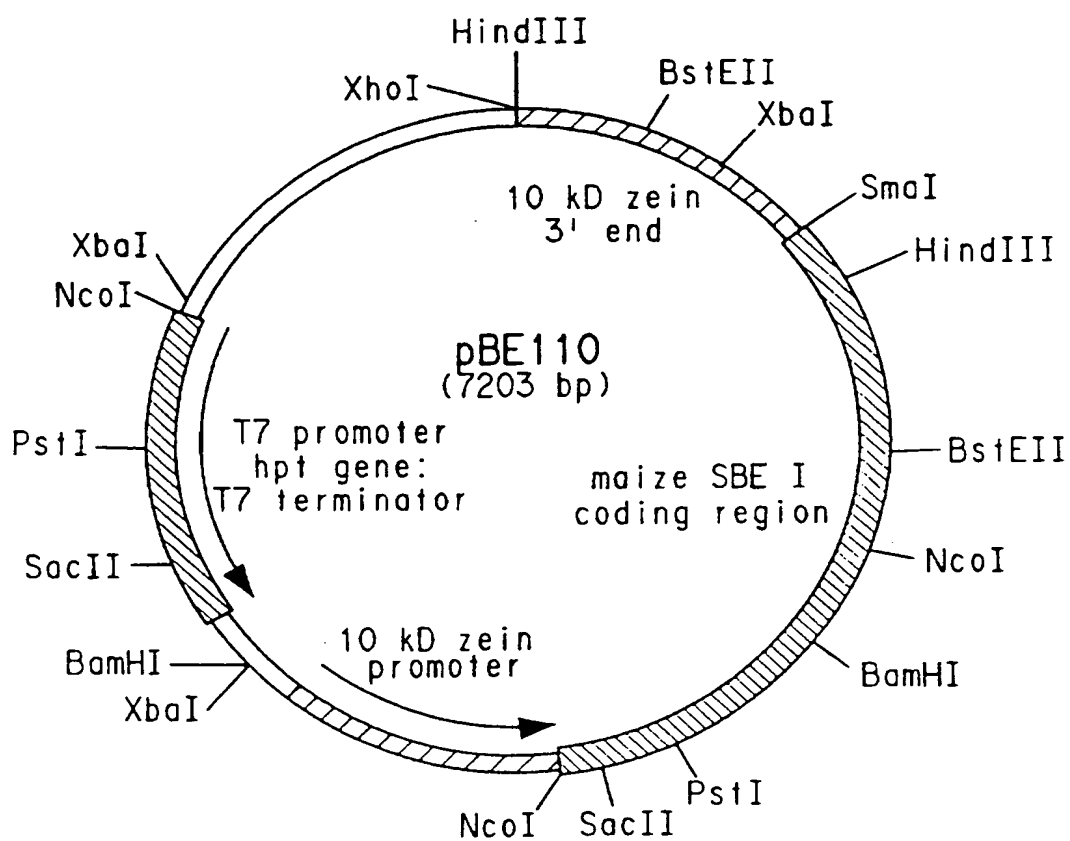


FIG. 14

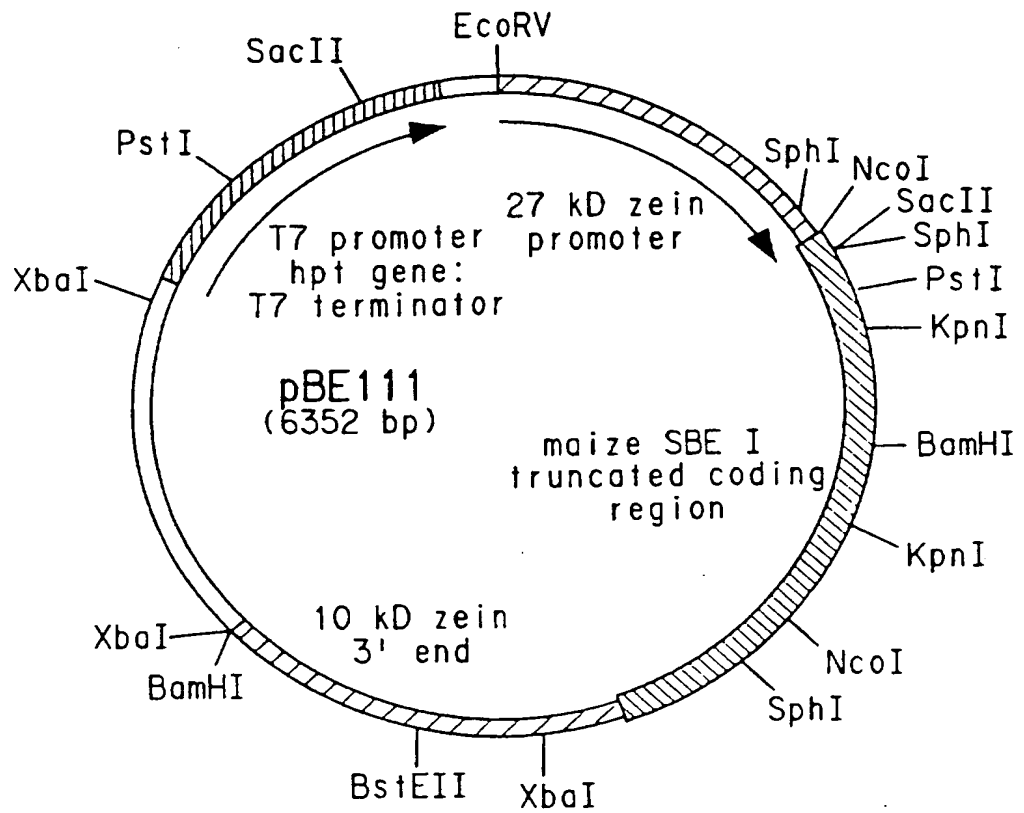
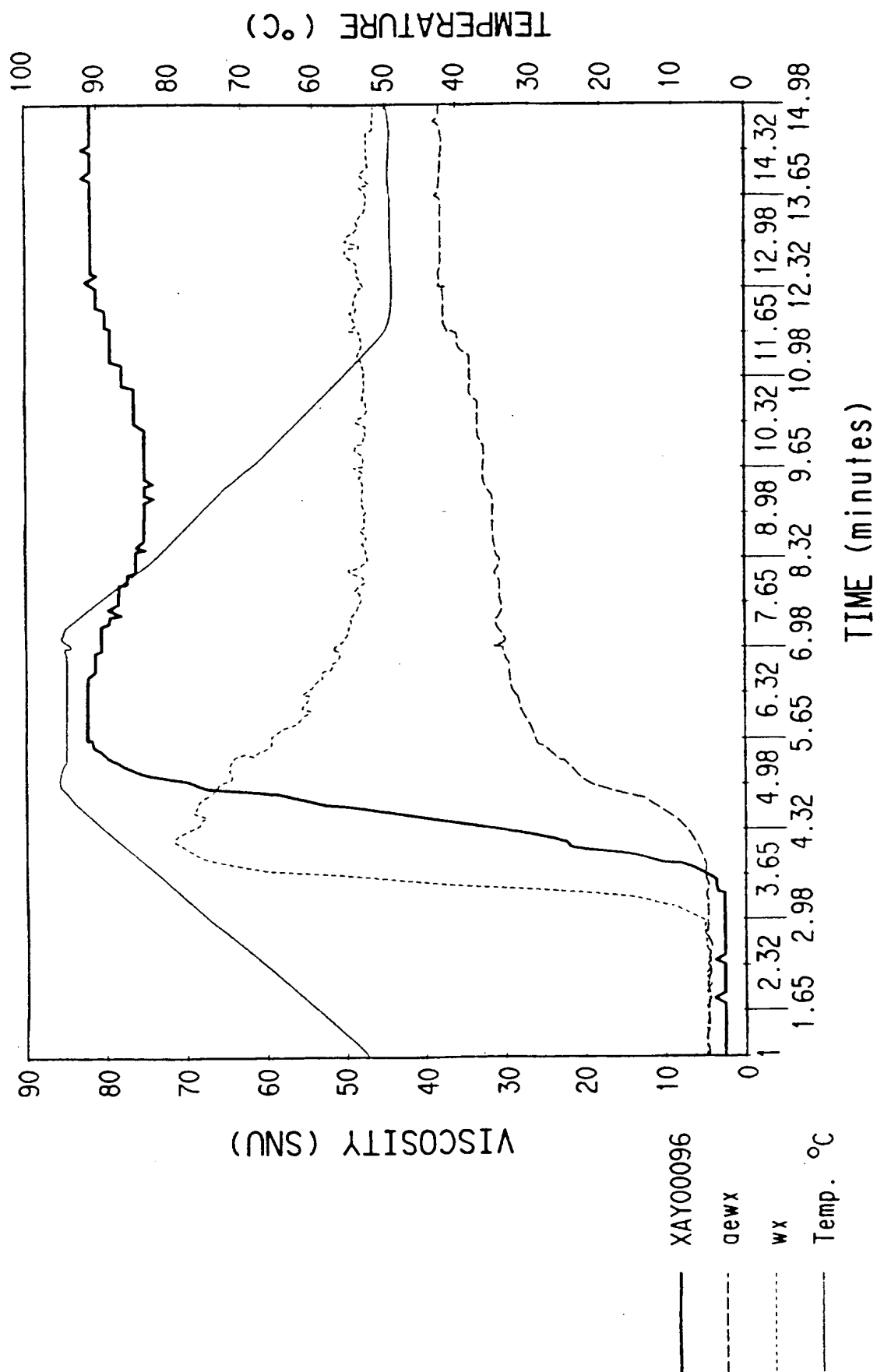


FIG. 15

FIG. 16



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PCT/US 96/19610
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A. The indications made below relate to the microorganism referred to in the description on page <u>23</u> , lines <u>14-16</u>	
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INTERNATIONAL SEARCH REPORT

Intern. Application No.
PCT/US 96/19678

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/54 C12N15/82 C12N9/10 C08B30/00 A01H5/00
A23L1/0522

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N C08B A01H A23L

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 94 11520 A (ZENECA LTD ;KEELING PETER LEWIS (GB)) 26 May 1994 see the whole document ---	1,9, 12-14, 18-20
X	WO 94 09144 A (ZENECA LTD) 28 April 1994 cited in the application	1,9,12, 13,18-20
Y	* see the whole document, esp. p.10,1.1-10, p.34,1.2-17, p.53,1.13-35 * --- -/--	2-8,10, 11, 14-17,21

☒ Further documents are listed in the continuation of box C.

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Date of the actual completion of the international search

28 May 1997

Date of mailing of the international search report

10.06.97

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PLANT PHYSIOLOGY, vol. 102, August 1993, pages 1269-1273, XP002031584 GUAN H. AND PREISS J.: "Differentiation of the properties of the branching isozymes from maize (Zea mays)" cited in the application see the whole document ---	2-8,10, 11, 14-17,21
A	BIOCHEMICAL GENETICS, vol. 20, no. 5/6, June 1982, pages 483-492, XP000674311 HEDMAN K. AND BOYER C.: "Gene dosage at the amylose-extender locus of maize: Effects on the levels of starch branching enzymes" see the whole document ---	1-21
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A	WO 92 14827 A (INST GENBIOLOGISCHE FORSCHUNG) 3 September 1992 * see the whole document, esp. p.2 1.6-11 * --- -/--	1-21



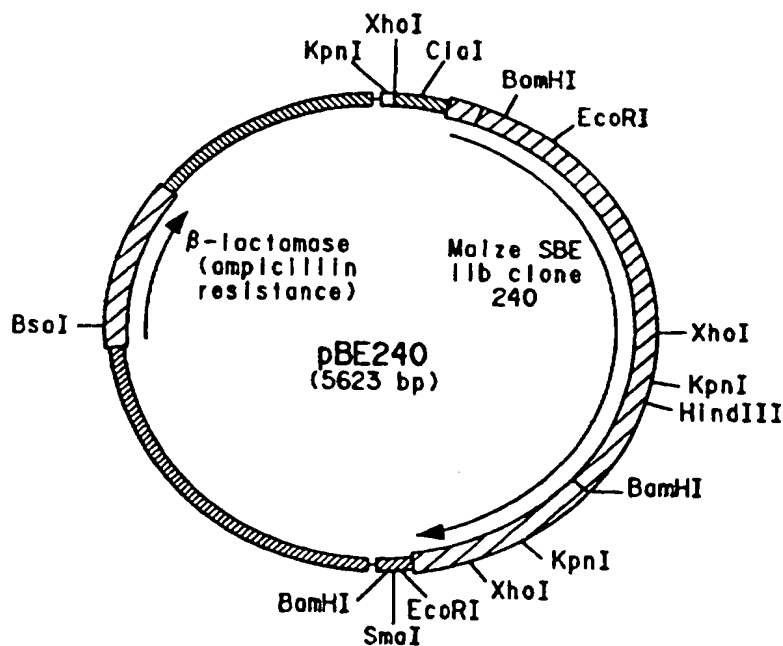
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(51) International Patent Classification ⁶ : C12N 15/54, 15/82, 9/10, C08B 30/00, A01H 5/00, A23L 1/0522		A3	(11) International Publication Number: WO 97/22703
			(43) International Publication Date: 26 June 1997 (26.06.97)
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(22) International Filing Date: 12 December 1996 (12.12.96)		Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> <i>With an indication in relation to a deposited microorganism furnished under Rule 13^{bis} separately from the description.</i> <i>Date of receipt by the International Bureau:</i> 25 February 1997 (25.02.97)	
(30) Priority Data: 60/009,113 20 December 1995 (20.12.95) US			
(71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US).			
(72) Inventors; and (75) Inventors/Applicants (for US only): HUBBARD, Natalie, Louise [US/US]; 2204 Mill Lane, Wilmington, DE 19810-4028 (US). KLEIN, Theodore, Mitchell [US/US]; 2229 Rosewood Drive, Wilmington, DE 19810-2815 (US). BROGLIE, Karen, E. [US/US]; 520 Port Royal Court, Landenberg, PA 19350-1333 (US).			
(74) Agent: MORRISSEY, Bruce, W.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US).		(88) Date of publication of the international search report: 24 July 1997 (24.07.97)	

(54) Title: NOVEL STARCHES VIA MODIFICATION OF EXPRESSION OF STARCH BIOSYNTHETIC ENZYME GENES

(57) Abstract

The instant invention discloses utilization of a cDNA clone to construct sense and antisense genes for inhibition of starch branching enzyme enzymatic activity in com. More specifically, this invention concerns a method of controlling the starch fine structure of starch derived from the grain of corn comprising: (1) preparing a chimeric gene comprising a nucleic acid fragment encoding a starch branching enzyme structural gene or a fragment thereof, operably linked in either sense or antisense orientation on the upstream side to a nucleic acid fragment encoding a promoter that directs gene expression in corn endosperm tissue, and operably linked on the downstream side to a nucleic acid fragment encoding a suitable regulatory sequence for transcriptional termination, and (2) transforming com with said chimeric gene, wherein expression of said chimeric gene results in alteration of the fine structure of starch derived from the grain of said transformed com compared to the fine structure of starch derived from com not possessing said chimeric gene.



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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PLANT PHYSIOLOGY, vol. 104, 1994, pages 1449-1453, XP002031588 GUAN H. ET AL.: "Expression of branching enzyme I of maize endosperm in Escherichia coli" * see the whole document, esp. p.1452, last chapter *</p> <p style="text-align: center;">---</p>	1-21
A	<p>P.N.A.S., U.S.A., vol. 92, no. 4, 14 February 1995, pages 964-967, XP002031589 GUAN H. ET AL.: "Maize branching enzyme catalyzes synthesis of glycogen-like polysaccharide in glgB-deficient Escherichia coli" see the whole document</p> <p style="text-align: center;">-----</p>	1-21

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 96/19678

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